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Propofol-induced GABAergic Tonic Inhibition Diminishes α -rhythms and Induces δ -rhythms in neuronal populations

Laure Buhry^{1*} and Axel Hutt¹

¹*INRIA-Nancy Grand Est Research Center
Cortex Team - Building C
615, rue du Jardin Botanique
54600 Villers-ls-Nancy
France*

**Email: laure.buhry@inria.fr*

Abstract

Anaesthetic agents such as propofol are known to have an effect on both synaptic and extra-synaptic receptors. On the one hand, binding of propofol to GABA_A synaptic receptors induces a phasic inhibition, as opposed to tonic inhibition which seems mainly induced by binding to extra-synaptic receptors. On the second hand, under anaesthesia, an increase in amount of slow oscillations, mainly δ (0-4Hz), concurrent to a decrease of α oscillations (8-12Hz), is observed in EEG recordings of occipital areas in most mammals including humans. The latter observation cannot be explained by sole phasic inhibition. Therefore, we propose to investigate, through numerical simulations, the role of tonic inhibition in the increase (in amount) of slow oscillations under propofol anaesthesia. To account for the biological realism of our simulations, the cortical model includes two neuronal populations, one excitatory modeled by Type I Leaky integrate-and-fire neurons, one inhibitory modeled by Type II Morris-Lecar neurons, the stimulations are noisy, and the intrinsic cellular properties heterogeneous. The cells are connected through exponential conductance-based synapses. We show that, in presence of tonic inhibition, 1) the oscillation frequency of the network decreases as well as subthreshold oscillations, inducing δ -rhythms in EEG-like recordings; 2) simultaneously, the amount of α -rhythms decreases supporting experimental evidence about the role of tonic inhibition.

Keywords: general anaesthesia, alpha oscillations, delta oscillations, numerical simulations, propofol, tonic inhibition

0.0.1. Highlights

- Tonic inhibition induces a decrease in spiking frequency of neural network activity;
- The spike synchrony of the excitatory neuronal population decreases as tonic inhibition increases;
- α -rhythms in neuronal subthreshold activity tend to disappear in presence of tonic inhibition;
- The peak in the power spectrum of the mean membrane voltage shifts toward δ -frequencies with the increase of tonic inhibition.

1. Introduction

Several anesthetic agents such as propofol (2,6-di-isopropylphenol) are known to elicit δ -rhythms (2-4 Hz) (Gugino et al., 2001) in the EEG of the occipital lobe and α -rhythms (10-15 Hz) in the EEG of more frontal parts of the cortex (Feshchenko et al., 2004; Gugino et al., 2001). Moreover, these phenomena are correlated with loss of consciousness. At the molecular level, propofol has been relatively well studied: the anesthetic induces inhibition effects through the binding to GABA_A receptors (Houston et al., 2012). Two major effects were especially highlighted experimentally: phasic (Kitamura et al., 2003) and tonic inhibition (Glykys and Mody, 2007). In addition to the phasic inhibition obviously resulting from synaptic action, a process of tonic inhibition takes place when propofol is injected. Indeed, as the blood concentration increases, molecules bind to extra-synaptic GABA_A receptors, resulting in tonic inhibitory currents (Houston et al., 2012; Glykys and Mody, 2007; Farrant and Nusser, 2005; Scimemi et al., 2005; Semyanov et al., 2004; Mody, 2001).

Although the molecular effects of propofol are relatively well known, the interplay between the microscopic effects and the macroscopic observations in EEG, reflecting neuronal population activity, remains to be elucidated.

Models of synaptic action of propofol have been recently proposed in an attempt to explain the appearance of frontal α -rhythms (Ching et al., 2010), however no model can successfully reproduce the important amount of occipital δ -rhythms found under general anaesthesia nor the decrease of α -rhythms. The latest model included only synaptic effects of propofol, but never took into account consequences of extra-synaptic bindings. Hence, a potentiation of GABA_A synaptic current, only, does not seem to account for the latter phenomenon.

The present work uses a computational model and numerical simulations to demonstrate, through tonic inhibition, that binding to extra-synaptic receptors has non negligible effects at the neural network level under propofol anaesthesia. The model shows different effects of tonic inhibition, both on spiking and neuronal subthreshold activity: 1) the mean spiking activity is decreased by tonic inhibition, 2) in our study, spike synchrony of excitatory cells decreases as propofol concentration increases, 3) the amount of alpha oscillations in EEG-like signals decreases when tonic inhibition increases, 4) it induces an increase in the ratio between the amount of δ -oscillations and α -oscillations.

The paper is organized as follows: in section 2, we describe the neuron models used and the neural network designed for the simulations; section 3 summarizes the simulation results showing the effect of tonic inhibition on the raster plots of inhibitory and excitatory cells, and EEG-like data (mean excitatory membrane voltage).

2. Material and methods

2.1. Neuron models and tonic inhibition

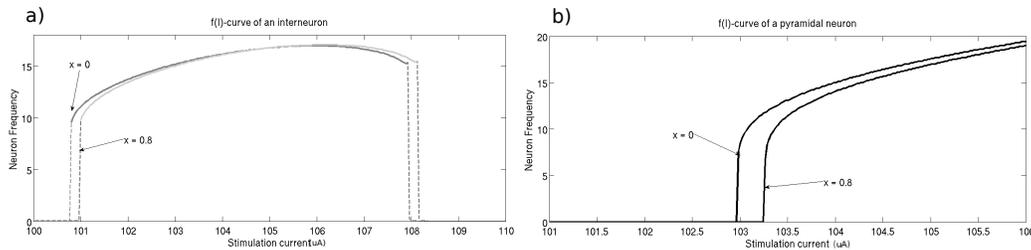


Figure 1: $f(I)$ curves – Effects of tonic inhibition. x represents the amount of tonic inhibition in μS : here $g_{ton1} = 20x \mu\text{S}$ namely $6\mu\text{S}$ and $16\mu\text{S}$, and $g_{ton2} = 100x \mu\text{S}$ namely $30\mu\text{S}$ and $80\mu\text{S}$.

On the one hand, tonic inhibition apparently is mainly due to the presence of extra-synaptic receptors (Houston et al., 2012; Glykys and Mody, 2007; Farrant and Nusser, 2005; Scimemi et al., 2005; Semyanov et al., 2004; Mody, 2001). On the other hand, we know that extra-synaptic receptors are found on inhibitory as well as on excitatory neurons and, therefore, that tonic inhibition affects the two types of neuronal populations in different brain areas (Song et al., 2011; Belelli et al., 2009; Kullmann et al., 2005). Our modeling thus takes into account the effects of tonic inhibition on two different neuronal populations: pyramidal cells ($400 \mu\text{m}^2$) modeled by Type I excitatory cells according to a Leaky-Integrate-and-Fire (LIF) neuron published in (London et al., 2008) and interneurons modeled by Type II inhibitory cells described by the Morris-Lecar model (Borisjuk and Rinzel, 2005).

Shunting/tonic inhibition occurs when the equilibrium potential of the tonic inhibition terms is very close to the resting potential of the cell; here, we model tonic inhibition by setting a pair of equilibrium potential and maximal conductance for the excitatory cells and a pair for the inhibitory cells. The

equations of the two neuron models including tonic inhibition are provided below.

2.2. Equations of the Leaky-Integrate-and-Fire model (LIF)

$$\begin{aligned}\frac{dV}{dt} &= (g_L(E_L - V) + I + g_e * (E_{eif} - V) + g_i(E_{iif} - V) + g_{ton1}(E_{ton1} - V)) / C1 \\ dg_{syn} &= -\frac{g_{syn}}{\tau_{syn}}\end{aligned}\quad (1)$$

where $syn = e$ or i ,
when spike, $g_{syn} \rightarrow g_{syn} + w_{syn}$

where $E_{ton2} = -76mV$ (resting potential of Type I neurons) $C1 = 33.181nF$ (membrane capacitance), $g_L = 22.88nS$, $\tau_m = C1/g_L = 14.5ms$ (membrane time constant), $E_L = -76.mV$, $V_T = -58.0mV$ (threshold), $V_r = -68.0mV$ (reset), refractory period = $8.0ms$.

2.3. Equations of the Morris-Lecar model (ML)

$$\begin{aligned}\frac{dV}{dt} &= (-g_{Ca}m_\infty(V)(V_{Ca}) - g_Kw(V - V_K) - g_L(V - E_L) \\ &\quad - g_e(V - E_e) - g_i(V - E_i) - g_{ton2}(V - E_{ton2}) + I_{app}) / C2 \\ \tau_w(V) \frac{dw}{dt} &= w_\infty(V) - w \\ dg_{syn} &= -\frac{g_{syn}}{\tau_{syn}}\end{aligned}\quad (2)$$

where $syn = e$ or i ,
when spike, $g_{syn} \rightarrow g_{syn} + w_{syn}$

where g_L is the leak conductance, E_L is the resting potential, $g_{e,i}$, $E_{e,i}$, $\tau_{e,i}$ the parameters for the synapses, and g_{ton} , E_{ton} the parameters for tonic inhibition. m_∞ , w_∞ , τ_w are given by:

$$\begin{aligned}m_\infty &= 0.5(1 + \tanh((V - V1)/V2)) \\ w_\infty &= 0.5(1 + \tanh((V - V3)/V4)) \\ \tau_w &= 1/(\cosh((V - V3)/(2 \times V4))) \text{ (homogeneous to time)}\end{aligned}$$

The fixed parameter values have been chosen as following:

$$E_{ton2} = -60.9mV(\text{resting potential of Type II neurons}) C2 = 20\mu F/cm^2; V1 = -1.2mV; V2 = 18mV; g_K = 8mS/cm^2; g_L = 2mS/cm^2; V_{Ca} = 120mV; V_K = -84mV; V_L = -60mV$$

$$V3 = 2mV; V4 = 30mV; g_{Ca} = 4mS/cm^2; \varphi = 0.04;$$

Regarding tonic inhibition, we define tonic conductances that are physiologically reasonable for GABA_A receptors (Song et al., 2011; Farrant and Nusser, 2005) with respect to the stimulation needed to make the model neurons fire: $g_{ton1} = 20x \mu S$ and $g_{ton2} = 100x \mu S$ where x varies between 0 and 1, namely g_{ton1} varies between $0\mu S$ and $20\mu S$, and g_{ton2} varies between $0\mu S$ and $100\mu S$.

In our study, the conductance of tonic inhibition in inhibitory cells is higher than for excitatory cells. This is consistent with the experimental observation that the tonic GABA_A current usually is significantly smaller than those in interneurons (Song et al., 2011; Scimemi et al., 2005).

The firing profiles of the two neurons are shown in Fig 2.1 without and with different amounts of tonic inhibition. At the cellular level, we notice a shift of the $f(I)$ -curves to the right as tonic inhibition increases.

2.4. Noise and heterogeneity

2.4.1. Noisy stimulations

The external input current applied during the simulations is given by constant values disturbed by white noise the following way:

$$I_{app} = I_0 + \mathcal{W}[-60.0 \mu A; 60.0 \mu A](t, neuron) \text{ (for Type II neurons)}$$

$$I_{app} = I_{constant1} + \mathcal{W}[-2.0 \mu A; 2.0 \mu A](t, neuron) \text{ (for Type I neurons)}$$

where $I_{constant1}$ is a constant value fixed for each simulation and I_0 , defined below, depends on a constant value fixed for each simulation. This means that, for the values of I_0 and $I_{constant1}$ we use, the firing frequencies of the neurons in the network range from 0 to 17 Hz, which is a rather high level of noise.

2.4.2. Heterogeneity

Heterogeneity can be added in the following way:

- For Type II
we add a spatial (i.e. over neurons) perturbation to the external input I_{app} such that: $\forall i \in \{\text{Neurons}\} I_0(i) = I_{\text{constant2}} + \mathcal{N}(0, \epsilon_2)$ (in μA), where ϵ and $I_{\text{constant2}}$ are constant values fixed for each simulation.
- For Type I
we add a spatial (i.e. over neurons) perturbation to the firing threshold such that: $V_T = -49 + \mathcal{N}(0, \epsilon_1)$ (in mV)

For the results shown in the present article, we chose $\epsilon_2 = 2$ and $\epsilon_1 = 0.0001$.

2.5. The neuronal network

1000 (750 LIF excitatory Type I and 250 ML inhibitory Type II) neurons are randomly connected with exponential synapses and the following parameters:

- $\tau_e = 5ms$ (excitatory time constant)
- $\tau_i = 20ms$ (inhibitory time constant)
- $w_{ee} = 0.005mS$ (excitatory synaptic weight to excitatory neurons),
- $w_{ie} = 0.008mS$ (inhibitory synaptic weight to excitatory neurons)
- $w_{ei} = 0.4mS$ (excitatory synaptic weight to inhibitory neurons)
- $w_{ii} = 0.5mS$ (inhibitory synaptic weight to inhibitory neurons)
- $E_i = -27.mV$
- $E_e = 0mV$ (excitatory resting state)
- $E_{iif} = -75mV$

The time constants of the synapses correspond, for the excitatory neurons, to those found in NMDA-type synapses, and for the inhibitory, to those found in GABA_A synapses.

The connectivity, very sparse, reflects cortical connectivity (Binzegger et al., 2004). We used the following probabilities of connections:

- inhibitory neurons to inhibitory neurons: 0.05
- inhibitory neurons to excitatory neurons: 0.02
- excitatory neurons to inhibitory neurons: 0.01
- excitatory neurons to excitatory neurons: 0.005

2.6. Analysis tools

2.6.1. Coherence Measure

Several ways of calculating the coherence of neural network activity have been described in the literature. The method that we have retained is the coherence measure, κ , based on the normalized cross-correlation of neuronal pairs in the network and described in Wang and Buzsáki (1996). The coherence of two neurons x and y is calculated as following by the cross-correlation of their spike trains represented by X and Y within a time bin, τ_{bin} over a time interval T :

$$\kappa_{xy}(\tau_{\text{bin}}) = \frac{\sum_{l=1}^K X(l)Y(l)}{\sqrt{\sum_{l=1}^K X(l) \sum_{l=1}^K Y(l)}} \quad 0 \leq \kappa_{xy} \leq 1.$$

The spike trains are given by: $X(l) = 0$ or 1 and $Y(l) = 0$ or 1 , $l = 1, 2, \dots, K$ ($K = \tau_{\text{bin}}/T$), which mean that $X(l) = 1$ if there is at least one spike in the bin.

The population coherence measure

$$\kappa(\tau_{\text{bin}}) = \sum_{i=1}^{N_1} \sum_{j=1}^{N_2} \frac{\kappa_{ij}(\tau_{\text{bin}})}{M}$$

computed in this work is the the coherence measure of pairs averaged over M pairs of spike trains of the numbers N_1, N_2 . For instance, $M = N(N - 1)/2$ for intra-network coherence with $N_1 = N_2 = N$ neurons, while $M = N_1N_2$ for coherence measures between different networks of number N_1, N_2 neurons. When the time bin τ_{bin} is very small, a strong synchrony renders $\kappa(\tau_{\text{bin}})$ close to 1, and the smaller $\kappa(\tau_{\text{bin}})$ the less synchronized the network activity.

2.6.2. Power spectrum

The power spectra are computed with a fast Fourier transform and average over 10 simulations of the network with different random initializations (of noise, heterogeneity and connections) with a duration of 5s and a time resolution of 5ms, however, the simulation are run with an integration time of 0.5ms to avoid numerical integration problems. The fft Matlab routine is used to compute the spectra; this method decomposes the problem using the Cooley-Tukey algorithm; then, we take its absolute value, center it and keep only the positive frequencies.

3. Results

Propofol modifies GABAergic receptor dynamics and changes neural firing activity, exerting either inhibition or even excitation Borgeat et al. (1991); McCarthy et al. (2008) It is also known that anesthetic agents induce changes in EEG recordings indicating that they alter the subthreshold activity of excitatory neurons since EEG is known to originate from subthreshold dendritic currents on spatially aligned apical cortical dendrites (Nunez and Srinivasan, 2005). In order to investigate whether tonic inhibition can induce such changes in EEG and decrease in neural firing activity, we perform a study of the power spectra of the subthreshold activity in excitatory neurons and the neural firing statistics in the populations.

To understand the effect of tonic inhibition on single neuron activity, Fig. 2.1 illustrate the firing rate of excitatory and inhibitory neurons subjected to external constant currents. Pyramidal cells exhibit a sigmoid-like $f - I$ -curve and tonic inhibition shifts it to the right, i.e. a larger stimulation current is necessary to elicit spikes in presence of tonic inhibition (Fig. 2.1(a)).

These findings, have consequences on neuronal population activity as described in the following.

Figure 2 illustrates the behavior of the network when tonic inhibition is applied. In the absence of tonic inhibition, the network displays synchronized patterns of oscillations at about 9.5Hz (α -band) visible in the raster plots of the two neuronal populations and which also show up in the power spectrum of the subthreshold activity of the pyramidal neuronal population (Fig 2a).

When weak tonic inhibition is added (Fig 2b), the spiking activity slows down slightly, the synchrony visibly decreases and the activity becomes more random. Moreover, the frequency of the maximum power spectral density shifts to lower values, i.e. the amplitude of the α -activity decreases.

As the level of tonic inhibition keeps increasing (Fig 2c), the neurons fire more randomly accompanied by an extinction of α -activity in the neural firing and subthreshold activity and a clear enhancement of δ -activity in the subthreshold activity as observed in the power spectral density. Finally, when the level of tonic inhibition x is increase further ($x = 1.2$), the firing of excitatory neurons stops and the inhibitory neurons remain active only for a transient period before its neuronal activity fully dies out as well (result not shown).

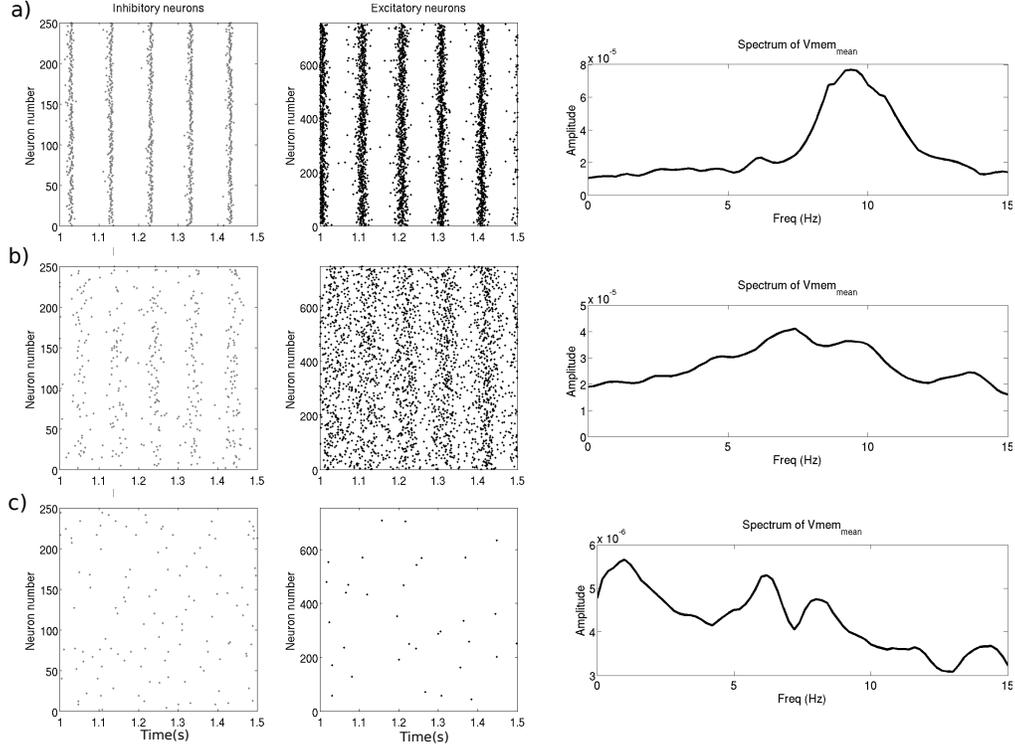


Figure 2: Raster plots of inhibitory and excitatory populations and smoothed power spectrum of the membrane voltage of the excitatory cells: a) without tonic inhibition, b) with tonic inhibition, $x = 0.3$, $g_{ton1} = 6 \mu\text{S}$ and $g_{ton2} = 30 \mu\text{S}$, and c) with $x = 0.8$, $g_{ton1} = 16 \mu\text{S}$ and $g_{ton2} = 80 \mu\text{S}$.

To further quantify the effect of tonic inhibition on the network activity, Fig. 3(a) presents the intra-population coherence in the excitatory and inhibitory population and the inter-correlation between the excitatory and inhibitory population. It shows that the coherence in the excitatory population (dashed grey curve) decreases monotonically with the increase of tonic inhibition. In contrast the coherence of the inhibitory population (plain black curve) exhibits a minimum at $x \approx 0.45$ and increases as the activity of the excitatory population becomes more random for $x > 0.45$. The coherence between the pyramidal and the interneuron populations (plain grey curve) is maximal, one notices a maximal synchrony at about $x = 0.16$ (zoom in

not shown), then the coherence becomes weaker in conjunction with the reduction of the excitatory activity. A computation of κ for various bin sizes τ shows that it does not increase linearly with τ (not shown) and hence the relative firing time of neural pairs is not uniformly distributed between 0 and the oscillation period and thus that there really is synchronization, see the paper of Wang and Buzsaki for details..

Moreover, Fig. 3b) shows that the ratio between the amount of δ -rhythms and the amount of α -rhythms (dashed black curve) increases as a sigmoid-like function of the propofol concentration x and becomes larger than 1 from $x = 0.575$ (namely the quantity of α -rhythms is then smaller than the quantity of δ -rhythms); the ratio between the amount of θ and α -rhythms (plain gery curve) increases for $x \in [0; 0.6]$, becoming larger than 1 for $x > 0.49$. The curve of the θ ratio crosses the curve of the δ ratio at $x = 0.65$ where the mount of delta remains higher. The ratio between β and α (dotted grey line) is larger than the ratio θ/α for $x \in [0; 0.19]$, and larger than the ratio δ/α for $x \in [0; 0.32]$, then is significantly smaller than the two other ratios reaching a maximum of 0.54 for $x = 0.8$.

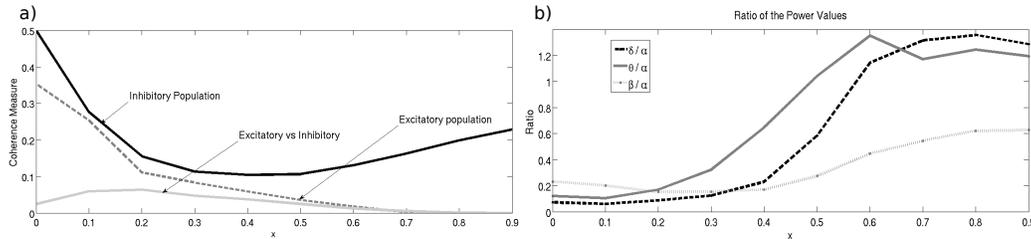


Figure 3: Summary of the effects of tonic inhibition on the synchrony and the subthreshold activity of a cortical neural network. a) Coherence measure of the neuronal population as a function of propofol concentration (x). b) Ratio of the power values δ , θ and β compared to α .

4. Discussion – Conclusion

We could show with a two neuronal population model that, despite the role of extrasynaptic receptors is often neglected, the resulting tonic inhibition greatly modifies the behavior of neural networks under propofol anaesthesia.

On the one hand, regarding single neurons, we observed a shift to the right of the $f(I)$ curve in presence of tonic inhibition which implies, for a same stimulation value, a lower firing frequency for the excitatory cells.

As a consequence, since excitatory cells entrain the inhibitory population, the whole network fires less often (raster plots of Fig. 2 and Fig. 3a). Hence, the amount of fast oscillations, like α decreases and the relative amount of δ increases as the propofol's concentration increases (Fig. 2 power spectra and Fig. 3b), suggesting that tonic inhibition could be at the origin of increased δ -frequencies in occipital EEGs δ -frequencies in occipital EEGs.

On the other hand, in our case, synchrony is lower with than without tonic inhibition: tonic inhibition tends to “disconnect” the network. This decrease of spike synchrony explains in turn the diminution of the mean power spectra amplitude in Fig. 2.

Slow oscillations in presence of tonic inhibition emerge as a network effect in the sense that single cells we used here, in particular the inhibitory cells, cannot fire at frequencies below about 8Hz. Moreover, noise and cell's heterogeneity play a major role in “stabilizing”, making more robust, the effect of tonic inhibition; indeed, in absence of noise and heterogeneity, the network activity is very sensitive to tonic inhibition level that rapidly has a shunting effect as the level of tonic inhibition increases. We suspect that the effects of tonic inhibition would be even more stable with cells whose the firing profile, i.e. the $f(I)$ -curve, would be less steep at slow frequencies. This would suggest that the appearance of δ -rhythms is not only due to tonic inhibition but also depends on intrinsic cellular properties.

To conclude, tonic inhibition through binding to extrasynaptic receptors probably accounts for the increased amount of δ -rhythms recorded in occipital EEGs. This, together with the fact that extrasynaptic receptors are largely present in the hippocampus, could also, at least partly, explain observations made in the MTL under general anaesthesia (Fell et al., 2005).

References

- Belelli, D., Harrison, N.L., Maguire, J., Macdonald, R.L., Walker, M.C., Cope, D.W., 2009. Extrasynaptic GABAA receptors: form, pharmacology, and function. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 29, 12757–12763.
- Binzegger, T., Douglas, R.J., Martin, K.A.C., 2004. A Quantitative Map of

- the Circuit of Cat Primary Visual Cortex. *The Journal of Neuroscience* 24, 8441–8453.
- Borgeat, A., Dessibourg, C., Popovic, V., Meier, D., Blanchard, M., Schwander, D., 1991. Propofol and spontaneous movements: an EEG study. *Anesthesiology* 74, 24–27.
- Borisjuk, A., Rinzel, J., 2005. Understanding neuronal dynamics by geometrical dissection of minimal models, in: *Models, in Neurophysics*, M. (Eds.), Proc. Les Houches Summer School 2003, pp. 19–52.
- Ching, S., Cimenser, A., Purdon, P.L., Brown, E.N., Kopell, N.J., 2010. Thalamocortical model for a propofol-induced α -rhythm associated with loss of consciousness. *Proceedings of the National Academy of Sciences* .
- Farrant, M., Nusser, Z., 2005. Variations on an inhibitory theme: phasic and tonic activation of GABA_A receptors. *Nature Reviews Neuroscience* 6, 215–229.
- Fell, J., Widman, G., Rehberg, B., Elger, C.E., Fernández, G., 2005. Human mediotemporal EEG characteristics during propofol anesthesia. *Biological Cybernetics* 92, 92–100.
- Feshchenko, V.A., Veselis, R.A., Reinsel, R.A., 2004. Propofol-induced alpha rhythm. *Neuropsychobiology* 50, 257–266.
- Glykys, J., Mody, I., 2007. Activation of GABA_A Receptors: Views from Outside the Synaptic Cleft. *Neuron* 56, 763–770.
- Gugino, L.D., Chabot, R.J., Prichep, L.S., John, E.R., Formanek, V., Aglio, L.S., 2001. Quantitative EEG changes associated with loss and return of consciousness in healthy adult volunteers anaesthetized with propofol or sevoflurane. *British Journal of Anaesthesia* 87, 421–428.
- Houston, C.M., McGee, T.P., MacKenzie, G., Troyano-Cuturi, K., Rodriguez, P.M., Kutsarova, E., Diamanti, E., Hosie, A.M., Franks, N.P., Brickley, S.G., 2012. Are Extrasynaptic GABA_A Receptors Important Targets for Sedative/Hypnotic Drugs? *The Journal of Neuroscience* 32, 3887–3897.

- Kitamura, A., Marszalec, W., Yeh, J.Z., Narahashi, T., 2003. Effects of halothane and propofol on excitatory and inhibitory synaptic transmission in rat cortical neurons. *The Journal of pharmacology and experimental therapeutics* 304, 162–171.
- Kullmann, D.M., Ruiz, A., Rusakov, D.M., Scott, R., Semyanov, A., Walker, M.C., 2005. Presynaptic, extrasynaptic and axonal GABAA receptors in the CNS: where and why? *Progress in biophysics and molecular biology* 87, 33–46.
- London, M., Larkum, M.E., Häusser, M., 2008. Predicting the synaptic information efficacy in cortical layer 5 pyramidal neurons using a minimal integrate-and-fire model. *Biological cybernetics* 99, 393–401.
- McCarthy, M.M., Brown, E.N., Kopell, N., 2008. Potential network mechanisms mediating electroencephalographic beta rhythm changes during propofol-induced paradoxical excitation. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 28, 13488–13504.
- Mody, I., 2001. Distinguishing between GABA(A) receptors responsible for tonic and phasic conductances. *Neurochemical research* 26, 907–913.
- Nunez, P.L., Srinivasan, R., 2005. *Electric Fields of the Brain: The Neurophysics of EEG*, 2nd Edition. Oxford University Press, USA. 2 edition.
- Scimemi, A., Semyanov, A., Sperk, G., Kullmann, D.M., Walker, M.C., 2005. Multiple and plastic receptors mediate tonic GABAA receptor currents in the hippocampus. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 25, 10016–10024.
- Semyanov, A., Walker, M.C., Kullmann, D.M., Silver, R.A., 2004. Tonic active GABA A receptors: modulating gain and maintaining the tone. *Trends in neurosciences* 27, 262–269.
- Song, I., Savtchenko, L., Semyanov, A., 2011. Tonic excitation or inhibition is set by GABAA conductance in hippocampal interneurons. *Nature Communications* 2, 376+.
- Wang, X.J., Buzsáki, G., 1996. Gamma Oscillation by Synaptic Inhibition in a Hippocampal Interneuronal Network Model. *The Journal of Neuroscience* 16, 6402–6413.