

## **Point-of-Care Cepheid Xpert HIV-1 Viral Load Test in Rural African Communities Is Feasible and Reliable**

Sikhulile Moyo, Terence Mohammed, Kathleen Wirth, Melanie Prague, Kara Bennett, Molly Holme, Lucy Mupfumi, Philemon Sebogodi, Natasha Moraka, Corretah Boleo, et al.

► **To cite this version:**

Sikhulile Moyo, Terence Mohammed, Kathleen Wirth, Melanie Prague, Kara Bennett, et al.. Point-of-Care Cepheid Xpert HIV-1 Viral Load Test in Rural African Communities Is Feasible and Reliable. *Journal of Clinical Microbiology*, American Society for Microbiology, 2016, 54 (12), pp.3050 - 3055. <10.1128/JCM.01594-16>. <hal-01579540>

**HAL Id: hal-01579540**

**<https://hal.inria.fr/hal-01579540>**

Submitted on 31 Aug 2017

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Point-of-Care Cepheid Xpert HIV-1 Viral Load Test in Rural African Communities Is Feasible and Reliable

Sikhulile Moyo,<sup>a,b</sup> Terence Mohammed,<sup>a</sup> Kathleen E. Wirth,<sup>c,d</sup> Melanie Prague,<sup>e</sup> Kara Bennett,<sup>f</sup> Molly Pretorius Holme,<sup>a,e</sup> Lucy Mupfumi,<sup>a</sup> Philemon Sebogodi,<sup>a</sup> Natasha O. Moraka,<sup>a</sup> Corretah Boleo,<sup>a</sup> Comfort N. Maphorisa,<sup>a</sup> Boitumelo Seraise,<sup>a</sup> Simani Gaseitsiwe,<sup>a,d</sup> Rosemary M. Musonda,<sup>a,d</sup> Erik van Widenfelt,<sup>a,d</sup> Kathleen M. Powis,<sup>a,d,g</sup> Tendani Gaolathe,<sup>a</sup> Eric J. Tchetgen Tchetgen,<sup>c,e</sup> Joseph M. Makhema,<sup>a,d</sup> Max Essex,<sup>a,d</sup> Shahin Lockman,<sup>a,d,h</sup> Vladimir Novitsky<sup>a,d</sup>

Botswana-Harvard AIDS Institute Partnership, Gaborone, Botswana<sup>a</sup>; University of Stellenbosch, Division of Medical Virology, Faculty of Medicine & Health Sciences, Tygerberg, South Africa<sup>b</sup>; Harvard T. H. Chan School of Public Health, Department of Epidemiology, Boston, Massachusetts, USA<sup>c</sup>; Harvard T. H. Chan School of Public Health, Department of Immunology and Infectious Diseases, Boston, Massachusetts, USA<sup>d</sup>; Harvard T. H. Chan School of Public Health, Department of Biostatistics, Boston, Massachusetts, USA<sup>e</sup>; Bennett Statistical Consulting, Inc., Ballston Lake, New York, USA<sup>f</sup>; Massachusetts General Hospital, Departments of Medicine and Pediatrics, Boston, Massachusetts, USA<sup>g</sup>; Brigham and Women's Hospital, Division of Infectious Diseases, Boston, Massachusetts, USA<sup>h</sup>

**Routine monitoring of HIV-1 RNA or viral load (VL) in patients on antiretroviral therapy (ART) is important, but there are multiple impediments to VL testing in resource-constrained settings. An accurate point-of-care (POC) HIV-1 VL test could alleviate many of these challenges. We compared the performance of the Cepheid Xpert HIV-1 VL assay against the laboratory-based Abbott *m2000sp/m2000rt* assay (Abbott assay). ART-naïve individuals participating in the Botswana Combination Prevention Project in 20 communities provided EDTA-blood specimens during household surveys. Both the POC Xpert HIV-1 VL and Abbott assays were performed on specimens sampled from 277 individuals. We found a high correlation between the Xpert HIV-1 VL and Abbott assay results ( $r^2 = 0.92$ ;  $P < 0.001$ ). The overall mean difference in the HIV-1 RNA values obtained by Xpert HIV-1 VL assay and Abbott assay was  $0.34 \log_{10}$  copies/ml (95% confidence interval [CI], 0.26 to  $0.40 \log_{10}$  copies/ml) ( $P < 0.001$ ). Using a clinically relevant level of 1,000 copies/ml as a threshold, agreement was 90.6% (95% CI, 87.9 to 93.1%), with a sensitivity of 98.6% (95% CI, 97.2 to 100%). The two methods agreed on their detectability of HIV-1 RNA ( $>40$  copies/ml) at 97.1% (95% CI, 95.5 to 98.7%), with a sensitivity of 99.6% (95% CI, 97.2 to 100%). The POC Cepheid Xpert HIV-1 VL assay showed high agreement and accuracy with a laboratory-based method of HIV-1 RNA testing. The POC Xpert HIV-1 VL assay tended to overestimate HIV-1 VL, although the difference was below a clinically relevant threshold of  $0.5 \log_{10}$  copies/ml. The POC Cepheid Xpert HIV-1 VL assay is a promising tool for monitoring patients on ART in southern Africa.**

Virologic monitoring of patients on antiretroviral therapy (ART) is a cornerstone of effective clinical care for the individuals, helping to monitor and support adherence, decrease drug resistance, and minimize costs and adverse clinical outcomes associated with unnecessary switching of antiretroviral (ARV) regimens. Botswana has successfully initiated antiretroviral therapy in over 87% of the citizens living with HIV (1). The magnitude of this scale-up necessitates routine HIV-1 RNA testing for monitoring treatment failure and adherence (2–5).

An increasing number of resource-poor countries are including viral load testing within their revised guidelines, a policy which conforms with the World Health Organization's recommendations for universal testing and treatment (6, 7). In resource-limited settings, especially in rural areas, laboratory-based HIV-1 RNA testing remains challenging for many reasons, among them insufficient access to laboratory facilities, cold-chain management shortcomings, and inability to consistently provide for sample transportation (8–10). Delays in the reporting of results may also negatively impact the HIV treatment cascade (11–13). Point-of-care (POC) viral load testing has the potential to alleviate these challenges, particularly in rural and remote communities.

Despite the development of several POC viral load technologies over the last decade (14–24), there is a shortage of commercially available tests. In this study, we evaluated the performance of the POC Cepheid Xpert HIV-1 viral load test in rural communities in Botswana in order to provide additional data to policy

makers regarding the use of this assay in decentralized HIV treatment programs. The results obtained by the Xpert HIV-1 viral load assay were compared to those obtained by the laboratory-based Abbott *m2000sp/m2000rt* assay.

## MATERIALS AND METHODS

**Study participants.** This study was performed as a cross-sectional sub-study within the Botswana Combination Prevention Project (BCPP; registration no. NCT01965470 at ClinicalTrials.gov). The overall design of BCPP is described elsewhere (1). Aggregate viral load data, pooled across arms, on study participants are presented here. A total of 302 ART-naïve HIV-infected individuals residing in 20 rural communities in Botswana (and taking part in BCPP) provided written in-

Received 5 August 2016 Returned for modification 23 August 2016

Accepted 5 October 2016

Accepted manuscript posted online 12 October 2016

**Citation** Moyo S, Mohammed T, Wirth KE, Prague M, Bennett K, Holme MP, Mupfumi L, Sebogodi P, Moraka NO, Boleo C, Maphorisa CN, Seraise B, Gaseitsiwe S, Musonda RM, van Widenfelt E, Powis KM, Gaolathe T, Tchetgen Tchetgen EJ, Makhema JM, Essex M, Lockman S, Novitsky V. 2016. Point-of-care Cepheid Xpert HIV-1 viral load test in rural African communities is feasible and reliable. *J Clin Microbiol* 54:3050–3055. doi:10.1128/JCM.01594-16.

**Editor:** M. J. Loeffelholz, University of Texas Medical Branch

Address correspondence to Sikhulile Moyo, sikhulilemoyo@gmail.com.

Copyright © 2016, American Society for Microbiology. All Rights Reserved.

formed consent and took part in the study. The Botswana institutional review board (IRB) (Human Resource Development Council [HRDC]) and CDC IRB approved the BCPP study as well as this POC HIV-1 RNA assessment substudy.

**POC testing of HIV-1 RNA.** Venous blood was collected in EDTA tubes in households (during the household survey) from September 2015 to May 2016. The collected blood was transported to a mobile clinic (portacamp) that was stationed in the community, and the blood was processed within 4 h of collection. The POC Xpert HIV-1 viral load testing was also performed in these mobile community-based clinics. One milliliter of plasma was used as input in the quantitative POC viral load test, the Cepheid Xpert HIV-1 viral load (catalog no. GXHIV-VL-CE-10), according to the manufacturer's instruction. The Xpert HIV-1 viral load is a test that automates the test process, including RNA extraction, purification, reverse transcription, and cDNA real-time quantification in a single cartridge within 90 min. The quantification range of the Xpert HIV-1 viral load test is 40 to 10,000,000 copies/ml ( $1.6 \log_{10}$  to  $7.0 \log_{10}$ ).

**Laboratory-based testing of HIV-1 RNA load.** The Abbott *m2000sp/m2000rt* assay (Abbott assay) was used as a comparator for evaluation of the Xpert HIV-1 viral load test. The Abbott assay was performed at the Botswana-Harvard AIDS Institute Partnership HIV Reference Laboratory (BHHRL) in Gaborone, Botswana, according to the manufacturer's instructions. The BHHRL was accredited by the South African National Accreditation System for HIV-1 viral load testing and maintains certification in Rush University's Virology Quality Assurance Program (25). The minimum level of HIV-1 RNA detection by Abbott assay is the same as that by POC Xpert HIV-1 viral load assay, 40 copies/ml (or  $1.6 \log_{10}$  copies/ml).

**Specimen flow.** A total of 302 participants were enrolled in this study. No data were generated for six samples due to quality control (QC)-related errors during POC testing. Three of these errors were related to internal quality control failure, and the other 3 were related to hardware error. A subset of 19 samples were tested by a single assay due to a limited specimen volume. A total of 277 specimens were tested by both the POC test and Abbott assay.

**Statistical methods.** We assume that missing data (insufficient plasma volume and QC errors) were missing completely at random. Correlation analysis was performed using the Spearman rank test. To determine differences between two viral load assays, we used the Bland-Altman plot method. The concordance was considered good when the differences are within the limits of two standard deviations of the mean. Passing-Bablok regression was used to test for systematic and proportional differences using the "BlandAltmanLeh" and "mcr" packages in R 3.3.0 (26–28). Passing-Bablok regression is insensitive to outliers and assumes that measurement errors in both variables have the same distribution. For all analysis, 95% confidence intervals are computed using bootstrapping to account for clustering within communities. We used 1,000 replicates where communities were sampled with replacement. The clinically relevant difference between two viral load measurements was considered at  $0.5 \log_{10}$  copies/ml, as described previously (29–31). We used the Cohen's kappa coefficient, a statistic which measures interrater agreement for qualitative values between two assays. It is considered to be a more robust measure than simple percent agreement calculation, since it takes into account the agreement occurring by chance. The kappa coefficient indicates a satisfying agreement between 0.6 and 0.8, and an excellent agreement above 0.8.

It should be noted that assay comparison implies a comparison of results obtained by POC Xpert HIV-1 VL assay performed in the mobile clinic to those obtained by the laboratory-based Abbott *m2000sp/m2000rt* assay, and it does not imply comparison of the assays. Each assay was performed in a natural setting, so there are substantial differences between the performance environments of the two assays (different conditions, methods, times, personnel, etc.).

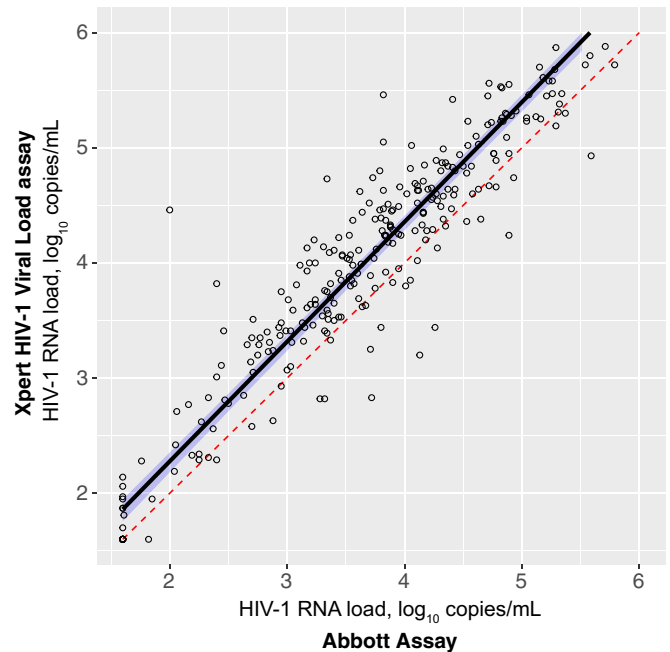


FIG 1 Passing-Bablok regression plot comparing POC Cepheid Xpert HIV-1 viral load assay against the Abbott *m2000sp/m2000rt* assay.

## RESULTS

Results from both the POC Xpert HIV-1 viral load test and the Abbott *m2000sp/m2000rt* assay were available for 277 participants. We observed a significant correlation between the HIV-1 RNA levels obtained by these two assays, using the nonparametric Spearman rank test ( $r^2 = 0.92$ ;  $P < 0.001$ ). With Pearson correlation, the results of the two assays also correlated highly linearly ( $r = 0.94$ ;  $P < 0.001$ ; Fig. 1). In Fig. 1, the estimated regression line (shown in blue) is compared to the diagonal dashed line (shown in red) that represents a hypothetically ideal match between two assays. Using Passing-Bablok regression, we observed significant systematic bias (intercept = 0.20; 95% confidence interval [CI], 0.05 to 0.32) and proportional bias (slope = 1.04; 95% CI, 1.02 to 1.07).

The similarities between the two assays were evaluated by the Bland-Altman plot method (Fig. 2). The overall mean difference in the HIV-1 RNA values obtained by POC and Abbott assay was  $0.34 \log_{10}$  copies/ml (95% CI, 0.16 to  $0.51 \log_{10}$  copies/ml) (overall POC mean [standard deviation {SD}],  $3.98 \log_{10}$  copies/ml [ $1.10 \log_{10}$  copies/ml] versus overall Abbott mean [SD],  $3.64 \log_{10}$  copies/ml [ $1.05 \log_{10}$  copies/ml]). Although statistically significant ( $P < 0.001$ ), the mean difference between the POC and Abbott assays was below the generally accepted clinically relevant difference of  $0.5 \log_{10}$  copies/ml (29–31). A significant difference between the two assays (i.e.,  $>2$  standard deviations of the mean) was observed for 4.7% ( $n = 13$ ) of the samples.

Using a threshold of 40 copies/ml (i.e.,  $1.6 \log_{10}$  copies/ml) for detectable HIV-1 RNA load, the two assays were in agreement at 97.1% (95% CI, 95.5 to 98.7%; Table 1), with a sensitivity of 99.6% (95% CI, 99.2 to 100%). Among the 7 individuals with detectable viral load using the POC Xpert HIV-1 viral load test ( $\geq 40$  copies/ml) and undetectable viral load using the Abbott test ( $< 40$  copies/ml), the mean  $\log_{10}$  HIV-1 RNA load (Q1, Q3) was 1.94 (1.87,

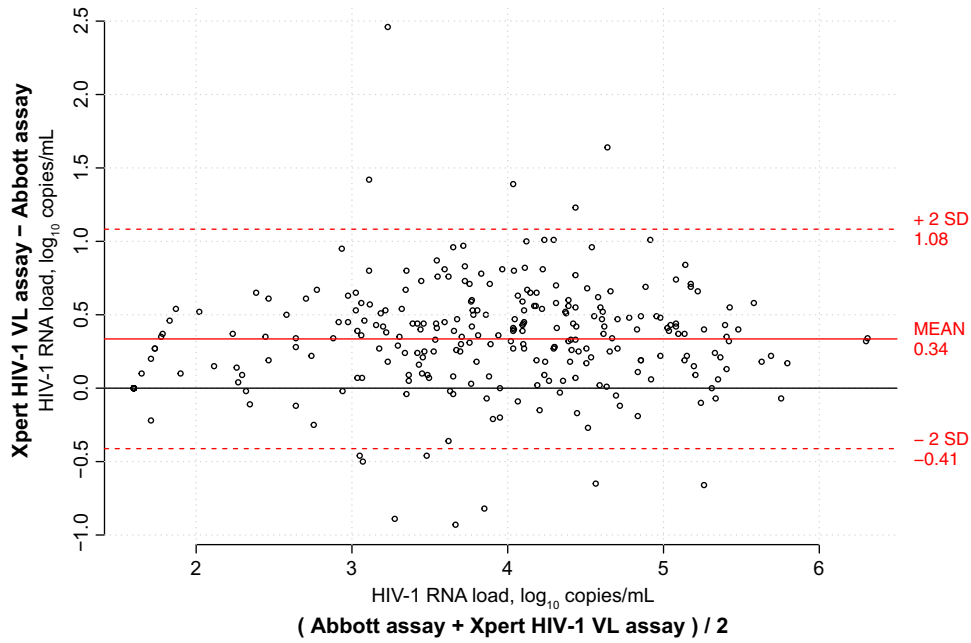


FIG 2 Bland-Altman plot comparing POC Cepheid Xpert HIV-1 viral load assay against the Abbott *m2000sp/m2000rt* assay.

2.01). This corresponded to a mean difference of 0.34 log<sub>10</sub> copies/ml with detectability threshold of 1.6 log<sub>10</sub>, which is not clinically relevant. Only one individual had an HIV-1 RNA load higher than 0.5 log<sub>10</sub> of the detectability limit. The only individual with undetectable POC Xpert HIV-1 viral load test had an Abbott quantification of 1.82 log<sub>10</sub> HIV-1 RNA load.

Using a threshold of 1,000 copies/ml (i.e., 3.0 log<sub>10</sub> copies/ml) for clinical monitoring of ART, the assays were in agreement for 90.6% (95% CI, 87.9 to 93.1%; Table 1), with a sensitivity 98.6% (95% CI, 97.2 to 100%). Among the 23 individuals with discordant results and high viral load with the POC Xpert HIV-1 viral load assay (Table 1), the mean (Q1, Q3) difference between the two tests was 0.66 log<sub>10</sub> copies/ml (0.44, 0.66 log<sub>10</sub> copies/ml). This difference was higher than 0.5 log<sub>10</sub> in 14 individuals (61%). Among the 3 individuals with discordant results and lower viral load with the POC Xpert HIV-1 viral load assay, the differences were 0.46, 0.5, and 0.89 log<sub>10</sub> copies/ml. Further analysis showed that the agreement is higher than 80% at any threshold used.

To assess whether agreement remains constant or depends on any particular threshold level, we calculated the level of agreement between the two assays against a range of HIV-1 RNA load mea-

surements from 1.6 to 6.0 log<sub>10</sub> copies/ml (Fig. 3A). The highest level of disagreement between the two assays occurred for HIV-1 RNA load between 3.5 and 4.5 log<sub>10</sub> copies/ml. However, the level of agreement was >80% across thresholds. Figure 3B shows the Cohen's kappa as a measure of agreement between POC and Abbott assays for different thresholds of the HIV RNA load. For all nonextreme HIV RNA load values, the Cohen's kappa indicates satisfying (above 0.6) or excellent (above 0.8) agreement between methods.

A sensitivity analysis was performed to further investigate differences of agreement between the two assays at a threshold of 3.0 log<sub>10</sub> copies/ml (1,000 copies/ml). For low HIV-1 RNA load ( $\leq 3.0$  log<sub>10</sub> copies/ml), statistically significant systematic (intercept = -0.63; 95% CI, -0.76 to -0.49) and proportional biases (slope = 1.39; 95% CI, 1.31 to 1.48) were observed (Fig. 4A). For high HIV-1 RNA load ( $> 3.0$  log<sub>10</sub> copies/ml), significant systematic bias (intercept = 0.36; 95% CI, 0.04 to 0.62) but no proportional bias (slope = 1.0; 95% CI, 0.95 to 1.07) was found (Fig. 4B). Overall, the POC assay tended to overestimate HIV-1 RNA load compared to the Abbott assay. In one case, HIV-1 RNA load was determined to be undetectable by POC assay but was borderline detectable by Abbott assay at 66 copies/ml.

TABLE 1 Agreement on detection of HIV-1 RNA between POC Cepheid Xpert HIV-1 viral load test and the Abbott *m2000sp/m2000rt* assay at two detection levels, 1.6 log<sub>10</sub> copies/ml and 3.0 log<sub>10</sub> copies/ml

Detection threshold for Xpert HIV-1 VL assay	Abbott assay detectability by HIV-1 RNA level			
	1.6 log <sub>10</sub> copies/ml		3.0 log <sub>10</sub> copies/ml	
	No. undetectable	No. detectable	No. undetectable	No. detectable
Undetectable	16	1	206	23
Detectable	7	253	3	45
Total	23	254	209	68

DISCUSSION

We found that POC Cepheid Xpert HIV-1 viral load testing was feasible in rural communities (after home-based collection of venous blood), and that the Xpert HIV-1 viral load results obtained in this setting correlated with the Abbott assay with a high level of agreement across a broad range of HIV-1 RNA values. We observed a slight overestimation of viral load by the Xpert HIV-1 viral load assay, which is consistent with other studies (21–24).

It is important to note that underestimation of HIV-1 RNA at low levels with the Abbott *m2000rt* assay was reported previously

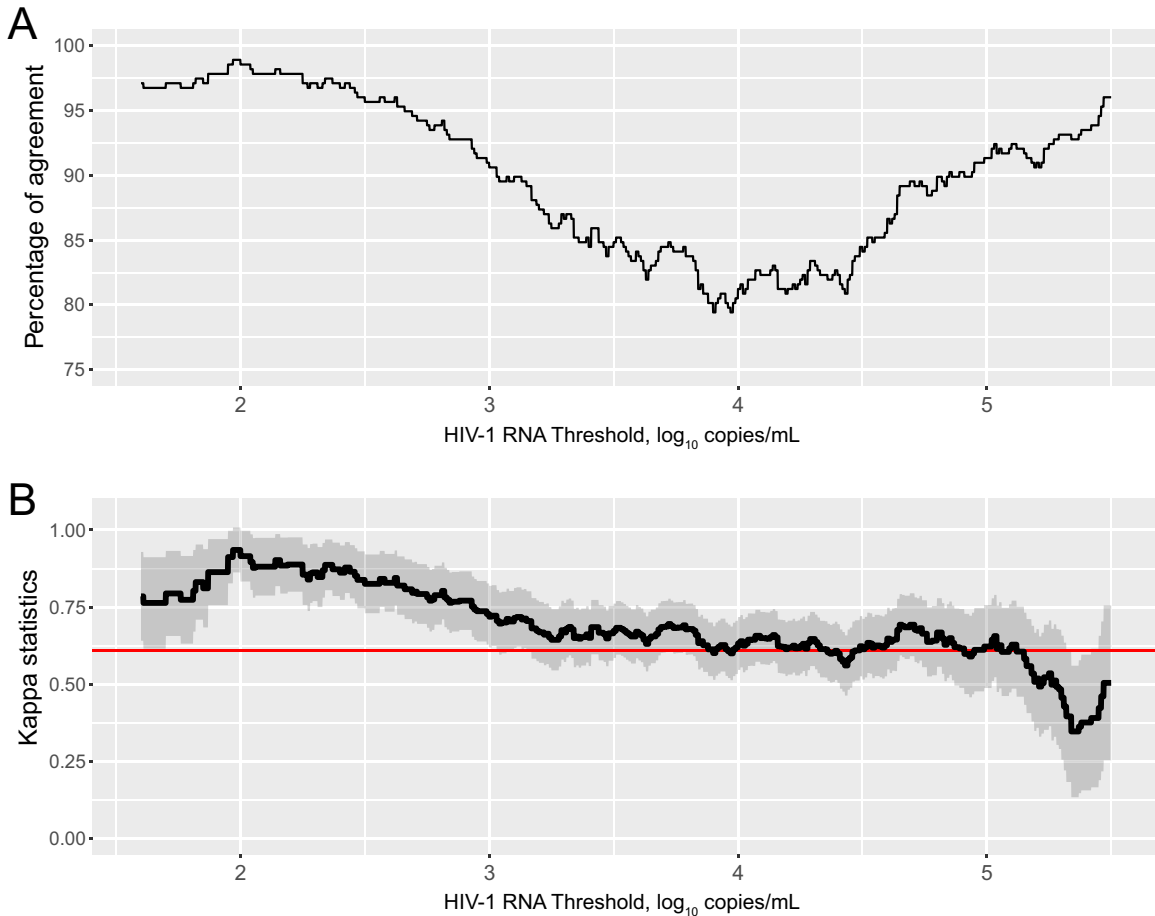


FIG 3 Level of agreement between POC Cepheid Xpert HIV-1 viral load assay and the Abbott *m2000sp/m2000rt* assay according to the viral load detection threshold. (A) Percentage of agreement between two assays. (B) Kappa statistics.

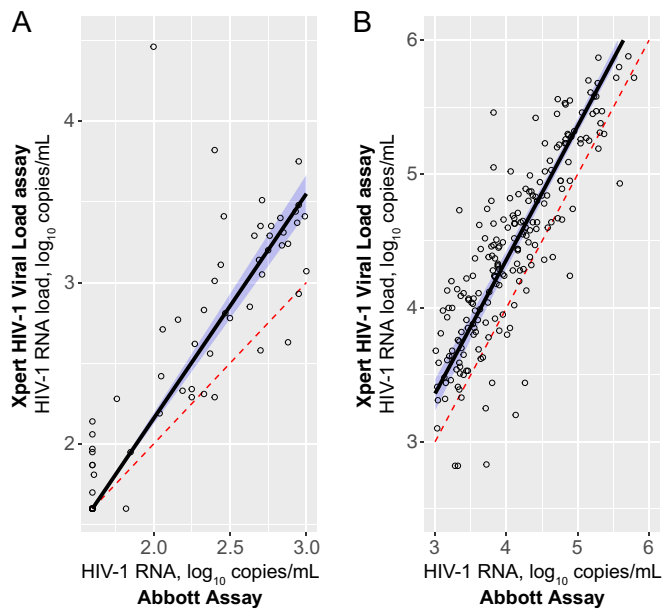


FIG 4 Passing-Bablok regression for POC Cepheid Xpert HIV-1 viral load assay and the Abbott *m2000sp/m2000rt* assay for low (≤3.0 log<sub>10</sub> copies/ml) (A) and high (>3.0 log<sub>10</sub> copies/ml) (B) HIV-1 viral load.

(29, 32). For example, while the overall correlation between the Abbott RealTime assay and the Roche Amplicor assay was 0.90, it was only 0.45 at a low level of HIV-1 RNA (32). According to a meta-analysis of the performance of HIV-1 RNA technologies, there is a tendency of the Abbott RealTime HIV-1 assay to underestimate HIV-1 RNA in plasma (29). Assuming that the Abbott assay underestimates HIV-1 RNA at low levels, it is possible that POC Xpert HIV-1 viral load results are more accurate at the low end of HIV-1 RNA distribution.

The primary goal of this study was to evaluate the performance of the Xpert HIV-1 viral load assay across the broad range of HIV-1 RNA values. In addition, we assessed the performance of the Xpert HIV-1 VL at two binary cutoffs, 40 copies/ml and 1,000 copies/ml. While we observed a high level of agreement across the range of more than 6 log<sub>10</sub> copies/ml, some values around the assessed thresholds were misclassified in binary analysis. Most mismatches were within 0.5 log<sub>10</sub>, which may be attributable to a natural variability associated with HIV-1 RNA measurements. Due to the small sample size around the assessed thresholds, the binary results obtained in this study should be taken cautiously. A dedicated study addressing the performance of the Xpert HIV-1 VL assay for the detection of HIV-1 RNA at the specified threshold among virologically suppressed individuals (e.g., individuals receiving ART) is warranted.



This study has some limitations. To preserve the integrity of data collected for an ongoing randomized trial, only limited aggregate information related to the characteristics of the study participants has been presented. To assess the performance of the Xpert HIV-1 VL test across a broad range of HIV-1 RNA values, we enrolled only ART-naive individuals. It might be necessary to evaluate the performance of the Xpert HIV-1 viral load assay among patients on ART in rural communities in southern Africa. This should assess the feasibility of the Xpert HIV-1 VL assay to be used for monitoring virologic suppression among individuals receiving ART. Finally, we used mobile clinics equipped with simple centrifuges for real-time processing of whole blood that was collected in households. However, this could be challenging in some settings where centrifugation facilities are not readily available. Finger-prick and whole-blood versions of the Xpert HIV-1 viral load assay are still under development. The current version of the Xpert HIV-1 viral load assay requires 1,000  $\mu$ l of plasma, and 19 samples did not have adequate volume to test using both platforms. This could be challenging for pediatric blood draws.

The major implementation lesson learned during the study is that quantification of HIV-1 RNA in rural southern Africa communities is feasible. Major challenges were related to initial steps, such as the collection of venous blood and its processing in a timely manner. This study benefitted from the extensive field network developed and maintained by the BCPP study (1). Nevertheless, smooth performance of the Xpert HIV-1 VL assay required proper training. This was evident from errors which occurred primarily at the beginning of the study, which could be attributable, at least partially, to the operator's mistake. In particular, a strict requirement that the input plasma volume for Xpert HIV-1 VL must be 1,000  $\mu$ l caused a few errors at the early stage of the study.

We conclude that Xpert HIV-1 viral load testing can be considered a feasible and reliable option for viral load monitoring, including in small rural communities in southern Africa. Same-day viral load could allow for immediate assessment of virologic failure, allow for triaging of patients for adherence counseling, and reduce loss to follow-up. The multiplexing functionality of the POC Cepheid Xpert assays could be an additional advantage for cost-effective diagnostics of other pathogens.

## ACKNOWLEDGMENTS

We thank the study participants and BCPP field study teams for making this study a success. We thank the BCPP team members for their contribution to this study: Tumulano Sekoto, Ngozana Seonyatseng, Unoda Chakalisa, Etienne Kadima Yankinda, One Pharathlathe, Vinoliah Simon, Rona Letlhogile, Atang Mbikiwa, Kutlo Manyake, Kutlwano Mukokomani, Mompoti Mmalane, Ria Madison, Chloe Auletta-Young, Data Management Centre Staff, and field laboratory assistants. We thank Lendsey Melton for excellent editorial assistance.

This study was supported by the U.S. President's Emergency Plan for AIDS Relief (PEPFAR) through the Centers for Disease Control and Prevention (CDC) under the terms of cooperative agreement U01 GH000447. Its contents are solely the responsibility of the authors and do not necessarily represent the official views of CDC. The funders had no role in the study design, data collection, analysis, interpretation, writing, or submission of the manuscript for publication.

## FUNDING INFORMATION

This work, including the efforts of Sikhulile Moyo, Terence Mohammed, Kathleen E. Wirth, Melanie Prague, Kara Bennett, Molly P. Holme, Natasha O. Moraka, Corettah Boleo, Comfort N. Maphorisa, Boitumelo Seraise, Simani Gaseitsiwe, Rosemary Musonda, Erik van Widenfelt, Kathleen M. Powis, Tendani Gaolathe, Eric J. Tchetgen Tchetgen, Joseph Makhema, M. Essex, Shahin Lockman, Vlad Novitsky, Lucy Mupfumi, and Philemon Sebogodi, was funded by U.S. President's Emergency Plan for AIDS Relief (PEPFAR) (U01 GH000447). This work, including the efforts of Simani Gaseitsiwe and Natasha O. Moraka, was funded by Wellcome Trust DELTAS Initiatives/Sub-Saharan Africa Network for TB/HIV Research Excellence (SANTHE) (107752/Z/15/Z).

This study was supported by the U.S. President's Emergency Plan for AIDS Relief (PEPFAR) through the Centers for Disease Control and Prevention (CDC) under the terms of cooperative agreement U01 GH000447. Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the CDC. The funders had no role in the study design, data collection, analysis, interpretation, writing, or submission of the manuscript for publication.

## REFERENCES

- Gaolathe T, Wirth KE, Holme MP, Makhema J, Moyo S, Chakalisa U, Yankinda EK, Lei Q, Mmalane M, Novitsky V, Okui L, van Widenfelt E, Powis KM, Khan N, Bennett K, Bussmann H, Dryden-Peterson S, Lebelonyane R, El-Halabi S, Mills LA, Marukutira T, Wang R, Tchetgen Tchetgen EJ, DeGruttola V, Essex M, Lockman S, Botswana Combination Prevention Project Study Team. 2016. Botswana's progress toward achieving the 2020 UNAIDS 90-90-90 antiretroviral therapy and virological suppression goals: a population-based survey. *Lancet HIV* 3:e221–e230. [http://dx.doi.org/10.1016/S2352-3018\(16\)00037-0](http://dx.doi.org/10.1016/S2352-3018(16)00037-0).
- Working Group on Modelling of Antiretroviral Therapy Monitoring Strategies in Sub-Saharan Africa, Phillips A, Shroufi A, Vojnov L, Cohn J, Roberts T, Ellman T, Bonner K, Rousseau C, Garnett G, Cambiano V, Nakagawa F, Ford D, Bansi-Matharu L, Miners A, Lundgren JD, Eaton JW, Parkes-Ratanshi R, Katz Z, Maman D, Ford N, Vitoria M, Doherty M, Dowdy D, Nichols B, Murtagh M, Wareham M, Palamountain KM, Chakanyuka Musanhu C, Stevens W, Katzenstein D, Ciaranello A, Barnabas R, Braithwaite RS, Bendavid E, Nathoo KJ, van de Vijver D, Wilson DP, Holmes C, Bershteyn A, Walker S, Raizes E, Jani I, Nelson LJ, Peeling R, Terris-Prestholt F, Murungu J, Mutasa-Apollo T, Hallett TB, Revill P. 2015. Sustainable HIV treatment in Africa through viral-load-informed differentiated care. *Nature* 528:S68–76. <http://dx.doi.org/10.1038/nature16046>.
- Marschner IC, Collier AC, Coombs RW, D'Aquila RT, DeGruttola V, Fischl MA, Hammer SM, Hughes MD, Johnson VA, Katzenstein DA, Richman DD, Smeaton LM, Spector SA, Saag MS. 1998. Use of changes in plasma levels of human immunodeficiency virus type 1 RNA to assess the clinical benefit of antiretroviral therapy. *J Infect Dis* 177:40–47. <http://dx.doi.org/10.1086/513823>.
- Thiébaud R, Morlat P, Jacqmin-Gadda H, Neau D, Mercie P, Dabis F, Chene G. 2000. Clinical progression of HIV-1 infection according to the viral response during the first year of antiretroviral treatment. Groupe d'Epidémiologie du SIDA en Aquitaine (GECSA). *AIDS* 14:971–978.
- Havlr DV, Bassett R, Levitan D, Gilbert P, Tebas P, Collier AC, Hirsch MS, Ignacio C, Condra J, Gunthard HF, Richman DD, Wong JK. 2001. Prevalence and predictive value of intermittent viremia with combination HIV therapy. *JAMA* 286:171–179. <http://dx.doi.org/10.1001/jama.286.2.171>.
- WHO. 2015. Consolidated guidelines on HIV testing services. World Health Organization, Geneva, Switzerland. [http://apps.who.int/iris/bitstream/10665/179870/1/9789241508926\\_eng.pdf?ua=1](http://apps.who.int/iris/bitstream/10665/179870/1/9789241508926_eng.pdf?ua=1).
- WHO. 2015. Guideline on when start antiretroviral therapy and on pre-exposure prophylaxis for HIV. World Health Organization, Geneva, Switzerland. [http://apps.who.int/iris/bitstream/10665/186275/1/9789241509565\\_eng.pdf?ua=1](http://apps.who.int/iris/bitstream/10665/186275/1/9789241509565_eng.pdf?ua=1).
- Bélec L, Bonn JP. 2011. Challenges in implementing HIV laboratory monitoring in resource-constrained settings: how to do more with less. *Future Microbiol* 6:1251–1260. <http://dx.doi.org/10.2217/fmb.11.121>.
- Ondoa P, Shamu T, Bronze M, Wellington M, Boender TS, Manting C, Steegen K, Luethy R, Rinke de Wit T. 2014. Performance and logistical

- challenges of alternative HIV-1 virological monitoring options in a clinical setting of Harare, Zimbabwe. *Biomed Res Int* 2014;102598. <http://dx.doi.org/10.1155/2014/102598>.
10. Ouma KN, Basavaraju SV, Okonji JA, Williamson J, Thomas TK, Mills LA, Nkengasong JN, Zeh C. 2013. Evaluation of quantification of HIV-1 RNA viral load in plasma and dried blood spots by use of the semiautomated Cobas Amplicor assay and the fully automated Cobas AmpliPrep/TaqMan assay, version 2.0, in Kisumu, Kenya. *J Clin Microbiol* 51:1208–1218. <http://dx.doi.org/10.1128/JCM.03048-12>.
  11. Lecher S, Ellenberger D, Kim AA, Fonjungo PN, Agolory S, Borget MY, Broyles L, Carmona S, Chipungu G, De Cock KM, Deyde V, Downer M, Gupta S, Kaplan JE, Kiyaga C, Knight N, MacLeod W, Makumbi B, Muttai H, Mwangi C, Mwangi JW, Mwasekaga M, Ng'Ang AL, Pillay Y, Sarr A, Sawadogo S, Singer D, Stevens W, Toure CA, Nkengasong J. 2015. Scale-up of HIV viral load monitoring—seven sub-Saharan African countries. *MMWR Morb Mortal Wkly Rep* 64:1287–1290. <http://dx.doi.org/10.15585/mmwr.mm6446a3>.
  12. Roberts T, Cohn J, Bonner K, Hargreaves S. 2016. Scale-up of routine viral load testing in resource-poor settings: current and future implementation challenges. *Clin Infect Dis* 62:1043–1048. <http://dx.doi.org/10.1093/cid/ciw001>.
  13. Rutstein SE, Golin CE, Wheeler SB, Kamwendo D, Hosseinipour MC, Weinberger M, Miller WC, Biddle AK, Soko A, Mkandawire M, Mwenda R, Sarr A, Gupta S, Mataya R. 2016. On the front line of HIV virological monitoring: barriers and facilitators from a provider perspective in resource-limited settings. *AIDS Care* 28:1–10. <http://dx.doi.org/10.1080/09540121.2015.1058896>.
  14. Cogswell HA, Ohadi E, Avila C. 2016. Viral-load point-of-care technologies to achieve an AIDS-free generation. *Future Microbiol* 11:5–9. <http://dx.doi.org/10.2217/fmb.15.121>.
  15. Maiers TJ, Gous N, Nduna M, McFall SM, Kelso DM, Fisher MJ, Palamoukian KM, Scott LE, Stevens WS. 2015. An investigation of fingerstick blood collection for point-of-care HIV-1 viral load monitoring in South Africa. *S Afr Med J* 105:228–231. <http://dx.doi.org/10.7196/SAMJ.7799>.
  16. Shafiee H, Kanakasabapathy MK, Juillard F, Keser M, Sadasivam M, Yuksekkaya M, Hanhauser E, Henrich TJ, Kuritzkes DR, Kaye KM, Demirci U. 2015. Printed flexible plastic microchip for viral load measurement through quantitative detection of viruses in plasma and saliva. *Sci Rep* 5:9919. <http://dx.doi.org/10.1038/srep09919>.
  17. Lee HH, Dineva MA, Chua YL, Ritchie AV, Ushiro-Lumb I, Wisniewski CA. 2010. Simple amplification-based assay: a nucleic acid-based point-of-care platform for HIV-1 testing. *J Infect Dis* 201(Suppl 1):S65–S72.
  18. Jani IV, Meggi B, Vubil A, Siteo NE, Bhatt N, Tobaiwa O, Quevedo JI, Loquiha O, Lehe JD, Vojnov L, Peter TF. 2016. Evaluation of the whole-blood Alere Q NAT point-of-care RNA assay for HIV-1 viral load monitoring in a primary health care setting in Mozambique. *J Clin Microbiol* 54:2104–2108. <http://dx.doi.org/10.1128/JCM.00362-16>.
  19. Ritchie AV, Ushiro-Lumb I, Edemaga D, Joshi HA, De Ruiter A, Szumilin E, Jendrulek I, McGuire M, Goel N, Sharma PI, Allain JP, Lee HH. 2014. SAMBA HIV semiquantitative test, a new point-of-care viral-load-monitoring assay for resource-limited settings. *J Clin Microbiol* 52:3377–3383. <http://dx.doi.org/10.1128/JCM.00593-14>.
  20. Scott L, Gous N, Carmona S, Stevens W. 2015. Laboratory evaluation of the Liat HIV Quant (IQuum) whole-blood and plasma HIV-1 viral load assays for point-of-care testing in South Africa. *J Clin Microbiol* 53:1616–1621. <http://dx.doi.org/10.1128/JCM.03325-14>.
  21. Garrett NJ, Drain PK, Werner L, Samsunder N, Abdool Karim SS. 2016. Diagnostic accuracy of the point-of-care Xpert HIV-1 viral load assay in a South African HIV clinic. *J Acquir Immune Defic Syndr* 72:e45–48. <http://dx.doi.org/10.1097/QAI.0000000000000978>.
  22. Ceffa S, Luhanga R, Andreotti M, Brambilla D, Erba F, Jere H, Mancinelli S, Giuliano M, Palombi L, Marazzi MC. 2016. Comparison of the Cepheid GeneXpert and Abbott *m2000* HIV-1 real time molecular assays for monitoring HIV-1 viral load and detecting HIV-1 infection. *J Virol Methods* 229:35–39. <http://dx.doi.org/10.1016/j.jviromet.2015.12.007>.
  23. Mor O, Gozlan Y, Wax M, Mileguir F, Rakovsky A, Noy B, Mendelson E, Levy I. 2015. Evaluation of the RealTime HIV-1, Xpert HIV-1, and Aptima HIV-1 Quant Dx assays in comparison to the NucliSens EasyQ HIV-1 v20 assay for quantification of HIV-1 viral load. *J Clin Microbiol* 53:3458–3465. <http://dx.doi.org/10.1128/JCM.01806-15>.
  24. Gueudin M, Baron A, Alessandri-Gradt E, Leme V, Mourez T, Etienne M, Plantier JC. 2016. Performance evaluation of the new HIV-1 quantification assay, Xpert HIV-1 viral load, on a wide panel of HIV-1 variants. *J Acquir Immune Defic Syndr* 72:521–526. <http://dx.doi.org/10.1097/QAI.0000000000001003>.
  25. HIV/AIDS Network Coordination. Virology quality assurance. HIV/AIDS Network Coordination, Seattle, WA. <https://www.hanc.info/labs/labresources/vqaResources/Pages/default.aspx>.
  26. R Core Team. 2014. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
  27. Lehnert B. 2015. BlandAltmanLeh: Plots (slightly extended). R package version 0.3.1. <http://cran.r-project.org/package=BlandAltmanLeh>.
  28. Manuilova E, Schuetzenmeister A, Model F. 2014. mcr: method comparison regression. R package version 1.2.1. <http://cran.r-project.org/package=mcr>.
  29. Sollis KA, Smit PW, Fiscus S, Ford N, Vitoria M, Essajee S, Barnett D, Cheng B, Crowe SM, Denny T, Landay A, Stevens W, Habiyambere V, Perrins J, Peeling RW. 2014. Systematic review of the performance of HIV viral load technologies on plasma samples. *PLoS One* 9:e85869. <http://dx.doi.org/10.1371/journal.pone.0085869>.
  30. Hughes MD, Johnson VA, Hirsch MS, Bremer JW, Elbeik T, Erice A, Kuritzkes DR, Scott WA, Spector SA, Basgoz N, Fischl MA, D'Aquila RT. 1997. Monitoring plasma HIV-1 RNA levels in addition to CD4<sup>+</sup> lymphocyte count improves assessment of antiretroviral therapeutic response. ACTG 241 Protocol Virology Substudy Team. *Ann Intern Med* 126:929–938.
  31. Saag MS, Holodniy M, Kuritzkes DR, O'Brien WA, Coombs R, Poscher ME, Jacobsen DM, Shaw GM, Richman DD, Volberding PA. 1996. HIV viral load markers in clinical practice. *Nat Med* 2:625–629. <http://dx.doi.org/10.1038/nm0696-625>.
  32. Swenson LC, Cobb B, Geretti AM, Harrigan PR, Poljak M, Seguin-Devaux C, Verhofstede C, Wirten M, Amendola A, Boni J, Bourlet T, Huder JB, Karasi JC, Zidovec Lepej S, Lunar MM, Mukabayire O, Schuurman R, Tomazic J, Van Laethem K, Vandekerckhove L, Wensing AM, International Viral Load Assay Collaboration. 2014. Comparative performances of HIV-1 RNA load assays at low viral load levels: results of an international collaboration. *J Clin Microbiol* 52:517–523. <http://dx.doi.org/10.1128/JCM.02461-13>.