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Antiviral-resistant Hepatitis B Virus: Can we prevent this monster from growing?

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Introduction

Despite the development of new nucleoside analogs that effectively inhibits hepatitis B virus (HBV) replication, antiviral therapy of chronic hepatitis B remains a clinical challenge mainly because of the slow kinetics of viral clearance and the subsequent emergence of drug resistant mutants (1). In this manuscript, we review the clinical impact of drug resistance, the new concepts of viral resistance management based on genotypic and phenotypic analysis of HBV drug resistant mutants, and measures to prevent antiviral resistant HBV.

Nucleoside analogs mainly target the different viral polymerase activities, i.e. RNA dependent DNA synthesis (reverse transcription) and DNA dependent DNA synthesis (1). This results in the inhibition of infectious virion production and a decreased rate of infection of new hepatocytes. However, none of the available polymerase inhibitors have been shown to prevent infection of uninfected hepatocytes and the *de novo* formation of covalently closed circular (ccc) DNA (2). Antivirals have a modest indirect effect on cccDNA by inhibiting the intracellular recycling of nucleocapsids, but the long half-life of infected hepatocytes and cccDNA necessitates very long duration of therapy placing patients at risk of developing drug resistance (1, 3).

How Do Antiviral-Resistant HBV Mutants Arise ?

Hepatitis B virus is a DNA virus that replicates via reverse transcription of pregenomic RNA. Unlike DNA polymerase, reverse transcriptase does not have proofreading ability. Therefore, HBV has a higher error rate than other DNA viruses. The high error rate and the large amount of virions produced (10^{12} - 10^{13} per day) mean that every possible mutation is generated daily (4). Because of the overlapping open reading frames, many of the spontaneous mutations may be detrimental to the virus and would not be propagated. Nevertheless, some spontaneously

occurring mutations may persist and antiviral-resistant HBV mutants may be generated spontaneously before exposure to nucleos(t)ide analogue treatment (5).

Antiviral-resistant HBV mutants are selected in the presence of antiviral therapy because they can replicate better than wild-type virus in the presence of antiviral drug. Various factors are associated with an increased likelihood of antiviral-resistant mutant selection. Patients with high pretreatment HBV DNA levels, those with slow or inadequate viral suppression, and those who had resistance to prior treatment that are cross-resistant to the current therapy are at increased risk of developing antiviral resistance (6, 7). The likelihood of antiviral resistance is higher if the treatment has intermediate antiviral activity (enough to exert selective pressure on the virus but not enough to result in rapid suppression of HBV replication) and a low genetic barrier to resistance (marked decrease in susceptibility due to a single amino acid substitution not affecting viral genome replication capacity). Host factors that contribute to an increased likelihood of antiviral resistance include non-compliance with medications, immunosuppression (resulting in enhanced replication by loss of immune control of infected cells) (8), abundant replication space (large number of uninfected hepatocytes as in the newly transplanted liver) (9), and possibly polymorphisms in kinases responsible for converting prodrugs to active drugs or for phosphorylation of nucleos(t)ide analogues to the active triphosphates.

Definition of HBV drug resistance

Clinically, HBV drug resistance manifests at different levels (10, 11): 1) *Genotypic resistance* corresponds to the detection of mutation(s) in the HBV genome which have been found to be selected during antiviral therapy and to confer resistance to the antiviral agent; i.e., the detection of specific HBV polymerase gene mutations; 2) *Virologic breakthrough* corresponds to a

confirmed increase (2 consecutive occasions) in serum HBV DNA levels by $>1 \log_{10}$ copies/mL during therapy, compared to nadir, in a medication compliant patient; 3) *Biochemical breakthrough* is defined as an increase in serum aminotransferase (ALT) levels that accompanies a virologic breakthrough. In most patients, these three levels of drug resistance present successively with intervals of months or years. However, progression from genotypic resistance to biochemical breakthrough can be abrupt particularly in patients who are immunosuppressed or have underlying cirrhosis.

Another situation of treatment failure is a *suboptimal treatment response and primary non response*. The definition is still debated but there is some agreement to define it by the inability of antiviral treatment to reduce serum HBV DNA by $\geq 2 \log_{10}$ IU/ml and to a level $<4 \log_{10}$ IU/ml after the first 6 months of therapy. This phenomenon is likely to be due to pharmacologic issue including the pharmacodynamics of the drug and its antiviral potency as well as patient's compliance. Recently, it was suggested that virological factors may be involved as pre-existing mutations, i.e. rtI233V, were associated with primary non response to adefovir in 3 patients (12).

Cross-resistance is defined as a mutation(s) that confers resistance to more than one antiviral drug by *in vitro* testing. This usually translates into failure of the other drugs as rescue therapy in the clinical setting. The knowledge of cross-resistance is therefore critical in developing treatment strategies after treatment failure to the first antiviral drug. The results of cross-resistance analysis for the most common HBV antiviral-resistant mutants are summarized in Table 1.

Phenotypic assays

Phenotypic assays are crucial in confirming if HBV polymerase mutations observed during

antiviral therapy confer resistance to the drug and to determine if those mutations have cross-resistance to other drugs. Several assays has been used to determine the in vitro phenotype of antiviral-resistant mutants and have been reviewed recently (13) : 1) Site directed mutagenesis of laboratory strains cloned in appropriate vectors and transient transfection of hepatoma cell lines; 2) Exchange of polymerase gene fragments between HBV genomes from clinical isolates and the laboratory strain; 3) Subcloning of the whole HBV polymerase gene; 4) Amplification of the whole HBV genome and its transfection in hepatoma cell lines, either directly or after a cloning step; 5) Cloning of 1.1 unit length HBV genome from clinical isolates and their transfection in cell lines. In each case, viral genome replication capacity is determined in the presence or absence of drugs. Other approaches have been used such as the cloning of HBV mutants in recombinant baculovirus vectors and the transduction of hepatoma cell lines by these baculoviruses.

Phenotypic assays are tedious but they have been instrumental in characterizing the mutations that confer resistance to lamivudine, adefovir, entecavir and telbivudine; the susceptibility of these mutants to other antiviral agents; and the differentiation between primary resistant mutations and secondary compensatory mutations (14-20). Phenotypic assays have demonstrated that combination of antiviral drugs does not adequately suppress multi-drug resistant mutants selected after sequential monotherapies (21, 22).

Another important aspect of phenotypic assays is the assessment of replication fitness of the drug resistant mutants, i.e. their capacity to spread in the liver and outgrow wild type virus (9). The principle relies on the production of HBV mutants and the inoculation of susceptible cells, i.e. primary human or tupaia hepatocytes and/or the highly differentiated HepaRG cell line (13).

However, the number of infected cells and virus production in these systems are low. Another approach is to inoculate humanized chimeric mice with these mutants to study their infectivity *in vivo* (23). These assays to study viral fitness are being improved to provide new insight in the selection process of the HBV drug resistant mutants.

Diagnosis of HBV Drug Resistance

Direct sequencing is the most convenient method for identifying resistance mutations to new treatments. However, direct sequencing is insensitive. Whereas direct sequencing can detect mutants that constitute 10% to 20% of the virus population, studies in patients with human immunodeficiency virus (HIV) infection who have received highly active antiretroviral treatment have shown that resistant mutants are not consistently detected unless they constitute $\geq 40\%$ of the virus population. For nucleos(t)ide analogues with well-characterized resistance mutations, more sensitive methods that can consistently detect mutants when present in $\geq 10\%$ of the virus population should be used (24). These more sensitive methods permit earlier detection of resistance mutants, prior to biochemical breakthrough (25, 26). Recent studies showed that institution of rescue therapy at this early stage is more effective. Early detection of antiviral resistance is particularly important for patients with cirrhosis and in liver transplant recipients. Restriction fragment length polymorphism and reverse hybridization using strips coated with oligonucleotide probes (line probes) are the most common methods used for detecting antiviral-resistant HBV mutations. They are simple and can provide results within 1 day; however, they can only detect previously characterized mutations, and adaptation of these methods to detect mutants that confer resistance to a growing list of nucleos(t)ide analogues will be challenging (27).

Incidence of Antiviral-Resistant HBV

Although antiviral-resistant HBV mutations can occur spontaneously, these mutants are present as a minor virus species (< 0.1%) in a large pool of viruses in most HBV carriers who have not been exposed to nucleos(t)ide analogues. Thus, unless ultra-sensitive assays are used to selectively amplify the mutants, antiviral-resistant HBV cannot be detected in patients who have not received nucleos(t)ide analogue treatment (5). Among patients receiving a specific nucleos(t)ide analogue, the frequency of detection of antiviral-resistant HBV mutants correlates with pretreatment serum HBV DNA level, rapidity of viral suppression, and duration of treatment (7, 28-30). The incidence of genotypic resistance also varies with the sensitivity of the methods used for detection of resistant mutations and the patient population being studied.

Clinical studies of antiviral resistance have varied from testing all patients with detectable HBV DNA by PCR to only patients with confirmed virologic breakthrough, those with viral rebound (increase in HBV DNA to >5 log copies/ml), and those with biochemical breakthrough. The latter approaches result in underestimation of the rate of genotypic resistance, particularly if this is coupled with the use of an insensitive method for detection of antiviral-resistant mutation such as direct sequencing. These differences in methodology make it difficult to compare the incidence of genotypic resistance to various HBV therapies across clinical trials. Variations in patient characteristics (such as pre-treatment HBV DNA level, prior treatment, and liver transplantation) further confound these comparisons, emphasizing the need for standardized definitions and assays for viral resistance.

Clinical impact of HBV drug resistance

The emergence of antiviral resistant strains of HBV leads to viral, and subsequently biochemical breakthrough, negating any benefits achieved during initial antiviral treatment. The clinical outcome for patients with antiviral resistance is related to their age, the severity of the underlying liver disease and the intensity of the hepatitis flare. One study investigated the use of lamivudine for a period of up to 6 years in nearly one thousand patients found that the proportion of patients who experienced hepatitis flares increased from 33% for those with lamivudine-resistant HBV for 1 year to 77% for those who had lamivudine-resistant HBV for more than 4 years (30). Furthermore, hepatic decompensation occurred in only 0% to 2% of patients with lamivudine resistance during the first 4 years, but thereafter the risk of developing hepatic decompensation increased to 6%. The favorable outcome in this study could be related to the relatively young age of the patients (mean: 32 years) and the fact that most patients had early-stage liver disease (only 10% had cirrhosis) (30). Another study examined the efficacy of lamivudine vs. placebo in the prevention of disease progression for patients with advanced liver fibrosis / cirrhosis (31). After a median of 32 months, 49% of the patients in the lamivudine-treated group had genotypic resistance. The Child-Pugh scores were more likely to be increased in these patients than in those without lamivudine resistant mutants (7% vs. <1%, $p<0.001$). Eight of the 14 deaths in the lamivudine-treated group were related to emergence of lamivudine resistance (31). Despite these complications, patients with lamivudine resistance were less likely to develop progressive liver disease than the placebo-treated patients indicating a clinical benefit of lamivudine therapy, during the study period.

However, there have been many reports that emergence of lamivudine resistance was associated with severe hepatitis flare, liver failure, and death, especially in older patients and those with

cirrhosis or immunosuppression (26, 30, 32). Several studies have also demonstrated that histologic improvements achieved during the first year of treatment are negated with the development of lamivudine resistance. In one study, patients underwent liver biopsies at baseline, year 1, and year 3. Among the patients with lamivudine resistance for more than 2 years, the proportion of patients with reduction in necroinflammation decreased from 56% at year 1 to 36% at year 3, and progression to bridging fibrosis was observed in approximately 27% of patients. By contrast, 83% of the patients without lamivudine resistance experienced a reduction of liver fibrosis in the same period (33).

Patients who develop lamivudine resistance are less likely to undergo hepatitis B e antigen (HBeAg) seroconversion. One study of 58 patients found that after 4-5 years of continued lamivudine therapy, HBeAg seroconversion was achieved in 38% of those with and 75%-80% of those without lamivudine resistance mutations (7).

Most of the data on the clinical impact of antiviral-resistant HBV have been derived from studies of lamivudine therapy. Limited data indicate that resistance to other antiviral drugs such as adefovir is associated with the same consequences: viral rebound, hepatitis flares, and rarely liver failure (34). The rate of resistance to adefovir dipivoxil and to entecavir is lower than that to lamivudine in nucleoside naïve patients (5, 34) (Figure 1). However, in lamivudine experienced patients, the risk of resistance to adefovir dipivoxil (when prescribed in monotherapy) or to entecavir increases significantly (6, 35, 36) (Figure 2).

Implementation of combination prophylaxis of nucleos(t)ide analogues and hepatitis B immune globulin has decreased HBV recurrence post-liver transplant to less than 10%. Currently, antiviral-resistant HBV is the main cause of HBV recurrence, accounting for over 90% of recurrent HBV post-liver transplant (37).

Another consequence of the development of antiviral resistant HBV is the presence of mutations that confer cross-resistance with other nucleos(t)ide analogues that are approved or currently under development, limiting future treatment options. This is of particular concern in young patients. Switching from one treatment to another may result in the selection of multi-drug resistant mutations, especially if sequential monotherapy is used. Recent data indicates that multi-drug resistant mutations collocate in the same HBV genome and are refractory to combination therapy (21, 38, 39). Thus, patients may eventually run out of treatment options.

Finally, a potential concern is the transmission of antiviral-resistant HBV and *de novo* infection with these resistant strains (40). Although antiviral-resistant HBV mutants are supposed to have decreased replication fitness compared to wild-type HBV and are expected to be outgrown by wild-type HBV in a nucleos(t)ide analogue-naïve host, transmission is possible and vigilance is necessary.

Prevention of Antiviral-Resistant HBV

As with other areas of medicine, prevention is better than treatment. Judicious use of nucleos(t)ide analogues in patients with chronic hepatitis B is the most effective prophylaxis against the development of antiviral-resistant HBV. Thus, patients with minimal disease (such as inactive carriers) and those who are unlikely to achieve sustained response (such as HBeAg-positive patients in the immune tolerant phase) should not be treated with nucleos(t)ide analogues, particularly if they are young (< 30 years of age). When possible, the most potent nucleos(t)ide analogue with the lowest rate of genotypic resistance should be administered and compliance reinforced. Although combination therapy has been shown to prevent antiviral resistance in patients with HIV infection, the promise of combination therapy has not yet been fulfilled for

patients with HBV infection. Sequential therapy with drugs that are cross-resistant will lead to stepwise selection of strains harboring increasing number of polymerase gene mutations conferring resistance to the first drug and then to both medications, as in the case of entecavir therapy for patients with lamivudine-resistant HBV. Thus, the benefits of antiviral treatment must be balanced against the risk of drug resistance during long-term treatment. Studies combining drugs with high genetic barriers to resistance and no cross resistance should be conducted to determine the optimal combination therapy for hepatitis B.

Management of Patients with Antiviral-resistant HBV (Figure 3)

Hepatitis B patients receiving antiviral therapy should be closely monitored for virologic breakthroughs. Recent studies indicate that initiating rescue therapy early, at the time of virologic breakthrough, is more effective than delaying until the patients have developed biochemical breakthrough (41).

Patients with lamivudine resistance can be managed by switching to adefovir or by adding adefovir. Several studies found that addition of adefovir is associated with a lower risk of subsequent adefovir resistance. This is confirmed in a large multicenter Italian cohort study involving 588 HBeAg-negative patients who had developed lamivudine resistance and in whom treatment with adefovir was initiated either in combination with or as a substitution for lamivudine. Over a period of up to 36 months, 11% of patients switched to adefovir monotherapy developed genotypic resistance to adefovir, compared to none of the patients who received add-on adefovir. Similarly, virologic breakthrough was only seen in 5% of the add-on group versus 24% in the monotherapy group (42).

Patients with lamivudine resistance can also be managed with entecavir. However, despite the use of a higher dose, virologic rebound due to entecavir resistance was observed in 9% and 17% after 2 and 3 years of entecavir treatment (35). Given our current understanding that pre-existing lamivudine-resistant mutations predisposes to further selection of mutations at 169, 184, 202 and 250 that confer resistance to entecavir (17, 38, 39), entecavir is not an optimal treatment for patients with lamivudine resistance.

Patients with adefovir resistance should be managed according to the cross-resistance profile of the mutant selected by adefovir. In vitro data of phenotypic assays show that the rtN236T mutation is susceptible to lamivudine and entecavir (15, 18, 22). The rtA181V mutation confers a decreased susceptibility to lamivudine but remains sensitive to entecavir (21). A few reports of clinical cases confirm that these patients should be managed according to the profile of resistance mutations (6, 21, 43).

To date, very few cases of entecavir resistance among nucleoside-naïve patients who have received entecavir treatment have been reported. The *in vitro* characterization of the entecavir resistant strains showed that they are also resistant to lamivudine, but remain sensitive to adefovir and tenofovir. A recent report confirmed the initial efficacy of adefovir in decreasing serum HBV DNA level in a patient with entecavir-resistant mutants (38).

Several studies found that the risk of selection of drug resistant mutants increases significantly when serum HBV DNA does not decline below 3 log₁₀ copies /mL after 6 months of lamivudine or telbivudine therapy (28, 44), or after 12 months of adefovir dipivoxil therapy (34). These data suggest that treatment should be adapted early, in patients with partial virologic response to prevent the subsequent selection of resistant viruses.

Conclusion

HBV circulates as a pool of heterogenous viral genomes and its mutants are archived in the cccDNA; thus, HBV drug resistance is bound to occur with nucleoside analog monotherapy. With the development of new nucleoside analogs that exhibit more potent antiviral activity and a lower rate of viral resistance, phenotypic analysis and cross-resistance data will be important for the adaptation of antiviral therapy in patients with drug resistance as well as for the design of combination trials with long-term end-points. HBV drug resistance is becoming a major clinical challenge in patients receiving long-term antiviral therapy. As many patients have now been exposed to several courses of antiviral drugs, the risk of selection of multi-drug resistant mutants

is increasingly common. Based on clinical experience and on results of detailed genotypic and phenotypic analysis, management of patients with drug-resistant HBV is evolving towards earlier add-on therapy. The availability of several antiviral drugs with different resistance patterns and excellent safety profiles offers a unique opportunity for the evaluation of *de novo* combination therapy or early add-on therapy to prevent drug resistance.

References

1. Zoulim F. Mechanism of viral persistence and resistance to nucleoside and nucleotide analogs in chronic hepatitis B virus infection. *Antiviral Res* 2004;64:1-15.
2. Delmas J, Schorr O, Jamard C, Gibbs C, Trepo C, Hantz O, Zoulim F. Inhibitory effect of adefovir on viral DNA synthesis and covalently closed circular DNA formation in duck hepatitis B virus-infected hepatocytes in vivo and in vitro. *Antimicrob Agents Chemother* 2002;46:425-433.
3. Werle-Lapostolle B, Bowden S, Locarnini S, Wursthorn K, Petersen J, Lau G, Trepo C, et al. Persistence of cccDNA during the natural history of chronic hepatitis B and decline during adefovir dipivoxil therapy. *Gastroenterology* 2004;126:1750-1758.
4. Gunther S, Fischer L, Pult I, Sterneck M, Will H. Naturally occurring variants of hepatitis B virus. *Adv Virus Res* 1999;52:25-137.
5. Colonna RJ, Rose R, Baldick CJ, Levine S, Pokornowski K, Yu CF, Walsh A, et al. Entecavir resistance is rare in nucleoside naive patients with hepatitis B. *Hepatology* 2006;44:1656-1665.
6. Fung SK, Chae HB, Fontana RJ, Conjeevaram H, Marrero J, Oberhelman K, Hussain M, et al. Virologic response and resistance to adefovir in patients with chronic hepatitis B. *J Hepatol* 2006;44:283-290.
7. Lai CL, Dienstag J, Schiff E, Leung NW, Atkins M, Hunt C, Brown N, et al. Prevalence and clinical correlates of YMDD variants during lamivudine therapy for patients with chronic hepatitis B. *Clin Infect Dis* 2003;36:687-696.
8. Benhamou Y, Bochet M, Thibault V, Di Martino V, Caumes E, Bricaire F, Opolon P, et al. Long-term incidence of hepatitis B virus resistance to lamivudine in human immunodeficiency virus-infected patients. *Hepatology* 1999;30:1302-1306.
9. Litwin S, Toll E, Jilbert AR, Mason WS. The competing roles of virus replication and hepatocyte death rates in the emergence of drug-resistant mutants: theoretical considerations. *J Clin Virol* 2005;34 Suppl 1:S96-S107.
10. Locarnini S, Hatzakis A, Heathcote J, Keeffe EB, Liang TJ, Mutimer D, Pawlotsky JM, et al. Management of antiviral resistance in patients with chronic hepatitis B. *Antivir Ther* 2004;9:679-693.
11. Zoulim F. Antiviral therapy of chronic hepatitis B. *Antiviral Res* 2006;71:206-215.
12. Schildgen O, Sirma H, Funk A, Olotu C, Wend UC, Hartmann H, Helm M, et al. Variant of hepatitis B virus with primary resistance to adefovir. *N Engl J Med* 2006;354:1807-1812.

13. Zoulim F. In vitro models for studying hepatitis B virus drug resistance. *Semin Liver Dis* 2006;26:171-180.
14. Allen MI, Deslauriers M, Andrews CW, Tipples GA, Walters KA, Tyrrell DL, Brown N, et al. Identification and characterization of mutations in hepatitis B virus resistant to lamivudine. Lamivudine Clinical Investigation Group. *Hepatology* 1998;27:1670-1677.
15. Angus P, Vaughan R, Xiong S, Yang H, Delaney W, Gibbs C, Brosgart C, et al. Resistance to adefovir dipivoxil therapy associated with the selection of a novel mutation in the HBV polymerase. *Gastroenterology* 2003;125:292-297.
16. Durantel D, Carrouee-Durantel S, Werle-Lapostolle B, Brunelle MN, Pichoud C, Trepo C, Zoulim F. A new strategy for studying in vitro the drug susceptibility of clinical isolates of human hepatitis B virus. *Hepatology* 2004;40:855-864.
17. Tenney DJ, Levine SM, Rose RE, Walsh AW, Weinheimer SP, Discotto L, Plym M, et al. Clinical emergence of entecavir-resistant hepatitis B virus requires additional substitutions in virus already resistant to Lamivudine. *Antimicrob Agents Chemother* 2004;48:3498-3507.
18. Villeneuve JP, Durantel D, Durantel S, Westland C, Xiong S, Brosgart CL, Gibbs CS, et al. Selection of a hepatitis B virus strain resistant to adefovir in a liver transplantation patient. *J Hepatol* 2003;39:1085-1089.
19. Yang H, Qi X, Sabogal A, Miller M, Xiong S, Delaney W. Cross-resistance testing of next-generation nucleoside and nucleotide analogues against lamivudine-resistant HBV. *Antivir Ther* 2005;10:625-633.
20. Delaney W, Yang H, Westland CE, Das K, Arnold E, Gibbs CS, Miller MD, et al. The hepatitis B virus polymerase mutation rtV173L is selected during lamivudine therapy and enhances viral replication in vitro. *J Virol* 2003;77:11833-11841.
21. Villet S, Pichoud C, Villeneuve JP, Trepo C, Zoulim F. Selection of a multiple drug-resistant hepatitis B virus strain in a liver-transplanted patient. *Gastroenterology* 2006;131:1253-1261.
22. Brunelle MN, Jacquard AC, Pichoud C, Durantel D, Carrouee-Durantel S, Villeneuve JP, Trepo C, et al. Susceptibility to antivirals of a human HBV strain with mutations conferring resistance to both lamivudine and adefovir. *Hepatology* 2005;41:1391-1398.
23. Dandri M, Lutgehetmann M, Volz T, Petersen J. Small animal model systems for studying hepatitis B virus replication and pathogenesis. *Semin Liver Dis* 2006;26:181-191.
24. Stuyver L, Van Geyt C, De Gendt S, Van Reybroeck G, Zoulim F, Leroux-Roels G, Rossau R. Line probe assay for monitoring drug resistance in hepatitis B virus-infected patients during antiviral therapy. *J Clin Microbiol* 2000;38:702-707.
25. Lok AS, Zoulim F, Locarnini S, Mangia A, Niro G, Decraemer H, Maertens G, et al. Monitoring drug resistance in chronic hepatitis B virus (HBV)-infected patients during lamivudine therapy: evaluation of performance of INNO-LiPA HBV DR assay. *J Clin Microbiol* 2002;40:3729-3734.
26. Nafa S, Ahmed S, Tavan D, Pichoud C, Berby F, Stuyver L, Johnson M, et al. Early Detection of Viral Resistance by Determination of Hepatitis B Virus Polymerase Mutations in Patients Treated by Lamivudine for Chronic Hepatitis B. *Hepatology* 2000;32:1078-1088.
27. Zoulim F. New nucleic Acid diagnostic tests in viral hepatitis. *Semin Liver Dis* 2006;26:309-317.
28. Yuen MF, Sablon E, Hui CK, Yuan HJ, Decraemer H, Lai CL. Factors associated with hepatitis B virus DNA breakthrough in patients receiving prolonged lamivudine therapy. *Hepatology* 2001;34:785-791.

29. Zoulim F, Poynard T, Degos F, Slama A, El Hasnaoui A, Blin P, Mercier F, et al. A prospective study of the evolution of lamivudine resistance mutations in patients with chronic hepatitis B treated with lamivudine. *J Viral Hepat* 2006;13:278-288.
30. Lok AS, Lai CL, Leung N, Yao GB, Cui ZY, Schiff ER, Dienstag JL, et al. Long-term safety of lamivudine treatment in patients with chronic hepatitis B. *Gastroenterology* 2003;125:1714-1722.
31. Liaw YF, Sung JJ, Chow WC, Farrell G, Lee CZ, Yuen H, Tanwandee T, et al. Lamivudine for patients with chronic hepatitis B and advanced liver disease. *N Engl J Med* 2004;351:1521-1531.
32. Hadziyannis SJ, Papatheodoridis GV, Dimou E, Laras A, Papaioannou C. Efficacy of long-term lamivudine monotherapy in patients with hepatitis B e antigen-negative chronic hepatitis B. *Hepatology* 2000;32:847-851.
33. Dienstag JL, Goldin RD, Heathcote EJ, Hann HW, Woessner M, Stephenson SL, Gardner S, et al. Histological outcome during long-term lamivudine therapy. *Gastroenterology* 2003;124:105-117.
34. Hadziyannis SJ, Tassopoulos NC, Heathcote EJ, Chang TT, Kitis G, Rizzetto M, Marcellin P, et al. Long-Term Therapy With Adefovir Dipivoxil for HBeAg-Negative Chronic Hepatitis B for up to 5 Years. *Gastroenterology* 2006.
35. Sherman M, Yurdaydin C, Sollano J, Silva M, Liaw YF, Cianciara J, Boron-Kaczmarek A, et al. Entecavir for treatment of lamivudine-refractory, HBeAg-positive chronic hepatitis B. *Gastroenterology* 2006;130:2039-2049.
36. Lee YS, Suh DJ, Lim YS, Jung SW, Kim KM, Lee HC, Chung YH, et al. Increased risk of adefovir resistance in patients with lamivudine-resistant chronic hepatitis B after 48 weeks of adefovir dipivoxil monotherapy. *Hepatology* 2006;43:1385-1391.
37. Schreiber IR, Schiff ER. Prevention and treatment of recurrent Hepatitis B after liver transplantation: the current role of nucleoside and nucleotide analogues. *Ann Clin Microbiol Antimicrob* 2006;5:8.
38. Villet S, Ollivet A, Pichoud C, Barraud L, Villeneuve JP, Trepo C, Zoulim F. Stepwise process for the development of entecavir resistance in a chronic Hepatitis B virus infected patient *J Hepatol* 2007;in press.
39. Yim HJ, Hussain M, Liu Y, Wong SN, Fung SK, Lok AS. Evolution of multi-drug resistant hepatitis B virus during sequential therapy. *Hepatology* 2006;44:703-712.
40. Thibault V, Aubron-Olivier C, Agut H, Katlama C. Primary infection with a lamivudine-resistant hepatitis B virus. *Aids* 2002;16:131-133.
41. Lampertico P, Vigano M, Manenti E, Iavarone M, Lunghi G, Colombo M. Adefovir rapidly suppresses hepatitis B in HBeAg-negative patients developing genotypic resistance to lamivudine. *Hepatology* 2005;42:1414-1419.
42. Lampertico P, Marzano A, Levrero M, Santantonio T, Di Marco V, Brunetto M, Andreone P, et al. Adefovir and lamivudine combination therapy is superior to adefovir monotherapy for lamivudine-resistant patients with HBeAg-negative chronic hepatitis B. *Hepatology* 2006;44:Abstract LB5, p693A
43. Fung SK, Andreone P, Han SH, Rajender Reddy K, Regev A, Keeffe EB, Hussain M, et al. Adefovir-resistant hepatitis B can be associated with viral rebound and hepatic decompensation. *J Hepatol* 2005;43:937-943.
44. Di Bisceglie AM, Lai CL, Gane E, Chen Y-C, Thongsawat S, Wang Y, Chen Y, et al. Telbivudine globe trial: maximal early HBV suppression is predictive of optimal two-year efficacy in nucleoside-treated hepatitis B patients. *Hepatology* 2006;44:Abstract 112, p230A.

45. Lai CL, Leung N, Teo EK, Tong M, Wong F, Hann HW, Han S, et al. A 1-year trial of telbivudine, lamivudine, and the combination in patients with hepatitis B e antigen-positive chronic hepatitis B. *Gastroenterology* 2005;129:528-536.

Legends

Figure 1 – Incidence of Genotypic Resistance in Nucleoside Naïve Patients.

Results were obtained from clinical studies (5, 7, 29, 30, 34, 45)

Figure 2 – Incidence of Genotypic Resistance in Lamivudine Resistant Patients

Results of selected studies are presented (35, 42)

Figure 3 – Recommendations for the Management of HBV Drug Resistance

Resistance rate of selected studies are presented (6, 35, 36, 42). Recommendations are based on the results of phenotypic analysis and clinical studies.

Table 1: Results of in vitro cross-resistance analysis

Results were obtained after in vitro phenotypic analysis of the resistant mutants (15, 17-19, 21, 22, 38)

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