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Sensitivity Analysis of Ventricular Activation and Electrocardiogram in Tailored Models of Heart-Failure Patients

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Abstract Cardiac resynchronization therapy is not effective in a variable proportion of heart failure patients. An accurate knowledge of each patient's electroanatomical features could be helpful to determine the most appropriate treatment. The goal of this study was to analyze and quantify the sensitivity of left ventricular (LV) activation and the electrocardiogram (ECG) to changes in 39 parameters used to tune realistic anatomical-electrophysiological models of the heart. Electrical activity in the ventricles was simulated using a reaction-diffusion equation. To simulate cellular electrophysiology, the Ten Tusscher-Panfilov 2006 model was used. Intracardiac electrograms and 12-lead ECGs were computed by solving the bidomain equation. Parameters showing the highest sensitivity values were similar in the six patients

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studied. QRS complex and LV activation times were modulated by the sodium current, the cell surface-to-volume ratio in the LV, and tissue conductivities. The T-wave was modulated by the calcium and rectifier-potassium currents, and the cell surface-to-volume ratio in both ventricles. We conclude that homogeneous changes in ionic currents entail similar effects in all ECG leads, whereas the effects of changes in tissue properties show larger inter-lead variability. The effects of parameter variations are highly consistent between patients and most of the model tuning could be performed with only ~ 10 parameters.

Keywords Computer simulation · ECG morphology · heart failure · left bundle branch block · patient-specific model · sensitivity analysis

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1 Introduction

Heart failure (HF) is one of the most commonly diagnosed cardiac diseases with an overall incidence approaching one percent of the population over 65 years of age [39]. A significant percentage of HF patients presents a prolonged QRS duration on the 12-lead electrocardiogram (ECG), and many of them are diagnosed with left bundle branch block (LBBB), indicating impairment of the cardiac conduction system specifically in the left ventricle (LV) [14]. Cardiac resynchronization therapy (CRT) has been shown to reduce mortality in most patients with clear LBBB, but its efficacy is reduced in patients with other concomitant electrical abnormalities, such as right bundle branch block, or diffuse ventricular conduction disturbances [57]. Better prediction of CRT efficacy in each particular HF patient through accurate description of their electrophysiology would help to avoid unnecessary surgical interventions. Computational models are a powerful tool to achieve this without ethical limitations.

Recent studies have shown the usefulness of patient-specific models to study complex electrophysiological phenomena in healthy and diseased subjects [28, 35]. However, it is challenging to create multiscale models able to mimic in a realistic way the entire ventricular electrical behavior from the subcellular level to the whole heart and body surface levels. Without prior knowledge, the tuning of patient-specific models is hampered by the high number of parameters involved, which may differ between patients, creating an ill-posed problem. Therefore, doing a sensitivity analysis is meant to quantify to what extent each single parameter affects the output and, finally, to identify those needing accurate tuning. Previous sensitivity analysis studies *in silico* have shown that extracellular conduction properties in the different anatomical structures (blood, lungs, fat, skeletal muscle...) of a human torso affect in different ways the electrical potentials measured at the body surface [5, 19].

In this study we created realistic models of both the anatomy and the electrical activation in six HF patients selected for CRT. The main purpose of the analysis performed in this work was to investigate the relative importance of myocardial properties in determining both the activation sequence on the LV endocardial surface and the ECG morphology in HF patients. For this analysis, individual variations of 39 parameters, including intracellular and extracellular tissue conductivity, and cellular membrane ionic properties, were evaluated. This set of 39 parameters is too large to tune a patient-specific model efficiently. Therefore, a second goal of this study was to delimit the number of parameters that are required for model tuning.

2 Methods

2.1 Patients

Six patients, with ages ranging from 53 to 79 years, diagnosed with HF and suitable for CRT implantation according to the criteria from the 2011/2013 ESC clinical practice guidelines [6] were included in this study (Table 1). Four of the six patients showed clear LBBB electrocardiographic features according to three widely used sets of criteria: the criteria used by the European Society of Cardiology (ESC) [3], the cri-

Table 1 Selected patient demographics and characteristics

| Patient | Age | Gender | Height (m) | Weight (kg) | LVEF (%) | NYHA | Etiology | QRSd (ms) | Morph. |
|---------|-----|--------|------------|-------------|----------|--------|----------|-----------|--------|
| 1 | 72 | Female | 1.57 | 75 | 39 | III-IV | CAD | 139 | LBBB |
| 2 | 69 | Male | 1.82 | 75 | 35 | II-III | IDCM | 179 | LBBB |
| 3 | 79 | Male | 1.87 | 94 | 28 | II-III | CAD | 138 | IVCD |
| 4 | 57 | Male | 1.60 | 67 | 30 | II-III | CAD | 126 | IVCD |
| 5 | 73 | Male | 1.70 | 80 | 24 | III-IV | IDCM | 154 | LBBB |
| 6 | 53 | Male | 1.78 | 110 | 34 | II-III | IDCM | 145 | LBBB |

Abbreviations: LVEF=Left Ventricular Ejection Fraction;
 NYHA=New York Heart Association Functional Classification;
 QRSd=QRS duration;
 CAD=Coronary Artery Disease;
 IDCM=Idiopathic Dilated Cardiomyopathy;
 LBBB=Left-Bundle-Branch Block;
 IVCD=Intraventricular Conduction Delay.

teria used by the American Heart Association (AHA) [49], and the criteria proposed by Strauss et al. [48]. The other two patients had non-specific ventricular conduction disturbances. These six patients underwent standard 12-lead ECG, cardiac magnetic resonance (CMR) imaging, electro-mechanical anatomical mapping (NOGA XP, Biosense Webster), and a coronary angiography procedure. In four of the six patients, the NOGA catheter was introduced in the left-ventricular (LV) cavity and had contact with the endocardium only, whereas in patients 5 and 6 the electrical activation on both right ventricular (RV) and LV endocardium as well as the ventricular epicardial layer underneath the coronary sinus was measured. Details of the data collection procedure were described in a previous study [35]. Data for the six patients were collected between 2012 and 2015 at Cardiocentro Ticino (Lugano, Switzerland). The patients provided written consent to each procedure, and the institutional review board approved the use of the data for research purposes.

2.2 Model Construction

Anatomical Segmentation Segmentation of cardiovascular structures, lungs, and torso of each patient was performed based on CMR data obtained with a 3T Siemens Magnetom Skyra Scanner equipped with a dedicated cardiac 36-channel coil and a standard body coil for thorax examinations. CMR images were obtained in axial, coronal, sagittal, cardiac short axis, and cardiac long axis planes, with a slice thickness between 1 and 8 mm [35]. Late gadolinium-enhancement images for scar detection were obtained 10 minutes after intravenous infusion of gadolinium (Gadobutrol, 0.2 mmol/kg body weight) using a T-weighted gradient-echo pulse sequence with a phase-sensitive inversion recovery reconstruction. Semi-automatic segmentation of ventricular epicardium and endocardium was performed as described in a previous study [12]. Atrial cavities, aorta, inferior and superior vena cava, and pulmonary trunk were manually segmented using custom-made software for tracing contours. Lungs, torso, and ECG electrode locations were manually traced on ultra-fast

volume-interpolated breath-hold examination (VIBE) images following the injection of the contrast agent.

Mesh Construction The segmentation contours were used to generate a three-dimensional (3D) mesh of the tissue boundaries. Mesh construction, i.e. creation of closed surfaces representing the different structures, was performed using the Blender software (The Blender Foundation, Amsterdam, The Netherlands). All cardiovascular structures were joined to define a 3D model of the heart, which together with the lungs and torso meshes constituted the complete anatomical model of the patient. In order to extract the 12-lead ECG from the simulations, nine virtual electrodes were placed on the torso surface at the same locations as in the real patients: three for the limb leads, and six for the precordial leads. Both in reality and in the model the arm electrodes were placed on the shoulders and the left-leg electrode was placed near the hip.

Definition of Tissue Properties From the surface meshes defining the anatomy, volumetric meshes for the computations were created. Since the computational code worked with semi-structured finite-difference meshes, this conversion consisted of simply overlaying the surfaces on a regular 3D grid and assigning tissue types to the grid elements according to the surfaces in which they were contained. We used a mesh with 0.2-mm spacing for the heart and 1-mm spacing for the torso. Passive tissue properties were associated with grid elements, while model variables and active properties such as densities of ionic currents were associated with the nodes of the grid [33]. The node types, which determine the parameters of the ionic model, were determined from the element types using a set of rules that ensure model consistency [33]. The passive properties were the scalar conductivities (σ) in longitudinal, transversal and cross-sheet directions, and cell surface-to-volume ratio (β), i.e. the amount of cellular membrane per unit volume. For ventricular myocardium, fiber rotation from endocardial to epicardial layers was included by using a rule-based method [33]. Four layers were simulated in the myocardium: epicardium (the most external layer), midmyocardium (middle layer), endocardium (inner layer), and a thin fast endocardial layer (0.6 mm) to mimic the rapid propagation occurring due to the Purkinje system in the endocardium. The first three layers (epicardium, midmyocardium and endocardium) were differentiated by their cellular electrophysiology [55]. A full layer was used to represent the network of Purkinje fibers because the actual topology cannot be recorded in vivo [27]. This fast endocardial layer was initially assumed to have the same properties in both ventricles, although previous studies proposed to differentiate them for the stimulation profile [18, 41].

2.3 Electrophysiological Simulation

Cardiac electrophysiology was simulated with a monodomain reaction-diffusion model [33]. Electrophysiological activity of ventricular myocytes was simulated using the Ten Tusscher and Panfilov 2006 membrane model, which includes transmembrane

ionic currents, regulation of sodium, potassium, and calcium intracellular ion concentrations, and intracellular calcium handling in the sarcoplasmic reticulum [55]. The ECG was simulated by solving the extracellular potential ϕ_e from the equation

$$\nabla \cdot ([G_i + G_e] \nabla \phi_e) = -\nabla \cdot (G_i \nabla V_m) \quad (1)$$

where G_i and G_e are the intracellular and extracellular conductivity tensor fields, respectively, and V_m is the transmembrane potential field [34]. The reaction-diffusion simulation was run on a mesh with 0.2 mm resolution. The right-hand side of equation (1) was evaluated on this fine mesh and then projected on the coarser (1 mm) torso mesh, which included a downsampled model of the heart. ϕ_e was solved on the coarse mesh. The 12-lead ECG was then computed by extracting the extracellular potential at the electrode locations.

All simulations were performed using propag-5, software specifically created for cardiac simulation and parallelizable to utilize supercomputing facilities of the Swiss National Supercomputing Centre (CSCS) [22, 33]. Most simulations ran on 1536 cores of a Cray XC40 system. Computation times for each single simulation were in the range of 40 to 60 minutes for 600 ms of simulations of V_m and ϕ_e .

2.4 Endocardial Activation Map and ECG Matching

Once the model was complete, parameter adjustments were iteratively performed in order to match as closely as possible the electrical activation pattern measured with the NOGA catheter on the endocardial surface, as well as the morphological features of the clinically obtained ECG. In this study, electrical activity was only generated in the ventricular myocardium. Therefore the P-wave was not simulated and the ECG analysis was focused on the QRS complex and the T-wave. Figure 1 shows the modeled RV and LV endocardial surfaces of patient 5 together with the measured activation times (color-coded) at 396 NOGA recording sites for this patient. Activation time at a recording site was computed as the time interval between the QRS onset (vertical red line on the right panel of Figure 1) and the steepest negative deflection of the electrogram (blue dot on the right panel of Figure 1).

The iterative strategy of parameter adjustments differed between patients, but in general consisted of: 1) finding the appropriate number and locations of early activation sites to match the initiation of the LV activation; 2) tuning surface-to-volume ratios, conductivities, and the sodium current conductance to adjust the conduction velocity in the myocardium until good matching in the LV activation pattern was obtained; 3) refining adjustments performed in step 2 to improve matching of the QRS complex; 4) adjusting properties of the L-type calcium current and potassium currents to match the T-wave. Note that changes in surface-to-volume ratios, conductivities, and the sodium current conductance affect conduction velocity, but the way they do it may be different, so steps 2) and 3) are typically performed simultaneously: changes in surface-to-volume ratios modify propagation features homogeneously in a specific part of the ventricles (LV, RV, septum...); changes in conductivities allow us modify propagation features of cardiac tissue anisotropically in specific directions (longitudinal, transversal, or cross-sheet); and changes in the sodium current conductance

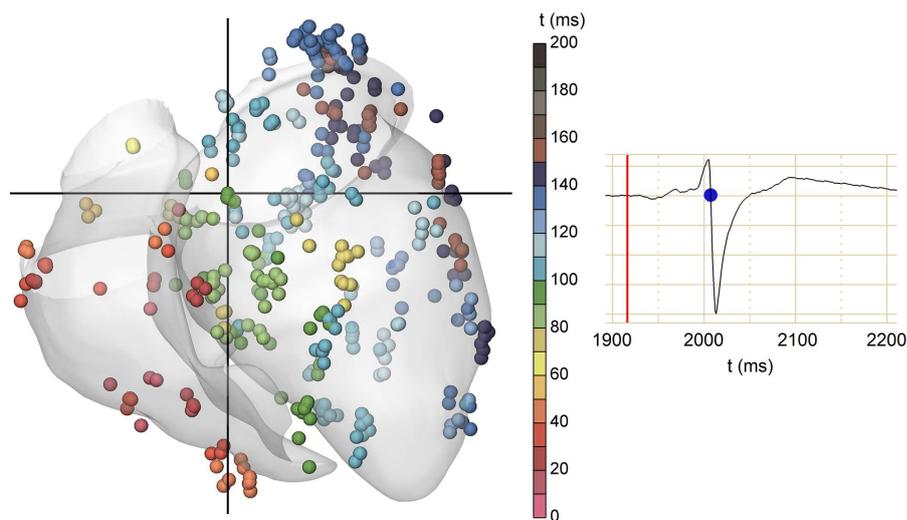


Figure 1 Modeled endocardial surfaces of RV and LV for patient 5 and measured activation times at NOGA recording sites in a color scale (left panel). Example of an electrogram recorded at the LV endocardium at the position of the crosshair in the left panel (right panel), together with the time of QRS onset (vertical red line) and the activation time (blue dot).

modify excitability of cardiac cells based on their type in both the LV and the RV (endocardial cell, M cell, or epicardial cell). Tuning was stopped when we could not find a way to improve it further. Parameters were changed in physiologically plausible ways, e.g. changes in β and σ in some patients corresponded to hypertrophied hearts. A complete list of tuned parameters and tailored models for the six patients is shown in Supplemental Material.

Figure 2 shows the anatomical models of the six patients together with leads I, aVL, V5 and V6 of the measured ECG (red) and simulated ECG (black). These four leads were chosen due to their importance for LBBB diagnosis. On the right panels of Figure 2, correlation between simulated and measured activation times is shown, including the fitted line and the correlation coefficient.

2.5 Univariate Sensitivity Analysis

A quantitative analysis to evaluate the relative importance for the cardiac electrical activity of individual parameters in the model was performed. Two groups of parameters were distinguished: those characterizing cellular ionic properties, and those characterizing passive electrical properties of the tissue. In the first group, maximum conductances of all transmembrane ionic currents were included: fast sodium current (I_{Na}), L-type calcium current (I_{CaL}), inward rectifier potassium current (I_{K1}), transient outward current (I_{to}), rapid delayed rectifier potassium current (I_{Kr}), slow delayed rectifier potassium current (I_{Ks}), sodium/potassium pump current (I_{NaK}) and sodium/calcium exchanger current (I_{NaCa}); and also two parameters to modify inactivation

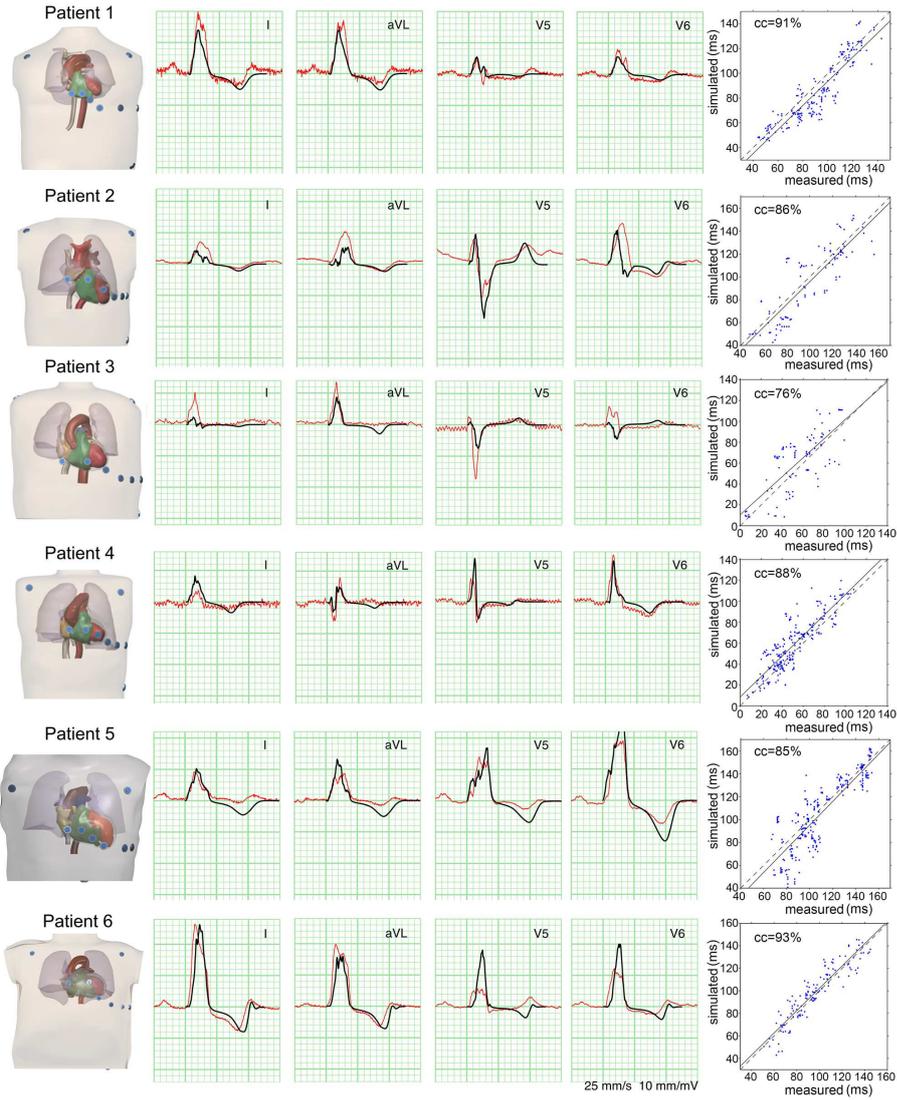


Figure 2 Anatomical models of the six patients (left), and corresponding measured (red), and simulated ECG (black) in leads I, aVL, V5, and V6. Correlation between simulated and measured activation times on the LV endocardium (right).

kinetics of I_{CaL} (τ_f) and activation kinetics of I_{Ks} (τ_{xs}), known to be relevant in the adaptation of cell repolarization to heart rate [36]. These parameters were chosen in agreement with their importance in determining ventricular cell properties as shown in a previous study [40]. The second group of parameters consisted of cell surface-to-volume ratio (β), intracellular conductivities (σ_i), and extracellular conductivities (σ_e) in the three directions of propagation: longitudinal (σ_{il} and σ_{el}), transversal (σ_{it} and σ_{et}), and cross-sheet (σ_{ic} and σ_{ec}). They were analyzed separately for each tissue

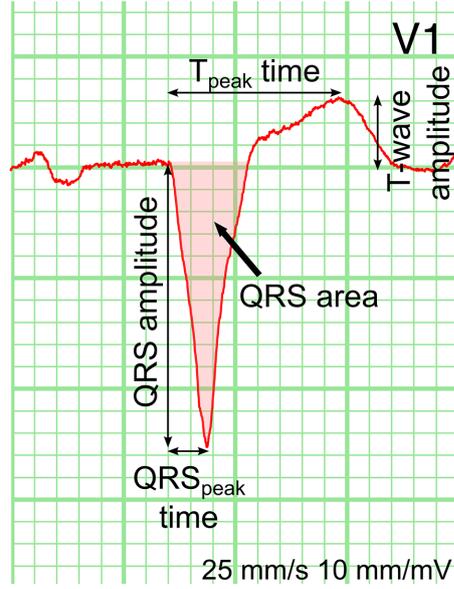


Figure 3 Example of lead V1 of a clinically measured ECG with five of the markers included in the sensitivity analysis: QRS peak time, QRS amplitude, QRS area, T peak time and T-wave amplitude.

type: LV, RV, fast endocardial layer (fendo), blood, skeletal muscle, and lungs. In the rest of this manuscript the tissue type will be shown as an additional subscript of β and σ . In total, changes in 39 parameters were studied.

To quantify the effects of each parameter on particular ECG markers and ventricular activation, $\pm 30\%$ changes with respect to the adjusted values of the model for each patient were simulated, in agreement with the magnitude of changes (25-30%) reported in previous studies [11, 19, 40, 42, 43].

Ten ECG markers were computed for each of the 12 ECG leads: QRS peak time, absolute value of QRS amplitude with respect to the baseline, QRS area, QRS skewness, QRS kurtosis, QRS peak-T peak interval, absolute value of T-wave amplitude with respect to the base line, T peak time, T-wave skewness, and T-wave kurtosis (examples of some of these markers are shown in Figure 3). Furthermore, catheter positions were projected on the simulated endocardial surface so that simulated activation times and measured activation times could be compared. Linear regression between the simulated and measured activation times was performed, and the slope of the fitted line was also included in the sensitivity analysis.

The sensitivity of a marker, m , to changes in a particular parameter, p , was quantified by computing:

$$S_{m,p}(\%) = \frac{m_{p+30\%} - m_{p-30\%}}{0.6m_{p\pm 0\%}} 100 \quad (2)$$

where $m_{p+30\%}$, $m_{p-30\%}$, and $m_{p\pm 0\%}$ are the values of the marker m when the parameter p is varied by $+30\%$, -30% and $\pm 0\%$ (unvaried), respectively, with respect to their default value in the model. A factor of 0.6 is included to account for the 60%

range of variation. The computed value, $S_{m,p}$, represents the percentage of variation of the marker m if the parameter p were varied by 100%, considering the effect of changes as linear.

A model parameter was considered to be relevant in the modulation of a marker if the sensitivity value was higher than 50% of the maximum sensitivity among all parameters for that particular marker. In the ECG markers, this consideration was taken in each of the 12 leads separately.

3 Results

Figure 4 shows examples of simulations conducted for the parameter sensitivity analysis. In particular, effects of changes in I_{Na} on the I, aVR, V5 and V6 ECG leads (a), LV endocardial activation map (b), displayed as a 2D polar diagram showing measured and simulated activation times in a color scale, and linear fit between simulated and measured activation times (c) are displayed. In this example, average computed values for sensitivity to I_{Na} changes were highly relevant for QRS peak time ($S_{QRSpeak,I_{Na}}=-30.1\%$), QRS area ($S_{QRSarea,I_{Na}}=-35.8\%$), and QRS morphology ($S_{QRSskew,I_{Na}}=-193.6\%$, $S_{QRSkurt,I_{Na}}=11.1\%$). Regarding the T-wave, the sensitivities of the T-wave amplitude ($S_{Tamp,I_{Na}}=-34\%$) and the T-wave morphology were also relevant ($S_{Tskew,I_{Na}}=52.1\%$, $S_{Tkurt,I_{Na}}=30.5\%$). I_{Na} changes entailed minimal effects on QRS amplitude ($S_{QRSamp,I_{Na}}=3\%$), T-wave peak time ($S_{Tpeak,I_{Na}}=-7.8\%$), and QRS-T interval ($S_{QRS-T,I_{Na}}=-1.1\%$) in this patient. Furthermore, effects of I_{Na} changes on the linear fitted slope were also notable ($S_{LVslope,I_{Na}}=-37\%$). Negative sensitivity values reflected that an increase in I_{Na} led to a decrease in the analyzed marker, and vice versa.

In general, the parameters showing the highest sensitivity values were similar in the six patients. Figures 5, 6 and 7 show the parameters whose variations exerted the strongest alterations on LV activation and ECG markers, respectively, in the six patients. Figure 5 shows the sensitivity values (horizontal axis) for each relevant parameter (vertical axis) and patient (color coded). In Figures 6 and 7 each horizontal line represents the average sensitivity value \pm standard deviation for the 12 ECG leads for a particular parameter (vertical axis) and patient (color coded).

LV Activation The slope of the fitted line between simulated and measured activation times was sensitive to several parameters, all related to conduction velocity in the myocardium in both ventricles, as shown in Figure 5. Effects were consistent between the six patients, but it was observed that changes in the tissue properties of the RV only entailed significant variations in the slope in patient 5 and, to a lesser extent, patient 1. This was probably due to anatomical particularities in the septum properties and the early activation sites of these patients (see Supplemental Material).

QRS-complex Morphology As shown in Figure 6, QRS peak time was mainly determined by those parameters affecting conduction velocity in the myocardium such as I_{Na} , β_{LV} , and β_{RV} . Parameters affecting conductivity in the fast endocardial layer,

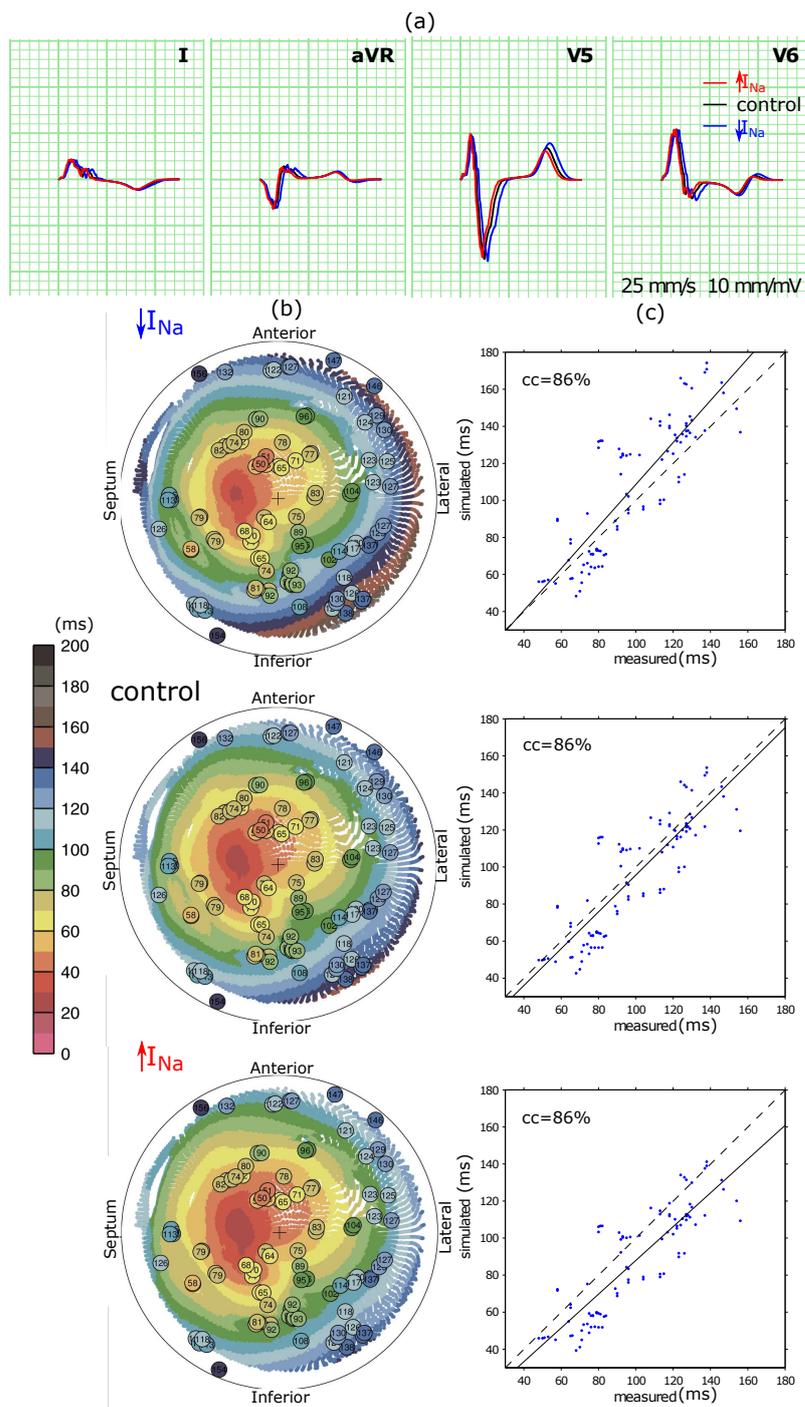


Figure 4 Simulation results. (a) ECG for patient 2. Leads I, aVR, V5, and V6 with I_{Na} reduction by 30% (blue), no changes (black), and I_{Na} increase by 30% (red). (b) Polar diagram of the LV endocardium showing measured activation times (circles with time in milliseconds) and simulated activation times (colored dots). Apex is marked with a cross, '+', at the center of the diagram. (c) Simulated versus measured activation times on the LV endocardium. Correlation coefficients are shown.

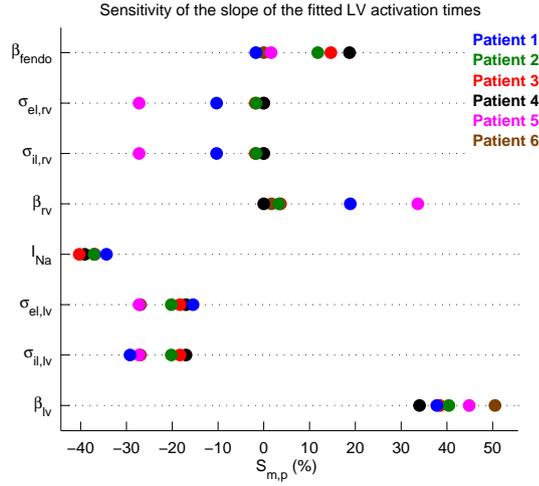


Figure 5 Significant parameters following sensitivity analysis results for the slope of the fitted line between simulated and measured activation times for the six patients (different colors for each patient).

like β_{fendo} , $\sigma_{il,fendo}$, or $\sigma_{el,fendo}$, also affect QRS peak time, but they are not represented in Figure 6 because they are not so notable. Variations in QRS amplitude were observed when applying changes in conduction properties in both ventricles, but also changes in some conductivities of the torso: $\sigma_{e,musc}$ (simultaneous changes in the three directions), and $\sigma_{et,musc}$ (changes only in the transversal direction). Similarly, QRS area was mainly modulated by changes in conduction in the LV: β_{LV} (with a large variability between leads), $\sigma_{il,LV}$ and β_{fendo} ; but also by I_{Na} and $\sigma_{e,blood}$. Morphology-related parameters like skewness and kurtosis were also modulated by the surface-to-volume ratios in the LV, the RV and the fast endocardial layer, and by I_{Na} and $\sigma_{il,LV}$. Changes in $\sigma_{e,lung}$ have some effects on QRS kurtosis in some of the patients, but their effects are not as notable as those provoked by the parameters previously mentioned. Regarding the time interval between the QRS peak and the peak of the T-wave, the parameters exerting the strongest influence were those affecting cell repolarization in the six patients: I_{CaL} , τ_f , I_{Ks} and τ_{xs} .

T-wave Morphology As shown in Figure 7, the temporal delay of the maximum peak of the T-wave, in absolute value, was mainly determined by ionic changes affecting cell repolarization, as occurred with the QRS-T interval: I_{CaL} , τ_f , I_{Ks} and τ_{xs} . The main modulators of the amplitude of the T-wave peak were the delayed rectifier potassium currents (I_{Kr} and I_{Ks}), as well as the sodium current (I_{Na}) and conduction properties in both LV (β_{LV} , $\sigma_{il,LV}$, $\sigma_{el,LV}$ and β_{fendo}) and muscle in the torso ($\sigma_{e,musc}$). T-wave skewness was mainly determined by the values of β_{LV} , β_{RV} , I_{Na} , I_{Kr} , and τ_f . Similarly, T-wave kurtosis was modulated by β_{LV} , I_{Kr} , and τ_f . Interestingly, the biphasic behavior of the T-wave in some ECG leads in patient 6 led to large variations in both skewness and kurtosis in the sensitivity analysis (see Figure 7). Only in this patient, another parameter played a role in the T-wave morphology ($\sigma_{ec,LV}$).

4 Discussion

The sensitivity analysis performed in this study has shown that the tuning of very complex patient-specific models with a very high number of variables can be achieved by changing only a few parameters. Furthermore, the role of these key parameters was highly consistent between different patients. The goal of the analysis was to estimate the relative importance of parameters that cannot be observed clinically and, thus, the lack of counterintuitive results gives scientific evidence of what scientists would infer by intuition. As stated throughout the paper, about ten parameters were enough to tune both ECG morphology and LV activation maps. The low impact of the rest of the parameters was directly related to their smaller influence on myocardial electrical conduction or cellular action potential, as shown in previous studies [40]. The tuning procedure is performed manually, but the results presented in this study are an important step forward to create realistic human models in a more automated way in the future.

Sensitivity analyses for the study of human cardiac properties have been previously performed *in silico* in single cells or small tissues [7, 11, 20, 40, 42, 43, 44, 45, 46]. The influence of tissue extracellular conductivities on the ECG has been analyzed in a previous study using a human torso model fitted to a normal volunteer [19]. In this paper we have gone a step further and performed a sensitivity analysis in models specifically tailored to six HF patients undergoing CRT implantation. We have reproduced the ECG measured at the body surface and the intracardiac electrical activation map with high accuracy, although some features like the morphology of the T wave in leads V5 and V6 for patients 1, 3 and 6 presented some differences with the measured ECG.

Previous studies have used patient-specific models for the study of cardiac properties in order to both provide better predictions adapted to each patient and study underlying differences between them [1, 15, 23, 25, 29, 35]. The importance of conductivity parameters in the torso has also been a matter of study in recent works in both forward and inverse problems, focusing on different aspects like uncertainty in 3D visualization [8], anatomical segmentation [56], and realistic ECG simulation [4, 34, 51]. To the best of our knowledge, the work presented in this study is the first to be developed with complete models of the human torso, **ventricular anatomy**, and cellular electrophysiology adapted to specific patients.

A percentage of HF patients are diagnosed with LBBB. These patients present abnormally long QRS complexes with notching in some ECG leads [49]. These features required particular adjustments in the models, such as making the septum less conductive in our simulations. This finding is in agreement with clinical studies reporting an augmented transseptal time in these patients which does not necessarily involve slower propagation on the LV endocardium [2, 38, 50]. Notching was also favored by making LV cells more hypertrophic than those of the RV by decreasing β_{LV} as occurs in HF patients [16, 24, 32, 35, 57].

Two of the six patients included in this study presented scar in LV myocardium. These scarred areas were precisely located with CMR images and simulated in the models as non-excitabile tissue, but the severity of tissue damage could not be assessed during the procedure *in vivo*. There could still be some conduction pathways

that allow some electrical propagation, as observed in clinical data [21, 47]. Nevertheless, the models of the two patients matched with notable accuracy both the ECG and the LV activation pattern, and the parameters modulating them were very consistent with those obtained in the other four patients.

4.1 Limitations

The model developed by Ten Tusscher and Panfilov in 2006 used for simulating ventricular cell electrophysiology has been used in numerous studies because it is one of the most complete human ventricular action potential models up to date [26, 33, 36, 37, 55]. Newer electrophysiological models of ventricular myocytes have been recently developed based on particular experimental datasets, but their robustness in reproducing tissue, cellular, and sub-cellular behavior has been shown to be similar [10, 17, 30]. Furthermore, previous studies have shown that HF entails remodeling of cellular electrophysiology in the myocardium [52] and included it in cellular models [13, 54]. In this study we used an undiseased human ventricular cell model as a reference, since we could not be sure about either to what extent ventricular cells had modified their electrophysiology for each patient or, considering they had modified it, whether their remodeling was homogeneous in both ventricles. Therefore, proper model tuning for each patient is an essential part of the model creation procedure. As a result of the choice of a different cell model, quantification of sensitivities could vary, but qualitative effects of particular parameter changes would probably remain similar. **Moreover, the results of the sensitivity analysis performed in this study could not be directly extrapolated to patients with healthy hearts or other cardiac diseases.**

Changes applied to the models in the sensitivity analysis were applied individually and in most cases homogeneously. Global sensitivity analysis with simultaneous changes in model parameters could provide more detailed information about possible synergistic effects, but the associated computational cost with such a large set of parameters would be extremely large [9, 53]. Furthermore, changes in parameters applied locally and/or heterogeneously could help in understanding properties of localized portions of tissue within the heart (low perfusion areas, scarred myocardium...). **Our models contained various intrinsic heterogeneities in ionic parameters related to repolarization. Based mostly on data from animal hearts, this heterogeneity was implemented in subendocardial, midmyocardial, and subepicardial layers with differences between the LV and RV [55]. In addition we used a G_{Ks} gradient related to the activation sequence. A recent study by Opthof et al. in 3 explanted normal human hearts has shown quite different patterns of heterogeneity [31]. It cannot be excluded that some results of our sensitivity analysis would have been different in the presence of these different baseline patterns.**

The electroanatomical models specifically adapted to each patient were not guaranteed to be unique since other combinations of tuned parameters could possibly lead to matching of similar quality. However, in this study we focused on quantifying the variations with respect to the default model, therefore the mechanisms involved in the modulation of the ECG and LV activation would not change. The number of patients used in the analysis was relatively small, but the high inter-patient consistency

of the results suggests similar results would be obtained with an increased number of patients.

5 Conclusion

The sensitivity analysis shows that the effects of parameter variations on both ECG and LV activation are highly consistent between patients with specific anatomies and prognoses. Changes in ionic properties entail similar effects in all ECG leads, whereas the effects of changes in tissue conduction properties may vary between leads. Results of the analysis suggest that, in addition to the location of early activation sites, about 10 parameters suffice to create an accurate patient-specific model. Alterations in the remaining parameters are not significantly reflected in the ECG or in the ventricular electrical activation.

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