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The Glycine Transporter GlyT2 Controls the Dynamics of Synaptic Vesicle Refilling in Inhibitory Spinal Cord Neurons

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At inhibitory synapses, glycine and GABA are accumulated into synaptic vesicles by the same vesicular transporter VGAT/VIAAT (vesicular GABA transporter/vesicular inhibitory amino acid transporter), enabling a continuum of glycine, GABA, and mixed phenotypes. Many fundamental aspects of the presynaptic contribution to the inhibitory phenotypes remain unclear. The neuronal transporter GlyT2 is one of the critical presynaptic factors, because glycinergic transmission is impaired in knock-out *GlyT2*^{-/-} mice and mutations in the human GlyT2 gene *slc6a5* are sufficient to cause hyperekplexia. Here, we establish that GlyT2-mediated uptake is directly coupled to the accumulation of glycine into recycling synaptic vesicles using cultured spinal cord neurons derived from GlyT2–enhanced green fluorescent protein transgenic mice. Membrane expression of GlyT2 was confirmed by recording glycine-evoked transporter current. We show that GlyT2 inhibition induces a switch from a predominantly glycine to a predominantly GABA phenotype. This effect was mediated by a reduction of glycinergic quantal size after cytosolic depletion of glycine and was entirely reversed by glycine resupply, illustrating that the filling of empty synaptic vesicles is tightly coupled to GlyT2-mediated uptake. Interestingly, high-frequency trains of stimuli elicit two phases of vesicle release with distinct kinetic requirements for glycine refilling. Thus, our results demonstrate the central role played by GlyT2 in determining inhibitory phenotype and therefore in the physiology and pathology of inhibitory circuits.

Key words: inhibitory transmission; glycine transporter; vesicular transporter; quantal size; spinal cord; paired recordings

Introduction

New and recycled synaptic vesicles are filled with a high concentration of neurotransmitter by specific vesicular H⁺ antiporters (Sulzer and Pothos, 2000; Edwards, 2007), yet despite the importance of this loading step for the completeness of vesicle recycling, little is known about its dynamics and the thermodynamic, kinetic, and osmotic constraints that regulate the vesicular storage of neurotransmitters under physiological conditions (Sulzer and Pothos, 2000; Liu, 2003; Edwards, 2007). In particular, it is not known how glycinergic, GABAergic, and mixed GABAergic/glycinergic vesicular phenotypes are specified at individual synapses in inhibitory neurons (Jonas et al., 1998), in which vesicular inhibitory amino acid transporter (VIAAT) (also named VGAT, for vesicular GABA transporter) is the unique vesicular transporter for the two inhibitory amino acids neurotransmitters (Burger et al., 1991; McIntire et al., 1997; Sagné et al., 1997; Wojcik et al., 2006; Aubrey et al., 2007). Although the proportions of glycine

and GABA in vesicles are expected to be determined by the relative concentrations of the neurotransmitters in the cytoplasm, vesicles released by isolated terminals (Katsurabayashi et al., 2004) or by VIAAT-transfected cells (Aubrey et al., 2007) evoked heterogeneous patterns of transmission, suggesting heterogeneous vesicular loading.

Although glycine is an ubiquitous intracellular metabolite, its basal synthesis cannot account for the 10- to 100-fold larger accumulation detected in glycinergic neurons (Daly, 1990; Ottersen et al., 1990). Thus, local recapture is considered to be the main mechanism of glycine supply at the terminals. Glycinergic neurons in the spinal cord and the brainstem express GlyT2, a transporter of the SLC6 family (Liu et al., 1993; Jursky and Nelson, 1995; Zafra et al., 1995) that mediates unidirectional transport of glycine coupled to 3 Na⁺/1 Cl⁻ (Roux and Supplisson, 2000). In a model system of inhibitory transmission, it has been shown that the only requirement for glycine accumulation into vesicles is the coexpression of GlyT2 and VIAAT (Aubrey et al., 2007). In addition, glycinergic transmission is impaired in brainstem slices of *GlyT2*^{-/-} knock-out mice (Gomez et al., 2003) and sporadic mutations in human *slc6a5*, the gene that encodes GlyT2 that abolishes glycine uptake, cause hyperekplexia (Rees et al., 2006), a neuromuscular startle syndrome attributed principally to mutations in the $\alpha 1$ subunit of the glycine receptor (GlyR) (Shiang et al., 1993).

Here, we investigate the relationship between GlyT2-mediated glycine uptake and vesicle filling in pairs of spinal cord neurons in culture. A GlyT2–enhanced green fluorescent protein (EGFP) transgenic mouse line enabled identification of GlyT2-positive (GlyT2⁺) neurons (Zeilhofer et al., 2005), and the cul-

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ture preparation allowed us to control intracellular and extracellular glycine. We show that uptake by GlyT2 drives the filling of synaptic vesicles with glycine and regulates quantal size in inhibitory neurons. Interestingly, we found that blocking GlyT2 induces a switch from predominantly glycinergic to predominantly GABAergic phenotypes. In addition, we highlight kinetic constraints in the supply of glycine for vesicle refilling during high-frequency release.

Materials and Methods

Embryonic mouse spinal cord neurons. Primary cultures of spinal cord neurons were prepared as described by Hanus et al. (2004) from embryonic day 13 or 14 C57BL/6 wild-type (WT) or heterozygous GlyT2–EGFP mouse pups (Zeilhofer et al., 2005). Embryos were obtained by cesarean section from pregnant mice anesthetized by intraperitoneal injection of ketamine–xylazine (100 and 10 mg/kg) and killed by cervical elongation. Spinal cords were dissected under sterile conditions into PBS with 33 mM glucose at pH 7.4 and then incubated in trypsin/EDTA solution (0.05% v/v; Sigma) for 10 min at 37°C. Cells were dissociated mechanically in a modified L15 Leibowitz's medium (Invitrogen) and plated at a density of 0.4×10^5 cells/cm² on sterilized glass coverslips coated with 60 μg/ml poly-DL-ornithine and with medium containing 5% inactivated fetal calf serum (Sigma). Cells were maintained at 37°C in 5% CO₂ in serum-free Neurobasal medium containing supplement B27 (Invitrogen) (Brewer et al., 1993) for up to 3 weeks. Medium was changed every 4–5 d.

Electrophysiology: procedures and solutions. Whole-cell patch-clamp recordings of spinal cord neurons [12–22 d *in vitro* (DIV)] were performed at ~30°C. Voltage-clamp and current-clamp modes were applied using a Multiclamp 700B or two Axopatch 200A amplifiers controlled by pClamp 9 or 10 acquisition software (Molecular Devices). Currents were filtered at 4 kHz and sampled at 20 kHz using a Digidata 1440A or 1322A (Molecular Devices). Patch pipettes were pulled from borosilicate glass capillaries (Hilgenberg) and had resistances of 4–6 MΩ. Miniature and evoked currents were recorded at a holding potential (V_H) of –60 mV using pipettes filled with a standard internal solution containing the following (in mM): 140 CsCl, 1 CaCl₂, 10 EGTA, 1 BAPTA, 1 MgCl₂, 4 Mg-ATP, 5 QX314 [N-(2,6-dimethylphenylcarbamoyl-methyl)triethylammonium-Cl], and 10 HEPES, adjusted to pH 7.4 with CsOH. For paired recordings, the internal solution for the presynaptic neurons contained the following (in mM): 155 K-gluconate, 4 KCl, 5 Mg-ATP, 0.1 EGTA, and 10 HEPES, adjusted to pH 7.4 with KOH. To record GlyT2-mediated currents, the internal solution was the following (in mM): 130 CsCH₃O₃S, 1 CaCl₂, 10 EGTA, 1 BAPTA, 5 Mg-ATP, 5 tetraethylammonium-Cl, 5 QX314-Cl, and 10 HEPES, adjusted to pH 7.4 with CsOH. Neurons were continuously bathed with an external solution containing the following (mM): 140 NaCl, 2.4 KCl, 4 CaCl₂, 1 MgCl₂, 10 glucose, and 10 HEPES, pH 7.4. AMPA and NMDA receptors were blocked with 2 μM 6-nitro-7-sulfamoylbenzo[f]quinoxaline-2,3-dione (NBQX) and 50 μM D-(±)-2-amino-5-phosphonovaleric acid (D-APV), respectively. GABA_A receptors (GABA_ARs) were selectively blocked with 5 μM gabazine (SR95531 [2-(3-carboxypropyl)-3-amino-6-(4-methoxyphenyl)pyridinium bromide]). GABA_B receptors (GABA_BRs) were blocked with CGP55845 [(2S)-3-((1S)-1-(3,4-dichlorophenyl)ethyl)amino-2-hydroxypropyl](phenylmethyl)phosphonic acid]. GlyRs were blocked with 3 μM strychnine, and larger concentrations (10–30 μM) were used when glycine was present in the external solution. GlyT2 was inhibited with 5 μM ORG25543 (4-benzyloxy-3,5-dimethoxy-N-[(1-dimethylaminocyclopentyl)methyl]benzamide) (generous gift from H. Sundaram and D. Hill, Organon, Newhouse, UK); other drugs were purchased from Sigma or Tocris Cookson.

Miniature events were recorded in the presence of 0.2 μM tetrodotoxin (TTX) from neurons isolated from WT or GlyT2–EGFP embryos between 16 and 22 DIV.

For paired recordings, presynaptic GlyT2⁺ neurons and putative postsynaptic neurons [GlyT2⁺ or GlyT2-negative (GlyT2[–])] were patched and tested for synaptic connection. Action potentials (APs) were triggered under current clamp by 10 ms depolarizing current steps com-

prised between 140 and 360 pA at 0.1–5 Hz. Evoked IPSCs were recorded in the presence of 50 μM D-APV and 2 μM NBQX.

GlyT2-mediated currents were recorded from neurons expressing EGFP, between 18 and 22 DIV. Neurons were held at V_H of –80 mV and bathed with standard external solution containing TTX (0.2 μM) and cadmium chloride (20 μM). Current–voltage (I – V) relationships were isolated by subtraction of glycine (200 μM) stimulated steady-state currents from those recorded in the external solution alone. Currents were normalized to the absolute value recorded at V_H of –50 mV.

Data analysis. Miniature synaptic currents were detected using the Clampfit template procedure. Recordings with less than five glycinergic miniature IPSCs (mIPSCs) were discarded from the amplitude analysis. To take into account combined changes in amplitude and frequency, the sum of mIPSC amplitudes per minute was calculated for each cell.

The latency for paired recordings was measured as the delay between the peak of the action potential and the maximum of the rising slope of the postsynaptic current. Evoked IPSC decays were fitted by one exponential or by the sum of two exponentials $I(t) = A_1 \exp(-t/\tau_1) + A_2 \exp(-t/\tau_2)$, where A_1 and A_2 are the amplitudes of the components (with identical sign), and τ_1 and τ_2 are the time constants. The amplitude-weighted decay time constant was calculated as $\tau_w = (A_1\tau_1 + A_2\tau_2)/(A_1 + A_2)$. CV² analysis was used to predict the locus of synaptic depression (Faber and Korn, 1991). For each train of 1000 APs, mean amplitude (μ), SD (σ), and coefficient of variation ($CV = \sigma/\mu$) of 40 IPSCs were computed at the onset (APs 2 and 41; CV_1, μ_1) and at the end (CV_2, μ_2) of the stimulation. Then, $\frac{CV_1^2}{CV_2^2}$ was plotted as a function of $\frac{\mu_2}{\mu_1}$ as described by Faber and Korn (1991).

Responses evoked by 5 Hz stimulation were normalized by the first IPSC amplitude or by the average amplitude between APs 3 and 32. In semilogarithmic plots, IPSC amplitudes between the APs 4 and 1000 were averaged using a constant logarithmic bin of 0.1.

Electrophysiological results are reported as mean ± SEM. All statistical tests were nonparametric. The Mann–Whitney U test and the sign test were used to assess differences between two independent and two related samples, respectively. The Kolmogorov–Smirnov test was used to assess the equality of the two distributions. For all tests, the number of asterisks in the figures corresponds to level of significance: * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$.

FM 4-64 imaging. GlyT2–EGFP spinal cord neurons were incubated for 3 min at 37°C in a depolarizing extracellular solution containing the following: 102.4 mM NaCl, 40 mM KCl, 1 mM MgCl₂, 2 mM CaCl₂, 10 mM glucose, 10 mM HEPES, pH 7.3, and 10 μM FM 4-64 [N-(3-triethylammoniumpropyl)-4-(6-(4-diethylamino)phenyl)hexatrienyl]pyridinium dibromide] (Invitrogen). Coverslips were washed for 20 min twice with the external solution containing TTX (0.5 μM) in the presence (1 mM) or absence of ADVASEP-7 [sulfonated β-cyclodextrin (Sigma)] (Kay et al., 1999). FM 4-64-labeled puncta were destained at 30°C by electrical stimulation as reported by Jungling et al. (2006) using large-diameter pipettes filled with standard extracellular solution attached to an Axopatch 800A (Molecular Devices). One-second bursts of square pulses (–60 V, 2 ms in duration) were applied at 10 Hz. Fluorescence images (11007v2 filter; Chroma Technology) were acquired at 0.2 Hz using a Coolsnap-ES CCD camera (Roper Scientific) and a LUCPLFL 40×/0.6 relief contrast objective (Olympus) with MetaView 5 software (Universal Imaging). Illumination (200 ms) was controlled with an electronic shutter (Uniblitz shutters; Vincent Associates). For time-lapse recordings, the average fluorescence intensities of individual boutons were normalized after background subtraction to the initial fluorescence intensity recorded before stimulation. Only boutons with colocalized FM 4-64 and EGFP fluorescence were analyzed. Final images were processed using Adobe Photoshop 9.0 software (Adobe Systems).

Glycine iontophoresis. Iontophoresis pipettes were pulled from thin-walled glass (WPI) to have a resistance of 150–200 MΩ when filled with 15 mM glycine, pH 2. The iontophoresis pipette was then moved close to neurites originating from the patch-clamped neuron. Fast glycine responses were elicited using a stimulator (A-M Systems 2100) with positive voltage steps of 1 ms in the range of 5–25 V and a small holding voltage (–0.1 to –1 V). The iontophoresis pipette position and holding

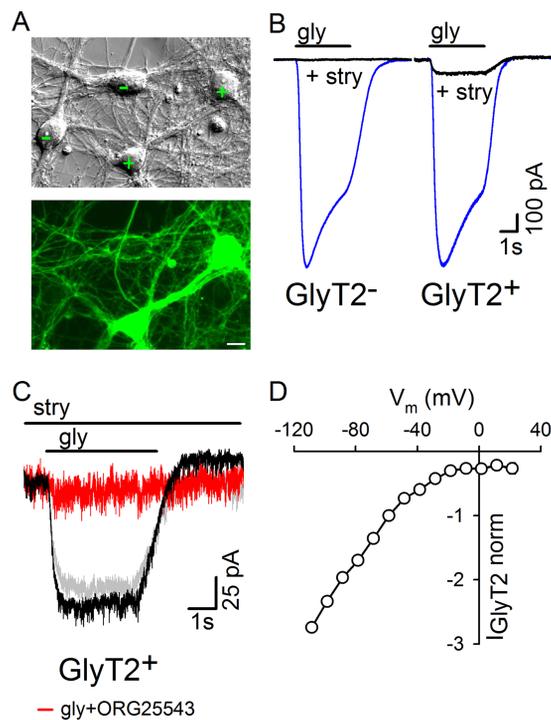


Figure 1. GlyT2-mediated currents in cultured spinal cord neurons. **A**, GlyT2 positive (+) or negative (–) neurons isolated from GlyT2–EGFP mice (top) were identified by their green fluorescence (bottom). Scale bar, 10 μ m. **B**, Traces of currents evoked by glycine (200 μ M) in GlyT2[–] neurons (left trace) and GlyT2⁺ neurons (right trace) in the absence (blue line) or presence (20 μ M; solid line) of strychnine (stry). **C**, The strychnine-insensitive glycine-evoked current of GlyT2⁺ neurons (solid line) was inhibited by ORG25543 (5 μ M; red line), reversibly (gray line; 40 min wash). **D**, *I*–*V* relationship of the GlyT2-mediated current (*I*–*V* values were normalized by their absolute values at V_m of –50 mV; $n = 10$).

voltage were not changed once it was optimally adjusted, and the amount of glycine release was modulated by changing the intensity of the positive voltage step. SR95531 (70 and 500 μ M) was bath applied. Control solutions all contained 5 μ M SR95531 (see above, Electrophysiology: procedures and solutions). The current traces presented in Figure 6A are the average from 20–30 consecutive iontophoretic pulses. The coefficients of variation were 0.197, 0.074, and 0.026 at 10, 15, and 20 V stimulation, respectively.

Results

We set out to study the role of GlyT2 for the refilling of synaptic vesicles with glycine, in pairs of cultured spinal neurons with identified presynaptic GlyT2 neurons, using transgenic GlyT2–EGFP mice (Zeilhofer et al., 2005).

GlyT2 transporter currents

To confirm the functional expression of GlyT2 in green fluorescent neurons, we examined the glycine-evoked current recorded from GlyT2⁺ or GlyT2[–] neurons (Fig. 1A) (50 \pm 1.7% GlyT2⁺ neurons/coverslip; $n = 127$). In all neurons tested, fast application of glycine (200 μ M) generated large inward currents (Fig. 1B, blue line), which slowly desensitized. In GlyT2[–] neurons, these responses were abolished by strychnine (20 μ M) (inhibition, 98.5 \pm 1.3%; $n = 4$) (Fig. 1B, left panel, solid line). In contrast, in GlyT2⁺ neurons, a stationary strychnine-insensitive current remained, with a mean amplitude of –43.5 \pm 6.8 pA ($n = 14$) (Fig. 1B, right panel, solid line). This residual current displayed the hallmarks of a transporter current because it was reversibly blocked by a specific GlyT2 inhibitor (5 μ M ORG25543; inhibition, 97.9 \pm 1.4%; $n = 5$) (Fig. 1C) and did not reverse between

–100 and +30 mV (Fig. 1D), as expected for a tightly coupled transporter. This recording of a glycine transporter current validated the fluorescence identification of GlyT2-expressing neurons in our spinal cord cultures.

GlyT2 determines the neuronal glycinergic phenotype

To examine synaptic transmission in GlyT2⁺ neurons, we recorded evoked postsynaptic currents (IPSCs) in pairs of connected neurons with an identified GlyT2⁺ presynaptic element (Fig. 2A). Presynaptic APs were triggered in current clamp by injection of depolarizing current pulses and IPSCs were recorded in connected voltage-clamped neurons (13 GlyT2⁺ and 3 GlyT2[–]) (Fig. 2A). The strength of synaptic transmission was variable between pairs (range, –35 to –2900 pA; $n = 16$), although the low coefficient of variation (CV of 0.2 \pm 0.03), the short latency (1.6 \pm 0.1 ms), and the absence of failures all indicated that the connections were monosynaptic. SR95531, a specific GABA_A receptor antagonist at 5 μ M, blocked one-quarter of the IPSC amplitude (26.3 \pm 3.3%; $n = 30$), whereas the remaining current was completely eliminated by strychnine (Fig. 2B,C).

The majority of GlyT2⁺ neurons (27 of 30) displayed a dominant (>50%) glycinergic phenotype (Fig. 2D), whereas the evoked postsynaptic currents of GlyT2[–] presynaptic neurons were either purely GABAergic, with a single component sensitive to SR95531 ($n = 5$ of 9), or glutamatergic, with evoked currents entirely blocked by a combination of NBQX and D-APV ($n = 4$ of 9; data not shown).

Having characterized control inhibitory transmission, we first investigated the role of GlyT2 by preincubating the cultures with ORG25543, a specific GlyT2 inhibitor (Caulfield et al., 2001). Inhibition of GlyT2 by ORG25543 for 12–24 h led to an increase in the weighted decay time constant of the evoked IPSC, from 23.0 \pm 3.2 ms ($n = 29$) in control neurons to 50.6 \pm 7.3 ms ($n = 24$) in ORG25543-incubated neurons ($p < 0.001$) (Fig. 2F). However, the average decay time constant of the glycinergic component decreased from 12.5 \pm 1.0 ms ($n = 30$) in control to 9.8 \pm 1.0 ms ($n = 20$) in ORG25543-treated ($p < 0.03$) neurons (Fig. 2B–F). This suggests an inversion in the relative contributions of fast glycinergic and slow GABAergic currents to the IPSCs. Indeed, the peak amplitude of the glycinergic component of the IPSCs decreased from –680 \pm 111 pA ($n = 30$) in control to –213 \pm 51 pA ($n = 26$) in ORG25543-treated ($p < 0.001$) neurons (Fig. 2E), whereas the amplitude of the GABAergic component increased significantly from –264 \pm 59 pA ($n = 30$) to –481 \pm 100 pA ($n = 26$; $p < 0.02$) (Fig. 2E). Furthermore, the decay time constant of the GABAergic IPSC increased from 36.4 \pm 4.5 ms ($n = 26$) to 53.5 \pm 7.7 ms ($n = 21$; $p < 0.02$) (Fig. 2F), suggesting an increase in released GABA (Jones and Westbrook, 1995). This phenotypic switch from dominant glycinergic to dominant GABAergic (Fig. 2C,D) was of presynaptic origin because the whole-cell current amplitude evoked by glycine (200 μ M) was similar in control (–4.0 \pm 0.5 nA; $n = 7$) and ORG25543-treated (–3.8 \pm 0.4 nA; $n = 5$; $p > 0.1$) neurons. Therefore, the reduction in amplitude of glycinergic IPSCs reflected a decrease in the amount of neurotransmitter released. Formally, this reduction could occur via one or both of two mechanisms: reduction in the number of glycinergic vesicles released (quantal content) or of their glycine content (quantal size).

Chronic GlyT2 inhibition reduces glycine quantal size

To determine whether the quantity of glycine in vesicles was reduced by preincubation with ORG25543, we recorded glycinergic mIPSCs in control and ORG25543-treated cells. Recordings

were performed in nominally glycine-free solution to prevent reaccumulation of glycine by the cells. Glycinergic mIPSCs isolated in the presence of TTX, NBQX, d-APV, and SR95531 (Fig. 3A) were severely reduced in neurons preincubated overnight with ORG25543 compared with control neurons (Fig. 3A–C), with a 39% decrease of the mean mIPSC amplitude, from -48.9 ± 3.7 pA ($n = 41$) to -29.8 ± 0.9 pA ($n = 57$; $p < 0.001$). The time course of mIPSCs was faster in ORG25543-treated neurons (5.0 ± 0.24 ms; $n = 57$) than in control neurons (6.1 ± 0.32 ms; $n = 41$; $p < 0.01$) (Fig. 3C). Nevertheless, the main effect of GlyT2 inhibition was to decrease the mIPSC frequency, from 3.6 ± 0.4 Hz ($n = 41$) in control neurons to 0.6 ± 0.13 Hz ($n = 72$; $p < 0.001$) in ORG25543 neurons (Fig. 3B). We examined whether this reduction in frequency corresponded to a decrease in mIPSC amplitude below the detection threshold by recording mIPSCs in the presence of zinc (Fig. 4A, B), a positive high-affinity allosteric modulator of GlyRs (Bloomenthal et al., 1994; Laube et al., 1995; Suwa et al., 2001; Laube, 2002). In both control and ORG25543-treated neurons, addition of $1 \mu\text{M}$ free zinc increased the mean amplitude of mIPSCs by $17.8 \pm 4\%$ ($n = 10$) and $12.8 \pm 6\%$ ($n = 6$), respectively (Fig. 4C, D). Although zinc doubled the frequency of mIPSCs in ORG25543-treated neurons (Fig. 4D), glycinergic mIPSC activity remained much lower than in control neurons (Fig. 4A–C), indicating that a large majority of synaptic vesicles in ORG25543-treated neurons do not release glycine or not enough to activate postsynaptic receptors and can therefore be considered as “glycine-empty.”

To assess the specificity of ORG25543 action, we recorded glycinergic, GABAergic, and glutamatergic mPSCs successively and compared their average activity expressed as the sum of mIPSC amplitudes per unit of time. Although the cumulative glycinergic mIPSC activity showed a 96% reduction, from 5.1 ± 1.6 nA/min ($n = 9$) in control to 0.22 ± 0.07 nA/min ($n = 11$) in ORG25543-treated neurons (Fig. 3D, $p < 0.01$), the average GABAergic activity increased from 0.65 ± 0.28 to 0.99 ± 0.32 nA/min (Fig. 3D, $p > 0.1$) primarily attributable to an increase in GABAergic mIPSC frequency (Fig. 3E), although the difference was not statistically significant. The average glutamatergic mEPSC activity was similar in control and ORG25543-treated neurons (8.9 ± 1.7 nA/min, $n = 9$ and 9.3 ± 1.7 nA/min, $n = 11$ for control and ORG25543-treated neurons, respectively) (Fig. 3E, $p > 0.5$), with no statistical difference in amplitude or frequency (Fig. 3E).

To examine whether preincubation with ORG25543 in some way altered exocytosis and/or endocytosis, neurons were loaded with FM 4-64 and their fluorescence was analyzed using conventional imaging techniques (Gaffield and Betz, 2006) at individual GlyT2⁺ boutons identified by the colocalization of FM 4-64 puncta and EGFP fluorescence (Fig. 3F). Comparable FM 4-64 fluorescence intensities were detected in control and ORG25543-

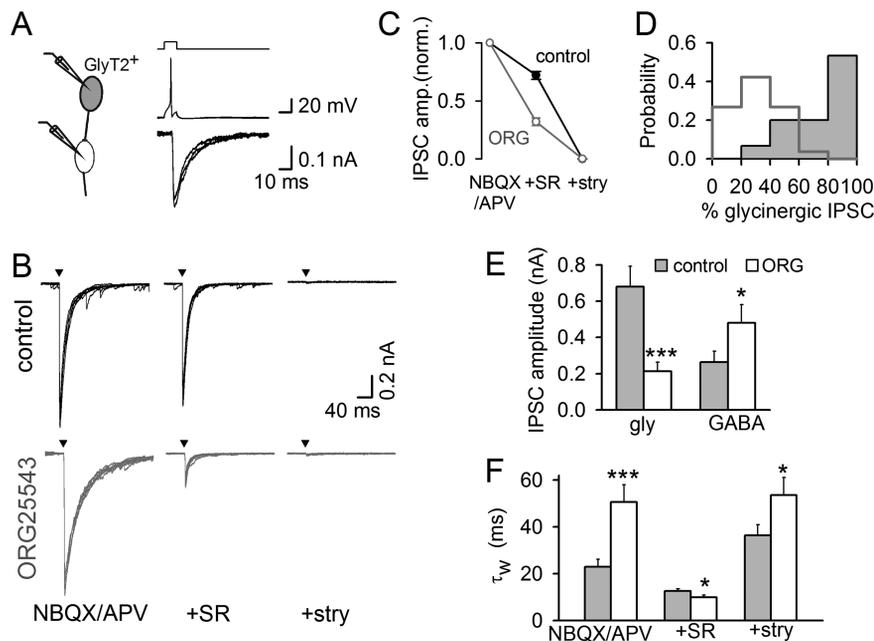


Figure 2. GlyT2 determines the neuronal glycinergic phenotype. *A*, Left, Schematic of a paired recording with an identified presynaptic GlyT2⁺ neuron. Right, Depolarizing current steps trigger spikes in a GlyT2⁺ neuron that evoked IPSCs in a connected neuron held at V_h of -60 mV. *B*, Representative IPSCs ($n = 7$) recorded in the presence of NBQX ($2 \mu\text{M}$) and d-APV ($50 \mu\text{M}$; left traces), SR95531 ($5 \mu\text{M}$; middle traces; SR), or strychnine ($3 \mu\text{M}$; right traces; stry) for control neurons (top, black) and neurons preincubated for 15–24 h with $5 \mu\text{M}$ ORG25543 (bottom, gray). *C*, Amplitude of the IPSCs recorded as described in *B*, for control neurons (black; $n = 30$) and neurons preincubated with $5 \mu\text{M}$ ORG25543 (gray; $n = 26$). The values were normalized by the IPSC amplitude recorded in the presence of NBQX and d-APV. *D*, Probability distribution of the strychnine-sensitive fraction of IPSCs for control neurons ($n = 30$; gray) and neurons preincubated with ORG25543 ($n = 26$; gray line). *E*, Bar graph of the absolute amplitude of glycinergic and GABAergic IPSCs recorded from control neurons ($n = 30$; gray) and neurons preincubated with ORG25543 ($n = 26$; white). *F*, Bar graph of the amplitude-weighted decay time constant of the mixed GABA/glycine ($p < 0.001$), pure glycinergic ($p < 0.02$), and pure GABAergic ($p < 0.03$) IPSCs for control neurons ($n = 30$; gray) and neurons preincubated with ORG25543 ($n = 26$; white).

treated neurons (24.1 ± 3.8 vs 25.8 ± 3.3 arbitrary units, respectively; $p > 0.5$), indicating similar rates of vesicle endocytosis during the loading phase. During repetitive high-frequency stimulation (HFS) (10 Hz for 1 min), rapid destaining of fluorescence-labeled vesicles was observed. This destaining was not detected in the presence of TTX, indicating that it was dependent on action potentials (Fig. 3G). Similar reductions in fluorescence were observed in response to HFS at individual boutons of control ($59 \pm 6\%$; $n = 6$) and ORG25543-treated ($56 \pm 4\%$; $n = 6$; $p > 0.4$) neurons.

These results strongly suggest that preincubation with ORG25543 reduced glycinergic quantal size and apparent quantal number via a reduction of vesicle filling. This effect appeared to be specific, because ORG25543 had no (or the opposite) effect on glutamatergic and GABAergic mPSCs and did not detectably alter vesicular endocytosis or exocytosis.

Restoration of mIPSCs by GlyT2-mediated glycine uptake

It seemed probable that GlyT2 inhibition acted on vesicular glycine content via a diminution of the cytosolic glycine concentration. This mechanism predicts that resupplying glycine to the transporter might restore mIPSCs. We tested this prediction by applying $100 \mu\text{M}$ glycine for 10 min (in the presence of strychnine to limit glycine receptor-mediated chloride fluxes) (Fig. 5A). In control neurons, the average amplitude of mIPSCs was the same before and after glycine application (-45.3 ± 3.2 and -45.2 ± 4.4 pA, respectively; $n = 16$). In contrast, we observed a remarkable restoration of mIPSC activity in ORG25543-treated neurons

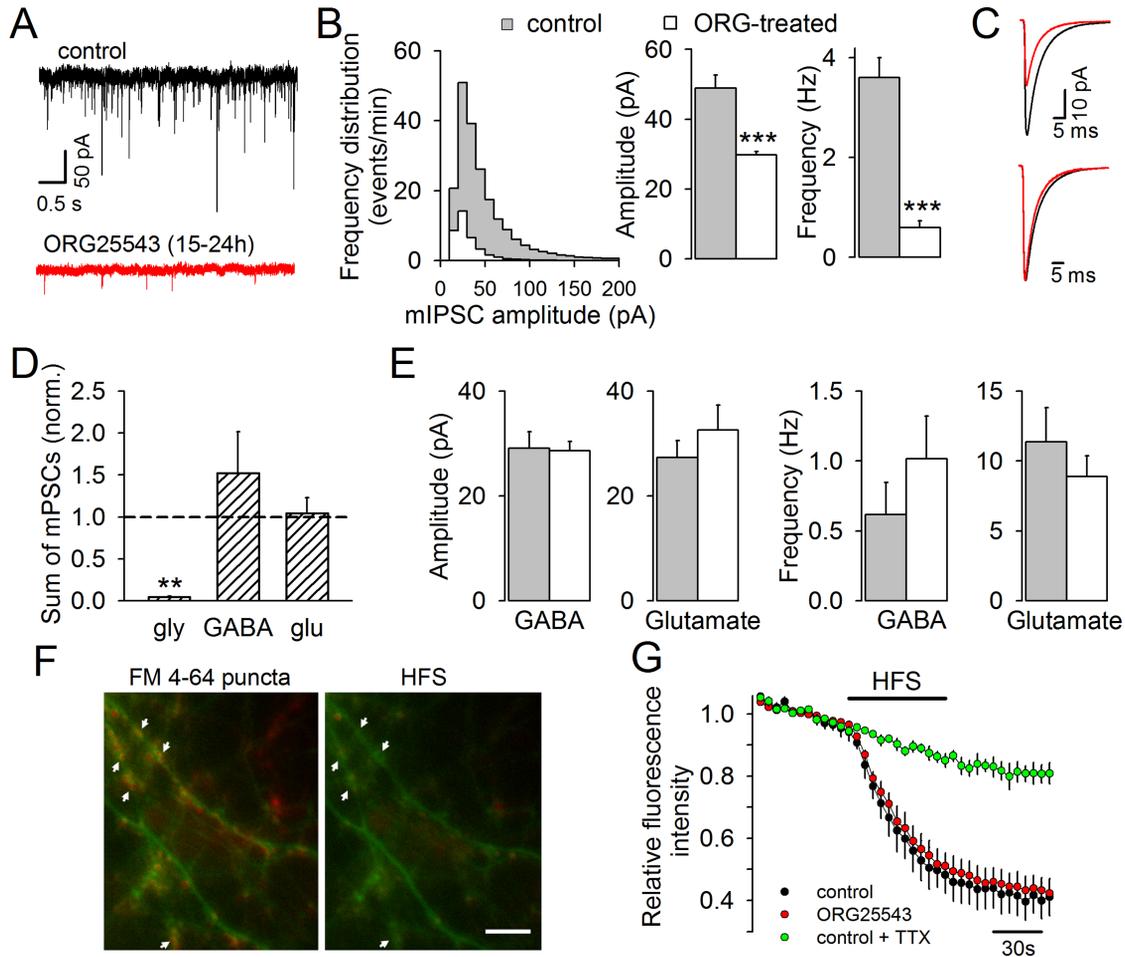


Figure 3. Chronic GlyT2 inhibition reduces glycine quantal size. **A**, Traces of glycinergic mIPSCs recorded from a control neuron (top trace, solid line) and from a neuron preincubated with 5 μM ORG25543 for 15–24 h (bottom trace, red line). **B**, Left, Distributions of mIPSC amplitude in control neurons (gray; $n = 41$) and in ORG25543-treated neurons (white; $n = 72$; $p < 0.001$). Right, Bar graphs of the amplitude and frequency of mIPSCs recorded in control neurons (gray; $n = 41$) and in ORG25543-treated neurons (white; $n = 57$ for amplitude and $n = 72$ for frequency; $p < 0.001$). **C**, Raw (top traces) and normalized (bottom traces) average mIPSCs recorded in control neurons ($n = 41$; solid line) and ORG25543-treated neurons ($n = 57$; red line). **D**, Bar graph of the sum of amplitudes per minute of glycinergic, GABAergic, and glutamatergic mIPSCs recorded in the presence of NBQX + D-APV + SR95531, NBQX + D-APV + strychnine, and SR95531 + strychnine, respectively, for neurons preincubated with ORG25543 ($n = 11$). Data are normalized to values obtained with control neurons ($n = 9$). **E**, Bar graphs of the amplitude and frequency of GABAergic and glutamatergic mIPSCs recorded in control neurons (gray; $n = 11$) and in ORG25543-treated neurons (white; $n = 9$; $p > 0.05$). **F**, Left, Visualization of synaptic boutons (arrows) in GlyT2⁺ neurons after FM 4-64 loading. Right, Destaining of FM 4-64 fluorescence after HFS (10 Hz, 1 min). Scale bar, 10 μm . **G**, Time course of the change in FM 4-64 fluorescence induced by HFS in control neurons (filled circle; 130–190 boutons/coverlips; $n = 6$) and neurons preincubated with 5 μM ORG25543 (red circle; 110–220 boutons/coverlips; $n = 6$; $p > 0.1$). HFS did not induce a change in FM 4-64 fluorescence in the presence of TTX (green circle; 110–150 boutons/coverlips; $n = 4$).

(Fig. 5A–C). The amplitude of mIPSCs returned rapidly to the control level (Fig. 5B), from -28.2 ± 1.2 to -42.8 ± 4.0 pA ($n = 20$), with very similar cumulative distributions (Fig. 5C, $p > 0.5$). The relative change in mIPSCs amplitude, frequency, and time constant induced by glycine is shown in more detail for individual cells in supplemental Figure 1 (available at www.jneurosci.org as supplemental material).

This almost complete restoration of transmission further strengthens the conclusion that GlyT2 acts exclusively via control of intracellular glycine, which in turn regulates vesicular filling. To check the specificity of the restoration for glycine transmission and to exclude any role for the small tonic activation of GlyRs during glycine application, glycine (100 μM) was replaced by β -alanine or taurine (200 μM), both GlyR agonists (Lynch, 2004) that are weak (β -alanine) or not (taurine) substrates of GlyT2 (supplemental Fig. 2, available at www.jneurosci.org as supplemental material). When applied in the presence of strychnine (3 μM), β -alanine and taurine evoked a small and noisy GlyR-mediated current (data not shown) similar to that evoked

by glycine but failed to restore mIPSC activity in ORG25543-treated neurons (Fig. 5D, E). Furthermore, when glycine was applied in the presence of ORG25543 (5 μM), the mIPSC activity remained unchanged after complete washout of strychnine (Fig. 5E). The restoration of mIPSCs by glycine displayed the expected pharmacological profile because, of the drugs tested, only glycine is a substrate for both GlyT2 and the vesicular transporter VIAAT at the concentration applied. These results show that ORG25543 pretreatment selectively and specifically reduces glycine accumulation and thus reduces vesicular glycine content. This effect can be fully reversed by the brief uptake of extracellular glycine by GlyT2.

As the amplitude of mIPSCs in control neurons were not potentiated by glycine uptake (Fig. 5B), we examined whether there was a presynaptic change in glycine release by taking advantage of SR95531, a low-affinity competitive antagonist of GlyRs (Wang and Slaughter, 2005; Beato et al., 2007). Inhibition of GlyRs by a competitive antagonist should be sensitive to changes in the amount of glycine released (Clements, 1996) because SR95531

has an estimated mean lifetime occupancy of 300 μ s and should unbind within the timescale of synaptic release (Beato et al., 2007). In outside-out or nucleated patches, SR95531 at 70 μ M, a concentration well below its binding equilibrium constant of 190 μ M (Beato et al., 2007), blocked the current evoked by 60 μ M glycine by $61 \pm 11\%$ ($n = 3$), whereas the current evoked by 200 μ M glycine was reduced only by $10.6 \pm 9.5\%$ (data not shown). To mimic fast synaptic release, we recorded currents evoked by brief 1 ms iontophoretic applications of glycine and confirmed that the fraction of the current blocked by 70 μ M SR95531 decreased as the amount of glycine release increased (Fig. 6A). Figure 6B shows that 70 μ M SR95531 reduced the amplitude of glycinergic mIPSCs by $28.6 \pm 2\%$ ($n = 6$) (Fig. 6B, C) and that this block remained unchanged ($29.9 \pm 1\%$; $n = 6$) (Fig. 6D, E) after glycine application (100 μ M for 10 min in the presence of 3 μ M strychnine), suggesting that there was no significant change in glycine released after GlyT2 uptake in control neurons.

Acute depletion of intracellular glycine and its reversal

To deplete the intracellular storage of glycine, the ORG25543 preincubation protocol described above lasted 15–24 h. We wanted to test whether acute manipulation of intracellular glycine levels could induce similar effects. We tested this by exploiting the dialysis of the cytoplasm by the patch pipette. In the absence of intracellular or extracellular glycine, the evoked synaptic response ran down during continuous stimulation at 1 Hz over a period of 30 min (Fig. 7A) [$87 \pm 3\%$ reduction ($n = 20$) compared with a basal rundown of $21.6 \pm 11.6\%$ ($n = 4$) in the absence of repeated stimulation]. The time course of this rundown was quite variable between cells, ranging from complete disappearance in 15 min to incomplete inhibition after 60 min (Fig. 7C) ($t_{80-20} = 28 \pm 4$ min; $n = 20$). A sharp acceleration in the time course was present after ~ 20 min of stimulation in 7 of 20 pairs, suggesting that the dialysis of glycine in the axon terminals became effective only after a long latency (Mathias et al., 1990).

Once the rundown had occurred, the response did not recover even if the stimulation frequency was reduced to a very low rate (0.1 Hz; the frequency dependence of the transmission is investigated below). However, application of 100 μ M glycine (in the presence of strychnine) induced a strong recovery of the evoked response to $77 \pm 13\%$ of the initial amplitude ($n = 10$) (Fig. 7B). The time course of the recovery was masked by the presence and long wash out of strychnine. Surprisingly, the IPSC decay time constant increased from 12.3 ± 2.0 ms ($n = 10$) to 19.9 ± 3.1 ms ($p < 0.05$) after recovery after glycine. Moreover, preventing the loss of glycine by addition of a high concentration of glycine in the intracellular solution limited the run down to $57 \pm 8\%$ of the

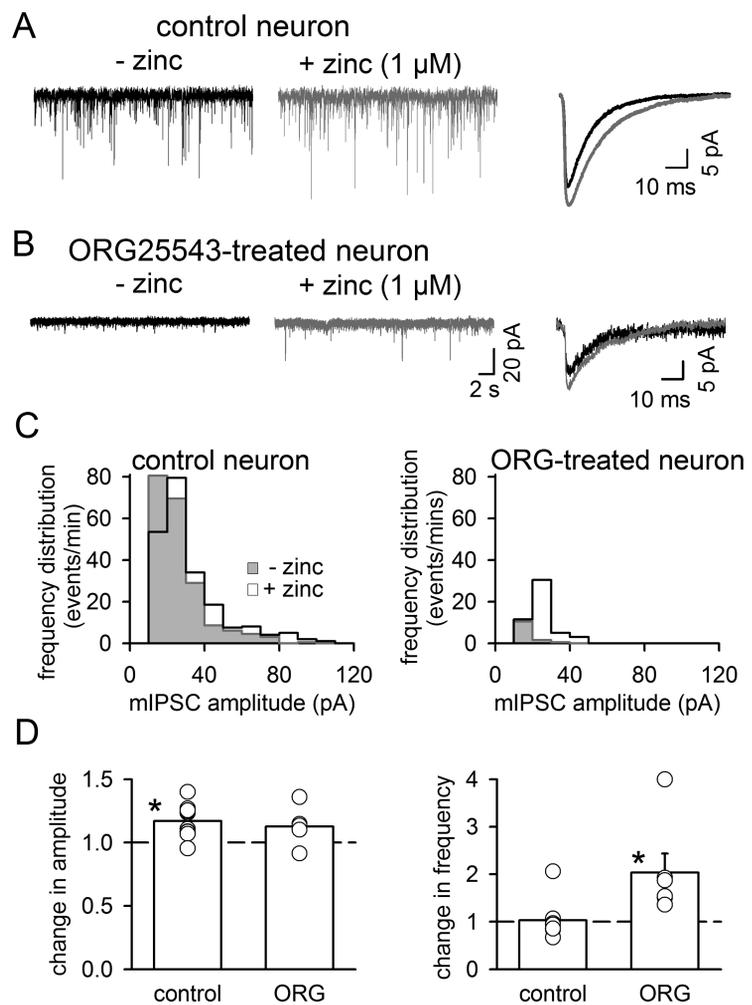


Figure 4. Zinc unmasks a small population of vesicles containing trace levels of glycine in ORG25543-treated neurons. Glycinergic mIPSCs were recorded from control (A) and ORG25543-treated (B) neurons, in the absence (solid) or presence (gray) of 1 μ M free zinc. To obtain 1 μ M free zinc, 200 μ M ZnCl₂ was added to the external solution containing 10 mM tricine (Paoletti et al., 1997; Fayyazuddin et al., 2000). C, D, Free zinc at 1 μ M slightly increased the mean glycinergic mIPSC peak amplitude in control and ORG25543-treated neurons ($n = 10$ for control neurons, $p < 0.5$; $n = 6$ for ORG-treated neurons; $p > 0.5$). Zinc had no net effect on mIPSC frequency in control neurons ($n = 10$; $p > 0.5$) but always increased the mIPSC frequency in ORG25543-treated neurons ($n = 6$; $p < 0.5$), suggesting that small events were below the detection threshold.

initial amplitude ($n = 4$; $p < 0.02$) after 30 min of dialysis (Fig. 7C).

Therefore, it is possible to acutely deplete and restore intracellular glycine levels, and these effects are consistent with those produced by chronic GlyT2 inhibition (Figs. 2, 3).

Glycine supply during high-frequency activity

A crucial role of glycine uptake is to supply neurotransmitter to fill newly formed vesicles and refill those recovered by endocytosis. To investigate the role of GlyT2 in this process, we recorded glycinergic IPSCs evoked by high-frequency stimulation and identified two phases of stationary release. Figure 8A–C shows the traces of IPSCs from a paired recording that was stimulated with 1000 APs at 5 Hz. IPSCs recorded during initial and late phase of the stimulation had the same kinetics, but the amplitude of late IPSCs was reduced (Fig. 8C). The average time course of IPSC amplitude during stimulation is shown in more detail in Figure 8D, using a logarithmic timescale. The very first IPSC (I_1) of the train was always larger than subsequent IPSCs and was ignored because its “frequency” is 0. Subsequent IPSCs show an

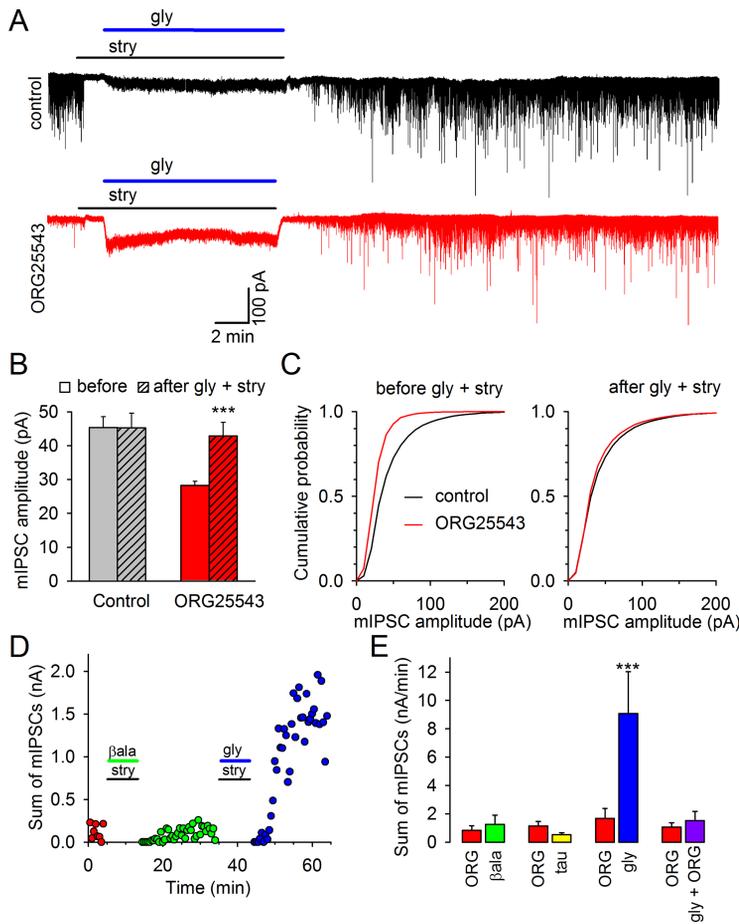


Figure 5. Glycine uptake by GlyT2 restores glycinergic mIPSCs. **A**, Representative traces of glycinergic mIPSCs recorded in control neurons (top, black) and in neurons preincubated with 5 μM ORG25543 for 12–24 h (bottom, red). Glycine (100 μM ; blue line) was applied in the presence of strychnine (3 μM ; solid line; stry). The average tonic GlyR current evoked by glycine was comparable for control and ORG25543-treated neurons (-37.9 ± 4.1 , $n = 10$ and -40.3 ± 4.6 pA, $n = 17$, respectively). **B**, Bar graphs of the mean amplitude of glycinergic mIPSCs from control neurons (gray bars; $n = 16$) and ORG25543-treated neurons (red bars; $n = 20$), before (solid bars) and after (striped bars) glycine application with strychnine (3, 10, or 30 μM , all data were pooled). **C**, Cumulative probability of mIPSC amplitude distribution recorded from control neurons (black line; $n = 16$) and from glycinergic neurons preincubated with 5 μM ORG25543 for 15–24 h (red line; $n = 20$) before (left; $p < 0.001$) and after (right; $p > 0.5$) glycine application. **D**, Time course of the sum of mIPSC peak amplitudes per 30 s recorded from a neuron preincubated with 5 μM ORG25543 for 15–24 h. β -Alanine (200 μM) and then glycine (100 μM) were applied in the presence of strychnine (3 μM). **E**, Sum of glycinergic mIPSC peak amplitudes per minute in ORG25543-treated neurons (red) after application of β -alanine (200 μM ; green; $n = 7$), taurine (300 μM ; yellow; $n = 4$), glycine (100 μM ; blue; $n = 23$; $p < 0.001$), and glycine (100 μM) + ORG25543 (5 μM) (purple; $n = 8$).

initial plateau (Fig. 8D) ($I_p/I_1 = 0.62 \pm 0.01$; $n = 18$), typically between APs 3 and 40, and then a rundown to a low steady-state (tonic) level ($I_{ss}/I_1 = 0.13 \pm 0.02$). In contrast, the IPSC amplitude remained stable during the same period when stimulated at low frequency (0.1 Hz) (Fig. 8D, triangles). The reduced response in the tonic phase of 5 Hz stimulation could be caused by many presynaptic and/or postsynaptic mechanisms underlying short-term plasticity (Foster and Regehr, 2004; Xu-Friedman and Regehr, 2004): lower release probability, decrease in the pool of releasable vesicle, lower vesicular glycine content, or GlyR desensitization (Rigo and Legendre, 2006). Nevertheless, we were interested to examine whether the rate of vesicle refilling with glycine played a role during tonic release. In principle, some information about the locus of the synaptic depression can be obtained from the coefficient of variation method by comparing initial and late release events (Faber and Korn, 1991; Foster and Regehr, 2004). The average reduction in IPSC amplitude was

inversely proportional to the variation in CV^2 (Fig. 8E), thus arguing against a uniform reduction of quantal size and hence against uniform desensitization or reduction of glycine content being the major cause for the reduction in amplitude during high-frequency stimulation. Directly sampling quanta by recording asynchronous IPSCs (aIPSCs) using strontium showed a reduction in aIPSC amplitude between initial and late release, from -58.6 ± 9.6 to -38.7 ± 4.3 pA ($n = 5$; $p > 0.05$) (supplemental Fig. 3, available at www.jneurosci.org as supplemental material), but this reduction cannot account for the extent of synaptic depression. We did not investigate further other mechanisms, except to exclude a reduction in release probability by the activation of presynaptic GABA_B receptors by coreleased GABA (Lim et al., 2000), because similar synaptic depressions were recorded in the presence of CGP55845 (0.25 and 1 μM) (supplemental Fig. 4, available at www.jneurosci.org as supplemental material), a specific GABA_B receptor antagonist (Blake et al., 1993).

In contrast to the slow rundown produced by whole-cell dialysis (Fig. 7), synaptic depressions evoked by high-frequency stimulation were reversible. Figure 8F shows the amplitude of IPSCs from a paired recording in which 0.1 and 5 Hz stimulation periods were interleaved. The low-frequency responses (Fig. 8F, triangles) chart the progressive depletion of intracellular glycine. The decrease in cumulative IPSC amplitude in four consecutive trials showed a progressive decline in initial and late tonic responses (Fig. 8G). Because both initial and late responses might include the release of slowly recruited and/or recycled vesicles, their co-decline suggests the capacity to refill recycled vesicles with glycine decreased as the rundown progressed.

Next, we examined whether GlyT2-mediated uptake could restore high-frequency release in glycine-depleted neurons. To do so, two trains of 1000 APs at 5 Hz were evoked before and after glycine application in dialyzed and ORG25543-treated neurons, our two models of glycine depletion. Figure 9A shows the time course of IPSC amplitude in a control neuron that is being dialyzed. In this cell, the slow rundown took 3000 APs evoked at 1 Hz (the average IPSC amplitude was computed every minute in Fig. 9A, gray circles). Subsequent application of glycine effectively potentiated the responses evoked by low-frequency stimulation (Fig. 9A, triangles), whereas the late phase of release was not restored during the second 5 Hz stimulation ($I_{ss}/I_p = 0.018 \pm 0.007$; $n = 3$) (Fig. 9A,B). Indeed, the cumulative amplitude of the second stimulation approached saturation after ~ 200 APs (Fig. 9C, green line), in contrast to the continuous rise observed during the first train (Fig. 9C, blue line). A similar result was obtained using ORG25543-treated cells that were already

glycine-depleted at the onset of the experiment (Fig. 9D,E,H). Once again, glycine uptake strongly potentiated the response evoked at low frequency and during the initial phase of high-frequency stimulation (Fig. 9D,E,G,I) but failed to potentiate the late tonic phase ($I_{ss}/I_p = 0.059 \pm 0.037$; $n = 4$) as shown by the saturating value of cumulative amplitude (Fig. 9F, green line).

The failure of GlyT2-mediated uptake to restore tonic synaptic release suggested that glycine did not reach the cytosolic concentration needed for efficient VIAAT loading, thus reducing the rate of vesicle refilling. To test this hypothesis, similar experiments were repeated with a presynaptic intracellular solution containing high concentrations of glycine (10 or 20 mM). Figure 10A shows a trace of IPSCs evoked by 5 Hz stimulation in a neuron dialyzed with a pipette solution containing 10 mM glycine. Although a synaptic depression was still present during the high-frequency stimulation, the mean amplitude of the last 200 IPSCs was potentiated by the addition of glycine in the pipette solution (Fig. 10A–C), from $I_{ss}/I_p = 0.18 \pm 0.03$ ($n = 16$) in the absence of glycine to 0.34 ± 0.07 ($n = 7$; $p < 0.05$) and 0.52 ± 0.07 ($n = 4$; $p < 0.01$) in the presence of 10 and 20 mM glycine, respectively (Fig. 10C). The cumulative amplitudes of IPSCs during 5 Hz stimulation (normalized to the initial amplitude) plotted in Figure 10D allow a direct comparison of the five different conditions of glycine supply described above. The slopes of cumulative amplitude during the tonic phase of release (after ~200 APs) rise with increasing glycine concentration (Fig. 10D, solid lines), suggesting that VIAAT kinetics limits the refilling of vesicles with glycine. Therefore, the absence of tonic release in dialyzed and ORG25543-treated neurons after short-term glycine uptake (Fig. 10D, dashed lines) may be explained by a lower cytosolic glycine concentration and thus lower rate of vesicles refilling.

Finally, we examined whether the uptake of glycine by GlyT2, at a concentration (5 μ M) near physiological levels, can support vesicle refilling. As shown previously in Figure 9D, the initial pool of glycinergic vesicles was exhausted in ORG25543-treated cells after high-frequency stimulation (Fig. 11A). Then the amplitude of IPSCs were monitored at low frequency (0.1 Hz) in the presence or absence of extracellular glycine (5 μ M) (Fig. 11A). Because 5 μ M glycine evoked only a small tonic current (-49.2 ± 11 pA; $n = 7$), addition of strychnine was not necessary, thus unmasking the coupling between uptake and release. During glycine application, the amplitude of IPSCs rose slowly, reflecting the growth in the pool of vesicles refilled with glycine (Fig. 11A). We estimated that IPSC amplitude increased by $63 \pm 10\%$ ($n = 7$) after 20 min of 5 μ M glycine application (Fig. 11C). Surprisingly, the washout of glycine in five of seven pairs evoked a monotonic decline of IPSC amplitude, indicating a direct coupling between GlyT2-mediated uptake and vesicles refilled with glycine (Fig.

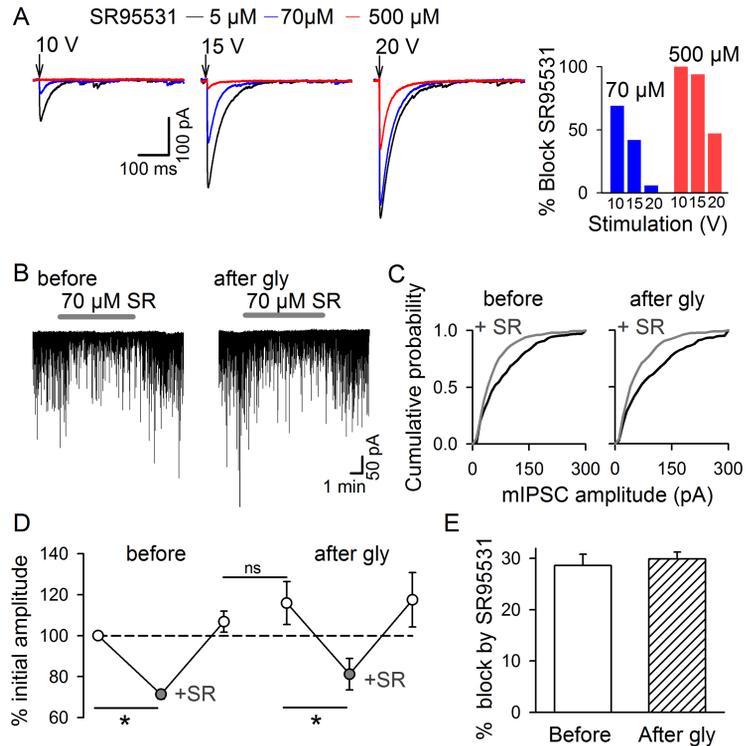


Figure 6. Glycine release is not changed after GlyT2 uptake in control neurons. **A**, SR95531 (70 and 500 μ M) inhibits fast iontophoretically evoked glycine currents. Left, Average current traces evoked by 1 ms pulses (10, 15, or 20 V) of iontophoretically applied glycine in extracellular solution containing 5 μ M (solid line), 70 μ M (blue line), or 500 μ M (red line) SR95531. Right, Percentage block by SR95531 for the three voltage intensities shown in **A**. **B**, Representative traces of glycinergic mIPSCs recorded from control neurons before (left) and after (right) glycine application (100 μ M for 10 min in the presence of strychnine at 3 μ M). In both conditions, addition of 70 μ M SR95531 (gray line) induced a reduction in glycinergic mIPSC amplitude. **C**, Cumulative probability of mIPSC amplitude distribution recorded from the same neuron as in Figure 5A before (left) and after (right) glycine application ($p > 0.05$) in control (solid line) and in the presence of 70 μ M SR95531 (gray line; SR; $p < 0.05$). **D**, Average change in mIPSC amplitude recorded from six neurons before and after glycine application ($p > 0.05$), in control (open circle) and in the presence of 70 μ M SR95531 (gray circle; $p < 0.05$). **E**, Percentage of glycinergic mIPSC amplitude blocked by 70 μ M SR95531 before (open bar) and after (dashed bar; $p > 0.05$) glycine application.

11A). In one tested cell, subsequent addition of glycine restored the transmission (Fig. 11A).

The time course of the IPSC was slower after glycine application (Fig. 11B) as shown previously in Figure 9G. In both experiments using ORG25543-treated cells, the decay time constant increased from 6.9 ± 0.7 ms at the beginning of the recording to 15.8 ± 3.8 ms ($n = 10$; $p < 0.05$) after recovery by glycine uptake. A hypothesis for this kinetic change will be discussed below.

Discussion

GlyT2 specifies the neuronal glycinergic phenotype

In this study, we took advantage of transgenic GlyT2–EGFP mice (Zeilhofer et al., 2005) to record evoked currents in pairs of connected spinal cord neurons with an identified GlyT2⁺ presynaptic element. Half of the neurons were fluorescent in the cultures, in good agreement with the 50% distribution of neurons expressing GlyT2 mRNA in laminae IV–VIII of the rat spinal cord (Hossaini et al., 2007). The probability of ectopic expression was reduced by using heterozygous GlyT2–EGFP mice (Zeilhofer et al., 2005), and we confirmed the membrane expression of GlyT2 by the observation that glycine uptake currents were only recorded in fluorescent neurons.

No strychnine-sensitive IPSCs were evoked in GlyT2[−] neuron pairs, and all GlyT2⁺ neurons were glycinergic, confirming that GlyT2 expression is therefore necessary and sufficient to

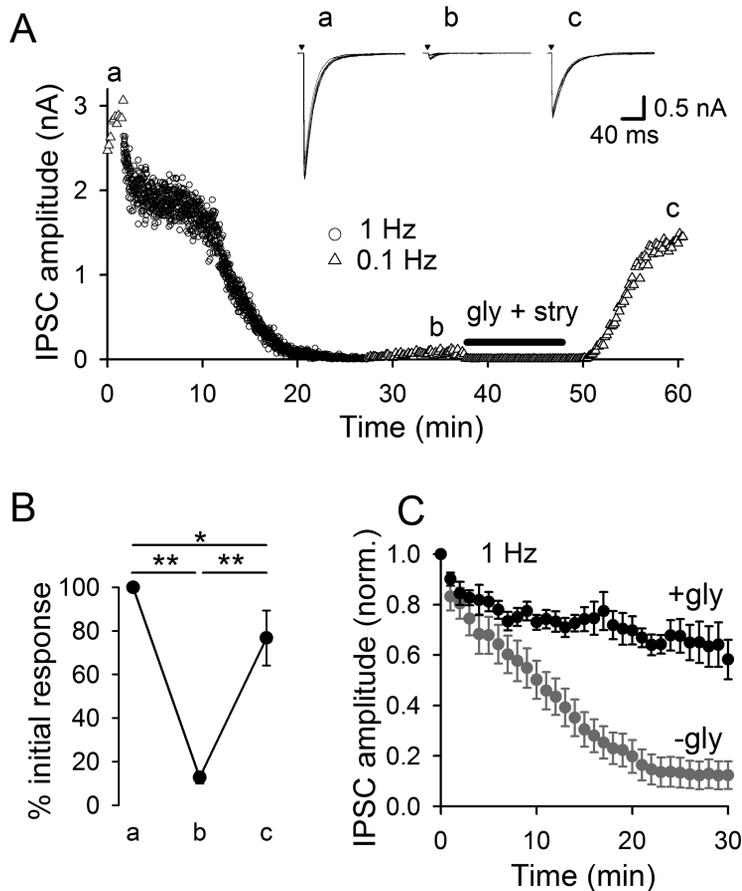


Figure 7. Acute depletion and refilling of glycine-containing vesicles by GlyT2-mediated uptake. **A**, Time course of the depression of glycinergic IPSCs recorded from a pair with a presynaptic GlyT2⁺ neuron during repeated stimulation at 1 Hz (open circles). After near complete rundown of the transmission, the stimulation frequency was lowered to 0.1 Hz (open triangles) and glycine (100 μ M; solid line) was applied for 10 min in the presence of strychnine (10 μ M). Inset, Current traces recorded for 10 consecutive IPSCs before 1 Hz stimulation (**a**), after the rundown (**b**), and after recovery with glycine (**c**). **B**, Average change in IPSC amplitude recorded in 10 pairs after complete rundown (**b**) and after glycine application (**c**). The responses are expressed as percentage of their initial amplitude recorded in **a**. **C**, Time course of glycinergic IPSC rundown during repeated stimulation at 1 Hz when the presynaptic GlyT2⁺ neuron is dialyzed without (gray; $n = 6$) or with (black circles) glycine [$p < 0.02$; data from 10 mM ($n = 2$) and 20 mM ($n = 2$) glycine were pooled] in the pipette solution.

determine a glycinergic phenotype (Jursky and Nelson, 1995; Zafra et al., 1995; Zeilhofer et al., 2005). However, the neuronal glycinergic phenotype was always associated with a GABAergic component. These mixed phenotypes are expected from embryonic spinal cord inhibitory neurons because expression of GlyT2 overlaps with that of glutamate decarboxylase (GAD) during the development of spinal cord and brainstem nuclei (Geiman et al., 2002; Mackie et al., 2003; Tanaka and Ezure, 2004). Furthermore, corelease of GABA and glycine is the main mode of immature inhibitory transmission in these structures (Jonas et al., 1998; Chéry and de Koninck, 1999; O'Brien and Berger, 1999; Russier et al., 2002). Considering that the EC_{50} of VIAAT for GABA transport is lower than for glycine (McIntire et al., 1997; Bedet et al., 2000), we interpret the dominance of glycinergic transmission in the majority of the synapses made by GlyT2⁺ neurons as reflecting a much higher supply of glycine (by uptake) than of GABA (by synthesis). Nevertheless, part of this glycinergic dominance could be also accounted for by differences in the relative expression or stabilization of the GlyR and GABA_AR in postsynaptic clusters (Todd et al., 1996), as shown in the cerebellum for the two targets of the Golgi cell (Dugué et al., 2005).

Blocking neuronal glycine uptake for several hours markedly

reduced glycinergic transmission in culture, as in slices (Gomez et al., 2003; Bradaia et al., 2004), but enhanced GABAergic transmission. The combination of these changes was sufficient to shift the neuron phenotype from mostly glycinergic to mostly GABAergic (Fig. 2D), thus illustrating the pivotal role of GlyT2 for the developmentally regulated switch of inhibitory phenotypes detected in many brainstem and spinal cord structures (Kotak et al., 1998; Gao et al., 2001; Keller et al., 2001). For example, in the lateral superior olive (LSO), a brainstem auditory nucleus, a transition from GABAergic to glycinergic transmission occurs during the second postnatal week (Kotak et al., 1998; Awatramani et al., 2005) that is associated with the formation of punctuate clusters of GlyT2 around LSO neurons (Zafra et al., 1995; Friauf et al., 1999) and a downregulation of glutamate decarboxylase (Nabekura et al., 2004). Interestingly, the peak of GlyT2 expression in the LSO is observed at postnatal day 10, near the onset of the shift (Kotak et al., 1998; Friauf et al., 1999; Awatramani et al., 2005), and then decreases, presumably because of lower competition of glycine with GABA (Nabekura et al., 2004).

Glycine and GABA content in synaptic vesicles

The experiments with FM 4-64 dye indicated that the recycling of synaptic vesicles was not impaired in ORG25543-treated neurons, thus confirming that recycling and refilling of vesicles are not tightly coupled (Edwards, 2007). Vesicles were not expected to be empty in ORG25543-treated neurons because glycine and GABA are thought to substitute for one another when their relative concentrations are changed, as both amino acids are coaccumulated in the same VIAAT-containing vesicles (Wojcik et al., 2006; Aubrey et al., 2007). However, restarting GlyT2-mediated glycine uptake in neurons treated overnight with ORG25543 restored miniature and evoked glycinergic IPSC amplitude within 10 min, suggesting that glycine-free vesicles were not completely filled with GABA, because they could be filled to control levels with glycine when it became available. Alternatively, glycine may replace GABA in vesicles if VIAAT mediates rapid exchange of the two amino acid neurotransmitters.

Manipulations that augment GABA supply in brain slices, by either reducing its degradation (Engel et al., 2001) or providing glutamate for GABA synthesis (Mathews and Diamond, 2003), increase the amplitude of GABAergic mIPSCs. In contrast, we show that glycine application did not potentiate the amplitude of glycinergic mIPSCs in control neurons and restored only to the level of control neurons in glycine-depleted ORG25543-treated neurons (Fig. 5C). These results can be explained by a saturation of postsynaptic GlyRs or alternatively by a saturation in the amount of glycine that can be stored in vesicles. In agreement with previous studies (Suwa et al., 2001; Laube, 2002; Rigo et al.,

2003; Hirzel et al., 2006), we provide evidence that GlyRs are not saturated in our preparation because the mIPSCs were potentiated by 1 μM zinc, a high-affinity allosteric modulator that increases the receptor affinity for glycine (Bloomenthal et al., 1994; Laube et al., 1995, 2000; Suwa et al., 2001; Miller et al., 2005). The fraction of the current blocked by SR95531 did not change after glycine uptake, suggesting that increasing glycine availability does not increase vesicular accumulation of glycine in control neurons.

Although glycine and GABA transport by VIAAT are presumably coupled to the same exchange with 1 H^+ (Burger et al., 1991; Edwards, 2007), their accumulation in synaptic vesicles may not be limited by the same thermodynamic or kinetic constraints if their cytosolic concentrations differ significantly. Glycine content measured in mice spinal cord tissues ($\sim 4 \mu\text{mol}/\text{mg}$ protein) (Daly, 1990) is approximately two orders of magnitude larger than GABA content measured in the cortex of adult mice ($\sim 15 \text{ nmol}/\text{mg}$ protein) (Asada et al., 1997). Furthermore, glycine loading during vesicle reacidification might be faster than for GABA because the recovery of glycinergic mIPSCs preceded GABAergic mIPSCs after transient inhibition of the vesicular H^+ -ATPase by bafilomycin (Katsurabayashi et al., 2004). Other factors may also regulate the relative content of glycine and GABA. For example, an association of VIAAT with GAD65 has been shown to provide a major kinetic advantage for the vesicular accumulation of newly synthesized GABA (Jin et al., 2003). Considering that synaptic vesicles contain up to 10 transporters (Takamori et al., 2006), their association or not with GAD65 is likely to alter the relative accumulation of glycine and GABA in vesicles (Katsurabayashi et al., 2004).

GlyT2 operation during vesicle refilling

Because vesicles are recycled locally, neurotransmitter must also be supplied locally, and our results establish the capacity of GlyT2 to drive the filling of synaptic vesicles by uptake of glycine at low extracellular concentration (Fig. 11). This result extends genetic studies in knock-out *GlyT2*^{-/-} mice and hyperekplexia patients (Gomez et al., 2003; Rees et al., 2006) that established GlyT2 as a critical presynaptic determinant of glycinergic transmission. The punctuate expression of GlyT2 around presynaptic boutons (Zafra et al., 1995; Arnsen et al., 2007) together with its 3 Na^+ coupling and unidirectionality (Roux and Supplisson, 2000) combine to produce a local accumulation of glycine at the synapse.

GlyT2-mediated glycine uptake has been shown to be coupled to the transfer of 2.1 charges/glycine in recombinant system (Roux and Supplisson, 2000). Using this proportionality, the av-

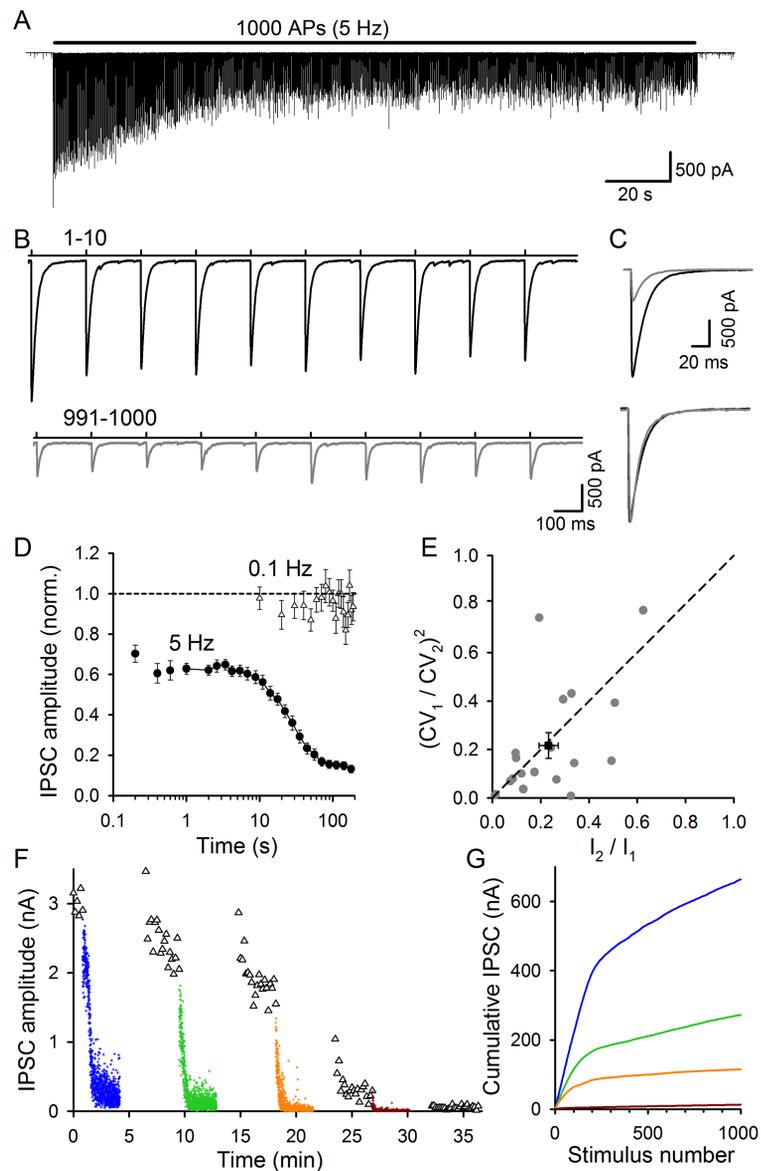


Figure 8. Different phases of glycine release during high-frequency stimulation. **A**, Trace of IPSCs evoked by 1000 APs triggered at 5 Hz. **B**, Trace of IPSCs recorded at the onset (top traces; APs 1–10) and at the end (bottom traces; APs 991–1000) of 5 Hz stimulation. **C**, Averaged (top traces) and peaked scaled (bottom traces) IPSCs shown in Figure 7B. **D**, Time course of the absolute peak amplitude of IPSCs evoked by 5 Hz stimulation [IPSC were normalized to the first IPSC and averaged using a constant logarithmic bin (0.1); $n = 18$]. **E**, Change in CV^2 plotted as a function of steady-state change in IPSC peak amplitude (see Materials and Methods). Individual pairs are represented with gray circles ($n = 16$), and the black square represents the mean \pm SEM. The dashed line represents a slope of 1. **F**, Time course of the amplitude of IPSCs recorded during four successive 5 Hz trains (crosses, depicted in different colors). After 2 min recovery, IPSCs were evoked at 0.1 Hz (triangles). **G**, Cumulative amplitude of IPSCs recorded in each 5 Hz trial (colors as in F).

erage transporter current recorded in GlyT2⁺ neurons (43 pA) (Fig. 1) is equivalent to a glycine influx of 208×10^{-18} mol/s. Assuming that synaptic vesicles store 1790 molecules of neurotransmitters on average (Takamori et al., 2006), GlyT2-mediated uptake can supply enough glycine to refill $\sim 70\,000$ vesicles/s. This calculated filling rate suggests that, despite their low turnover/rates and their restriction to two-dimensional membranes, transporters can compete favorably with enzyme synthesis as a supply mechanism.

Transient uptake of glycine mediated by GlyT2 restored the amplitude of the evoked response after glycine depletion but not the tonic release during train of high-frequency stimulation. In

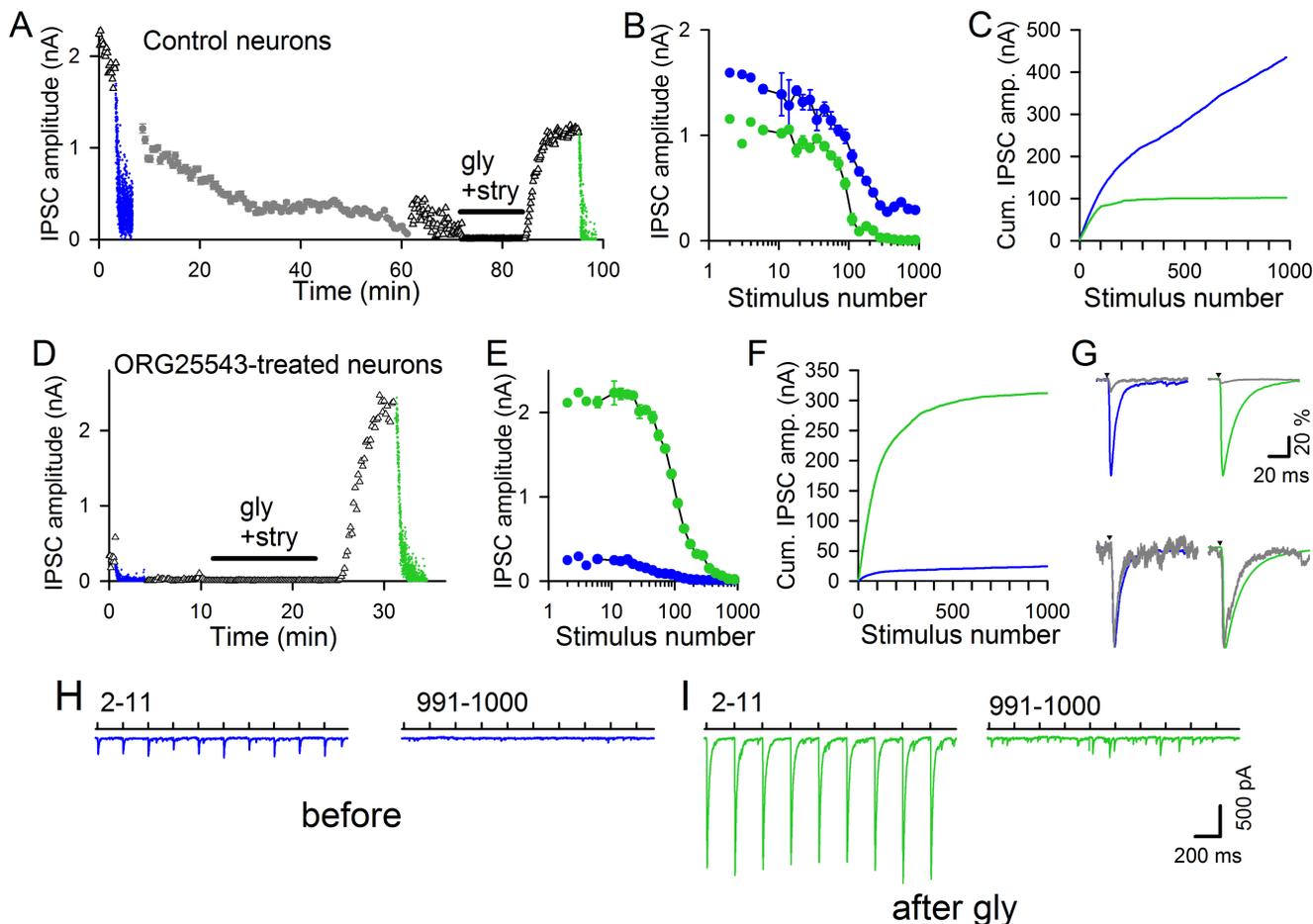


Figure 9. GlyT2-mediated uptake rescued the initial but not the late phase of release during high-frequency stimulation. **A**, Time course of the IPSC absolute amplitude evoked by continuous stimulation at different frequencies: 0.1 Hz (white triangles), 5 Hz (blue or green cross), and 1 Hz (gray circles represent the average amplitude per min). After a train of 1000 APs evoked at 5 Hz (blue crosses), IPSCs were evoked at 1 Hz until a near complete dialysis of the presynaptic neuron was achieved. Then, glycine (100 μ M; solid bar) was applied for 10 min in the presence of strychnine (10 μ M; stry), and, after strychnine washout, a second 5 Hz train of 1000 APs was triggered (green crosses). **B**, Peak amplitude of the IPSCs recorded during the two 5 Hz trains (colors as in **A**). **C**, Cumulative IPSC amplitude of the two 5-Hz stimulations. **D**, Time course of the absolute IPSC peak amplitude recorded in an experiment similar to **A** but using ORG25543-treated neurons. Note that the rundown was complete after the first 5 Hz train (blue crosses). **E**, Peak amplitude of the IPSCs recorded during the two 5-Hz trains (colors as in **D**). **F**, Cumulative IPSC amplitude before (blue line) and after (green line) glycine application. **G**, Raw (top traces) and normalized (bottom traces) average IPSCs recorded during the first (left) and the second (right) trains at the onset (blue or green lines) and at the end (gray line) of the 5 Hz stimulation. **H**, **I**, Traces of 10 consecutive IPSCs recorded at the onset (left) and end (right) of the first (**H**) and second (**I**) 5 Hz stimulation. Solid line represents the time when the APs were triggered.

contrast, high concentrations of glycine in the pipette prevented the rundown induced by dialysis (Fig. 7C) and potentiated the amplitude of evoked IPSCs during tonic release (Fig. 10C). Together, these results suggest that short-term glycine uptake may not be sufficient to restore the high cytosolic concentration of glycine request for rapid vesicles refilling because of VIAAT low affinity.

In our experiments, we consistently found an increase in the decay time of miniature and evoked IPSCs after glycine application. Although we cannot exclude a postsynaptic change of GlyR properties attributable to the application of high concentrations of glycine and strychnine, presynaptic factors might be also involved in this effect. Indeed, it has been shown recently that corelease of GABA, a weak partial agonist of GlyRs, alters the deactivation kinetics of GlyRs and controls the fast decay of glycinergic IPSCs in auditory brainstem nucleus (Lu et al., 2008). In our experiments, the proportion of GABA coreleased with glycine is likely to decrease after glycine application to dialyzed and ORG25543-treated neurons. According to Lu et al. (2008), lowering GABA corelease may alter the kinetics of glycinergic IPSCs

in the direction reported here (Lu et al., 2008), from a fast decay in glycine-depleted cells to slower decay after glycine uptake.

In conclusion, our data provide new evidence supporting the role of GlyT2 for the recycling and refilling of synaptic vesicles in glycinergic neurons. This mechanism takes advantage of the peculiar properties of GlyT2 for generating and maintaining high glycine content in axon terminals. During evolution, these properties may have obviated the need for a dedicated mechanism of enzymatic synthesis of glycine in inhibitory neurons. Furthermore, membrane trafficking of GlyT2 may offer a simple mechanism to adjust precisely glycine supply in the presynaptic terminal.

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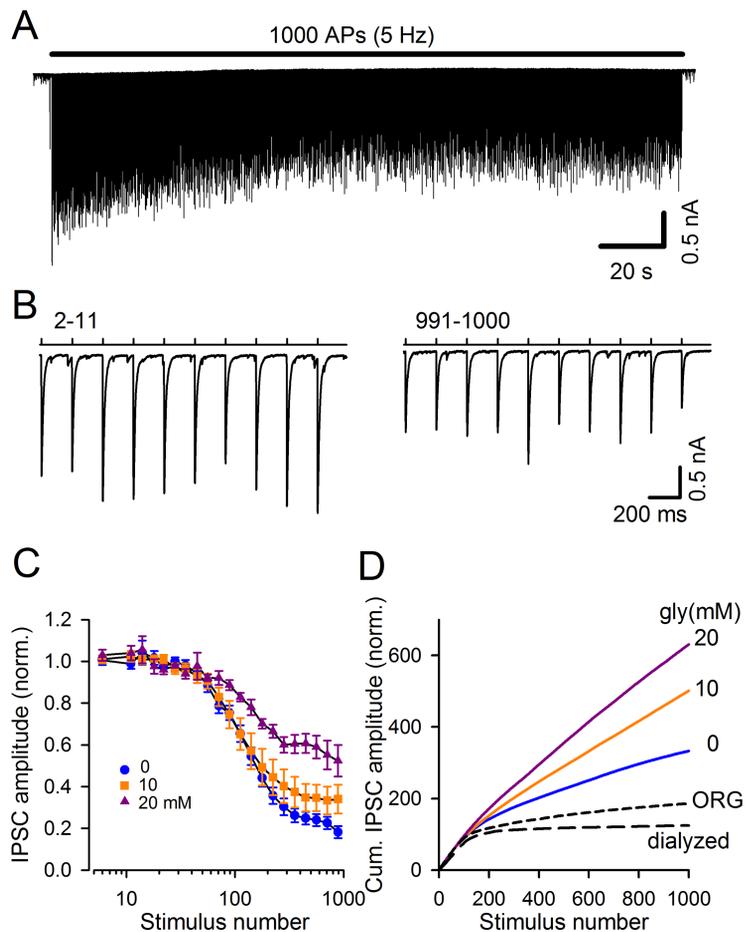


Figure 10. Glycine potentiates the late phase of release. **A**, Representative trace of IPSCs recorded during high-frequency stimulation (1000 APs at 5 Hz) with a GlyT2⁺ presynaptic neuron dialyzed with 10 mM glycine in the pipette. **B**, Traces of 10 consecutive IPSCs recorded at the onset (left traces; APs 1–10) and at the end of the 5 Hz stimulation (right traces; APs 991–1000). The solid line represents the time when the APs were triggered. **C**, Expanded time course of the peak amplitude of glycinergic IPSCs recorded at 5 Hz in pairs with presynaptic GlyT2⁺ neurons dialyzed with 0 (blue circle line; n = 16), 10 (orange square; n = 6), or 20 mM (purple triangle; n = 4) glycine. IPSC amplitudes were normalized to the average amplitude of 30 initial IPSCs (IPSCs 3–32). **D**, Average cumulative IPSC amplitudes during 5 Hz stimulations. IPSC amplitudes of control neurons dialyzed without glycine (blue line; n = 16) or with glycine at 10 mM (orange; n = 7) or 20 mM (purple; n = 4) were normalized to the average amplitude of 30 initial IPSCs (IPSCs 3–32). For comparison, the dashed lines represented the average cumulative IPSC amplitude measured after glycine application to dialyzed (long dashed; n = 3) and ORG25543-treated (short dashed; n = 4) neurons as shown in Figure 9.

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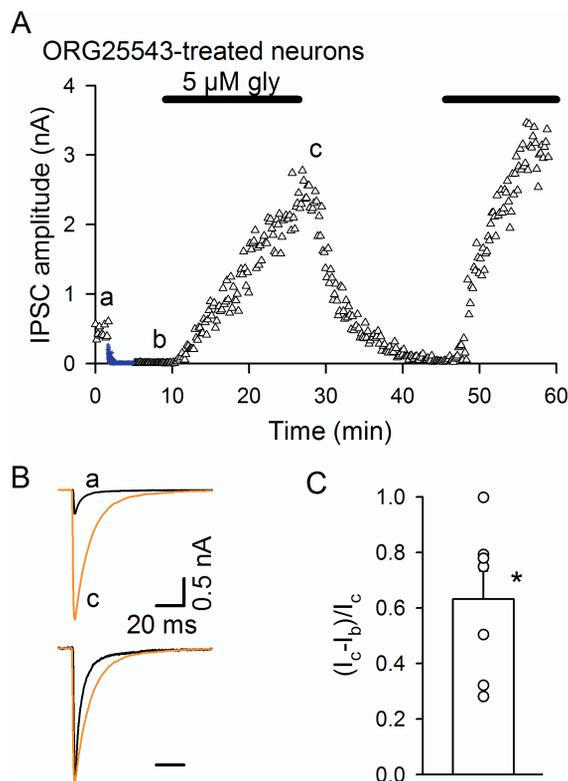


Figure 11. Direct coupling between GlyT2-mediated uptake and vesicle refilling. **A**, Time course of glycinergic IPSC amplitude during continuous stimulation at 0.1 Hz (white triangles) in the absence or presence of 5 μ M extracellular glycine (solid bar) in an ORG25543-treated neuron. As in Figure 9D, an initial 5 Hz stimulation train (1000 APs) was sufficient to induce a complete rundown of the response (blue crosses). **B**, Raw (top traces) and normalized (bottom traces) average IPSCs recorded at the beginning of the trace (a; solid line) and after glycine application (c; orange line). **C**, Relative change in IPSC amplitude during glycine application in ORG25543-treated pairs ($n = 7$; $p < 0.05$). I_b and I_c , Average amplitude of 10 IPSCs before (I_b) and at the end (I_c) of 5 μ M glycine application, as indicated in Figure 11A.

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