

Parallel Reconfigurable Operator for Genomic Sequence Comparison: Architecture and Performance Analyses

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***Parallel Reconfigurable Operator for Genomic
Sequence Comparison: Architecture and
Performance analyses***

Alexandre Cornu — François Dussaughey — Dominique Lavenier

N° 6776

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Parallel Reconfigurable Operator for Genomic Sequence Comparison: Architecture and Performance analyses

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Abstract: This document describes a parallel genomic sequence comparison operator for reconfigurable platforms. It has been implemented for speeding up the time consuming parts of the IRISA-TBLASTN program designed for comparing full genomes with large protein banks. Speed-up ranging from 25 to 200 has been measured on real genomic data.

Key-words: Parallelisation, similarity search, indexing, TBLASTN

Opérateur reconfigurable parallèle pour la comparaison de séquences Génomiques : Architecture et analyse des performances

Résumé : Ce document décrit un opérateur parallèle de comparaison de séquences génomiques pour des plates formes reconfigurables. Il a été mis en oeuvre pour accélérer les parties critiques du programme IRISA-TBLASTN conçu pour comparer des génomes entiers avec de grandes banques de protéines. Des accélérations importantes allant de 25 à 200 ont été mesurées sur des données génomiques réelles.

Mots-clés : Parallélisation, recherche de similarité, indexation, TBLASTN

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1 Introduction

With the increasing amount of complete sequenced genomes [8], the need for rapidly mining these genomic data is becoming critical. One of the first task is often to compared newly sequenced genomes with protein banks to locate genes and perform a first annotation.

This task is generally done with sequence comparison tools such as the TBLASTN program of the BLAST family [6] [5]. Even if the BLAST programs are fast, analyzing large genomes can be very time consuming.

Our approach is a first attempt to parallelize BLAST-like programs on parallel platforms. Contrary to TBLASTN (from the BLAST family), this version doesn't aim to scan large databases. Instead, it focusses on comparing consequent amount of data, that is a complete genome and a protein bank [1].

Like TBLASTN, our program (called here IRISA-TBLASTN) uses a seed heuristic to limit the search space. It proceeds into 3 steps:

1. find anchor points;
2. compute ungapped extension;
3. compute gap extension using dynamic programming techniques.

The main difference is that instead of only indexing the query (sequences from the protein bank), IRISA-TBLASTN indexes the complet genome following the 6 reading frame and performs a dynamic indexing of the protein bank. Furthermore, the index is enhanced with neighborhood information to avoid costly random memory accesses.

The central idea is to merge the two first steps of TBLASTN into a single one for generating very rapidly significant ungapped alignments or, at least, positions between the genome and the sequences from the protein bank where a high probability of similarity occurs. In that scheme, the first part of the algorithm only manipulates indexes which represent only local view of the genomes. This is enough to determine if it is worth to run (or not) a costly dynamic programming procedure. The great advantage is that these local computations can be highly parallized, as it is shown in the next section.

This technique has been sucessfully tested through various implementations: SSE instructions, multithreading and GPU devices [2]. This document presents an FPGA implementation through a Parallel Sequence Comparison operator (PSC operator).

Briefly, this operator is located on a reconfigurable board connected through a PCIe interface. It is made of hundreds of small dedicated processors working in parallel for computing a simple score between two short protein subsequences. It output only the short sequences (or more specifically these positions) having a relatively good similarity.

The rest of the document is organized as follows: the next section presents the IRISA-TBALSTN algorithm. Section 3 details the Parallel Sequence Comparison (PSC) operator. The last section gives performance results.

2 IRISA-TBLASTN Algorithm

2.1 Index

The IRISA-TBLASTN algorithm requires first to index a genome (DNA sequence) into its 6 protein reading frames. This is done offline. Results are stored into two specific files:

- genome.dat: this file contains a sorted list of seeds with their neighborhood;
- genome.idx: this file contains a list of 20^W integers corresponding to the seed number of elements of size W .

As an example, suppose the following DNA sequence S:

```
S = A T G G A C C A G G A T A G G A C C A C G A G T A G
      |                   |                   |
      0                   10                  20
```

Translation into its 3 reading frames gives:

```
M D A D R T T S (frame 1)
W T R I G P R V (frame 2)
G P G - D H E - (frame 3)
```

The reverse strand of S is :

```
C T A C T C G T G G T C C T A T C C T G G T C C A T
      |                   |                   |
      20                  10                   0
```

Translation into its 3 reading frames gives:

```
L L V V V S W S (frame -1)
Y S W S Y P G P (frame -2)
T R G P I L V H (frame -3)
```

With a size seed equal to 1 amino acid ($W = 1$) and a neighborhood of 3 amino acids, we get the intermediate index:

frame 1	frame 2	frame 3
*** M DAD 0	*** W TRI 1	*** G PG- 2
**M D ADR 3	**W T RIG 4	**G P G-D 5
*MD A DRT 6	*WT R IGP 7	*GP G -DH 8
MDA D RTT 9	WTR I GPR 10	GPG - DHE 11
DAD R TTS 12	TRI G PRV 13	PG- D HE- 14
ADR T TS* 15	RIG P RV* 16	G-D H E-* 17
DRT T S** 18	IGP R V** 19	-DH E -** 20
RTT S *** 21	GPR V *** 22	DHE - *** 23

frame -1	frame -2	frame -3
*** L LVV -25	*** Y SWS -24	*** T RGP -23
**L L VVV -22	**Y S WSY -21	**T R GPI -20
*LL V VVS -19	*YS W SYP -18	*TR G PIL -17
LLV V VSW -16	YSW S YPG -15	TRG P ILV -14
LVV V SWS -13	SWS Y PGP -12	RGP I LVH -11
VVV S WS* -10	WSY P GP* -9	GPI L VH* -8
VVS W S** -7	SYP G P** -6	PIL V H** -5
VSW S *** -4	YPG P *** -3	ILV H *** -2

The final index is sorted following the seed alphabetical order:

genome.idx	genome.dat
A 1	0: *MD A DRT 6 24: DAD R TTS 12
C 0	1: **M D ADR 3 25: IGP R V** 19
D 3	2: MDA D RTT 9 26: **T R GPI -20
E 1	3: PG- D HE- 14 27: **Y S WSY -21
F 0	4: -DH E -** 20 28: RTT S *** 21
G 5	5: *** G PG- 2 29: VSW S *** -4
H 2	6: *GP G -DH 8 30: VVV S WS* -10
I 2	7: *TR G PIL -17 31: YSW S YPG -15
K 0	8: SYP G P** -6 32: ADR T TS* 15
L 3	9: TRI G PRV 13 33: DRT T S** 18
M 1	10: G-D H E-* 17 34: **W T RIG 4
N 0	11: ILV H *** -2 35: *** T RGP -23
P 5	12: WTR I GPR 10 36: GPR V *** 22
Q 0	13: RGP I LVH -11 37: LLV V VSW -16
R 4	14: *** L LVV -25 38: *LL V VVS -19
S 5	15: GPI L VH* -8 39: PIL V H** -5
T 4	16: **L L VVV -22 40: LVV V SWS -13
V 5	17: *** M DAD 0 41: *** W TRI 1
W 3	18: TRG P ILV -14 42: VVS W S** -7
Y 2	19: WSY P GP* -9 43: *YS W SYP -18
	20 :YPG P *** -3 44: *** Y SWS -24
	21: **G P G-D 5 45: SWS Y PGP -12
	22: RIG P RV* 16 46: GPG - DHE 11
	23: *WT R IGP 7 47: DHE - *** 23

The genome.idx file contains 20^W entries, corresponding to the 20^W different possible seeds of size W . For each seed, it indicates the number of seeds in the genome.dat file.

2.2 General Algorithm

When comparing a genome (already indexed) to a protein bank, the IRISA-TBLASTN algorithm can be split into 2 steps as described below:

```

1: for i=0 to MAX_SEED reset(INDEX_PROT[i])           // step 1
2: for all sequences sq in the protein bank
3:   for all seeds sd of sq
4:     add_index(INDEX_PROT[sd])
5:     if size(INDEX_PROT[sd]) = MAX_ELMT
6:       get_index(INDEX_GENOME,sd)
7:       ungapped_alignments(RESULTS,INDEX_GENOME,INDEX_PROT[sd])
8:       add_align(UNGAPPED,RESULTS)
9:       reset(INDEX_PROT[sd])
10:
11: for all elements x of UNGAPPED                     // step 2
12:   compute_align(x)

```

INDEX_PROT[k] is an index structure of limited size located into the main memory of the computer. It has the same structure as the genome index but, for a specific seed, it can contain a maximum of MAX_ELMT elements (one element is made of a seed, its neighborhood and its position into the genome).

The protein bank is scanned only once and an index is dynamically created (line 4). When, for a seed, the maximum value is reached (line 5), the corresponding genome seed index is get (line 6). From these two indexes, ungapped alignments are performed. Results are stored into the UNGAPPED structure which memorizes only alignments having exceeded a given threshold.

The second step consists in extending ungapped alignments by using more sophisticated techniques such as dynamic programming.

In this document, we focus only on step 1 which represents a large percentage of the overall execution time. More specifically, code profiling shows that the `ungapped_alignment` procedure (line 7) represents more than 98% of step 1.

2.3 Ungapped Alignment Procedure

The `ungapped_alignment` procedure takes as input 2 index structures: one from the genome and the other from the protein bank. Both indexes are related to the same seed. If N is the number of elements in the genome index and M the number of elements in the protein bank index, then the procedure computes $N \times M$ scores based on an amino acid substitution matrix. The pseudo C-code is the following:

```

void ungapped_alignment(RESULTS, INDEX_GENOME, INDEX_PROT)
{
  for all elements G of INDEX_GENOME {
    for all elements P of INDEX_PROT {
      score = 0;
      maxi = 0;
      for (i=0; i<SIZE_SEED+2*SIZE_NEIGHBORHOOD; i++) {
        score = score + SUB(G.seq[i],P.seq[i]);
        if (score<0) score = 0;
        if (maxi<score) maxi = score;
      }
    }
    if (maxi>=THRESHOLD) add_results(RESULTS,G.pos,P.pos);
  }
}

```

The notation `G.seq[i]` represents the i^{th} amino acid of the sequences made of the concatenation of the left neighborhood, the seed and the right neighborhood of the element `G`. Similarly, `G.pos` represents the position of the seed inside the genome. At the end of the procedure, the structure `RESULTS` contains a list of position pairs corresponding to the locations where significant similarities have been detected. From these *anchoring points* the second step can be launched.

2.4 Parallelism

From the code of the `ungapped_alignment` procedure it can be seen that the computations performed inside the 2 nested `for all` loops are independant. Thus, the $N \times M$ score calculations can be attached to a specific task and run independantly.

The parallel operator describes in the next section is based on this observation.

3 Parallel Sequence Comparison Operator

3.1 Functionality overview

The PSC operator aims to hardwire onto a reconfigurable platform the computation of the `ungapped_alignment` procedure. This operator receives two data flows, the `INDEX_GENOME` and `INDEX_PROT` structures and output pairs of positions corresponding to significant ungapped alignments.

3.2 Connection to the host processor

The operator is implemented on a reconfigurable board connected through a PCIe interface. Figure 1 details the way the input/output of the PSC operator I/O are linked.

The two input are connected to two input FIFOs which are fed by two **read actors**. These two components are initialized by the host processor to performed DMA transfers from the host memory.

Similarly, the output of the PSC operator is connected to a FIFO which feed a **write actor**. This device is also initialized by the host processor for DMA transfers.

In that scheme, data transfers are initiated by the host. The operator starts working as soon as data enter the input FIFOs. The operator stops when the last data is read from the input FIFOs. To detect the end of the process, the FIFO are enhanced with a data flag indicating the data status. Thus, when the last data is sent to the FIFOs, the read actor set this flag.

3.3 Operator Architecture

The PSC operator architecture has been designed to be able to drive data to and from a large number of processing elements (PE). All the PEs are working in a parallel way as a Simple Instruction Multiple Data architecture. Each PE computes the distance between 2 sub-sequences of amino-acids.

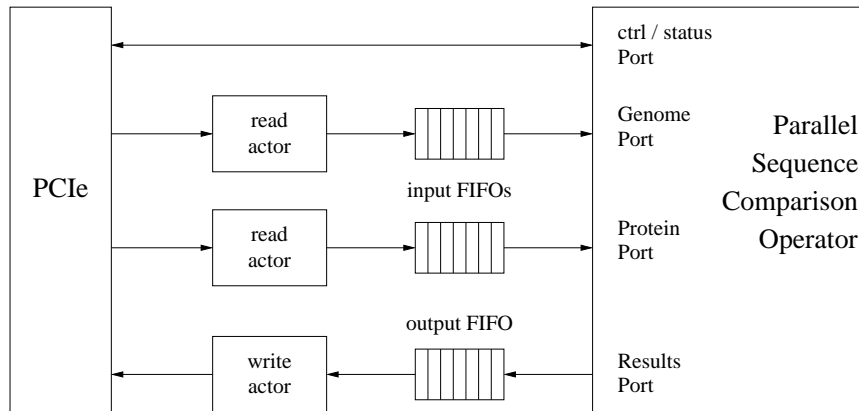


Figure 1: Connection of the PSC operator to the host processor through PCIe interface and read/write DMA actors

The design needs to run at 250 MHz to match PCIexpress frequency. This high frequency requires to prefer short and parallel data paths, rather than long and shared paths. This has led to develop a pipeline structure.

Indeed, PEs (or clusters of several PEs) are separated by register barriers, which delay and reinforce all signals (data and control signals). This structure, that delayed the computation process from a PE to the next one, makes the use of feedback signal impossible: PEs are entirely under the control of a master unit. Figure 2 represents this architecture.

The design is independant of the number of PEs. Here, this architecture permits to use a single PE for first validation (architecture simulation, software development and validation,...), then gradually increase the pipeline length and finally use a maximum of PE for highest acceleration (maximum PE number depends on FPGA ressources).

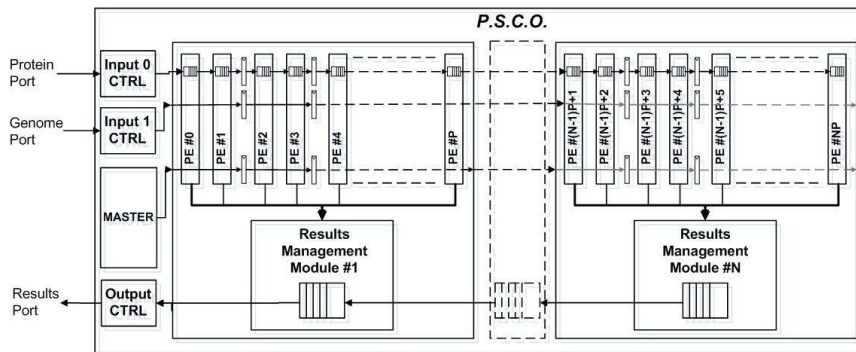


Figure 2: PSC Operator Architecture

PSC operator architecture is divided into the following components:

- *Input Controller 0*: reads query sub-sequences on protein port 0 and pushes them in the data pipeline 0.

- *Input Controller 1*: reads indexed genome sub-sequences on genome port and pushes them in the data pipeline 1.
- *PE Slots*: PE Slots consists of groups of PE with a common result management module. The factorization of Result management modules enable to save FPGA resources. PE Slots are made of:
 - *PE*: Processing Element that stores a query sub-sequences in order to reuse this data as much as possible and computes distance between stored query sub-sequences and new genome sub-sequences (see Processor Description section).
 - *Result Management Module*: scan results from a cluster of PE and store them into a Fifo if the distance computation result is higher than a predefined threshold. These Fifo are cascaded so that data join the Output Controller.
- *Output Controller*: reads results from cascaded Fifo and writes them on Result port.
- *Master controller*: manages the global process (process start, data loading, computation, results recovering, end of computation).

Each Input Controller has a Sequence Counter, so that a PE, knowing its position in the pipeline, can deduce the sequence identifier from this counter value. Results returned to the host consists of 32-bit-words of two sequence identifiers and some warning flags.

3.4 Processor Description

Figure 3 represents the PE architecture. First, during the load process, it stores a query sub-sequence into a shift-register Fifo. The feedback loop kept that sequence stored so that it can be reused for several computations. Then, during computation process, this sequence (query) is sent AA by AA to the processor, which computes a distance against every single sequence coming from the Genome Port controller (seq bank).

During a computation (N clock cycles for a computation between 2 sequences of size N), a distance between each couple of AA comes from a local RAM initialized with BLOSUM matrix values. These value are added. The maximum is stored as a result, which is then compared to a predefined threshold value by the Result Management module, and finally kept or thrown away.

4 Performance evaluation

4.1 Hardware platform

The reconfigurable platform is an XpressFX Card from PLDA, which has a 8 lanes PCIexpress interface giving a theoretical bandwidth of 2.5GBytes/s. The main components of the XpressFX card are:

- a FPGA: VIRTEX 4 XC4FX100 FF1152 from Xilinx;

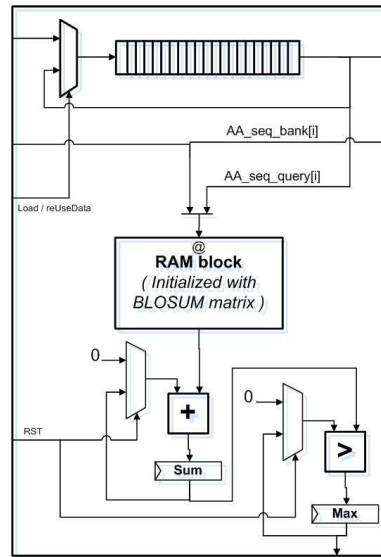


Figure 3: Processing Element Architecture



Figure 4: XpressFX Card from PLDA

- a couple of 512 MBytes DDR-SDRAM.

The XpressFX card is plugged in a X86 workstation having the following features:

- Processor : Intel(R) Pentium(R) D CPU 2.80GHz
- Processor Cache : 1 Mo
- Memory : 3 Go

- Operating System : Red Hat Enterprise Linux AS release 4

4.2 Data Set

The data set has been chosen according to a real bioinformatics application dealing with mitochondrial diseases [4]. The start of the study was the comparison of the human genome with 600,000 procaryotic proteins. Here, we select 5 protein subsets as follows:

- P1K: 1000 proteins (300,005 amino acids)
- P3K: 3000 proteins (811,411 amino acids)
- P10K: 10,000 proteins (2,984,635 amino acids)
- P30K: 30,000 proteins (6,868,965 amino acids)

The chromosomes of the human genome come from the Golden Database (NCBI Build 36.1, Mar. 2006 – hg18). We chose respectively the chromosomes 21, 9 and 1 as they represent short medium and long human chromosomes:

- Chr 21 : 46 944 323 nucleotides
- Chr 9 : 140 273 252 nucleotides
- Chr 1 : 247 249 719 nucleotides

4.3 Experiments

Experiments consist in measuring the execution time with and without the PSC operator using the different data sets. In the next subsections the time is reported in second and measured with the linux command `time` (there are no more jobs running on the platform).

Tests have been performed for the 3 chromosomes againts all the protein sets using a 256-processor PSC operator. For each experiment the software and the hardware versions have been run in the same condition in order to generate strickly identical results.

Chr21, Chr9 and Chr1 vs P1K

chr	host	PSC	speed-up
21	1603	49	32
9	4728	145	32
1	8758	320	26

Chr21, Chr9 and Chr1 vs P3K

chr	host	PSC	speed-up
21	4143	51	82
9	12266	143	85
1	21632	328	66

Chr21, Chr9 and Chr1 vs P10K

chr	host	PSC	speed-up
21	15144	69	219
9	45545	343	132
1	80755	696	116

Chr21, Chr9 and Chr 1 vs P30K

chr	host	PSC op	speed-up
21	34590	169	217
9	103878	753	138
1	180360	1441	125

4.4 Discussion

The main point is that larger the data set, better the speed-up. This is actually the situation where our implementation is supposed to give the best performance. When the protein data set is too small, the PSC operator is not used at its maximal capacity: all the processor cannot be fed with data. The best performances are achieved when the protein data sets exceed 10 000 sequences.

However, we note that small chromosomes provide extremely good performances compared to medium and large ones. Another way to interpret the results is that performances on medium and large chromosomes are not as promising as they should be! Investigations have shown that the architecture behaviour of the PSC operator is sensitive to burst of results: when many processors report simultaneously good similarity between pairs of protein sequences, the FIFO chain allowing to propagate the results to the output port can saturate, leading to the lost of some elements. This failure is detected by hardware and corrected by software. The underlying hypothesis is that this situation can exceptionally raise. Actually, the probability to detect burst of similarity on real genomic data is much higher than tests performed on synthetic data. This is a real weakness of this architecture which could be corrected by resizing the FIFO (but the problem is thus postponed for larger data sets) or by freezing temporarily the operator when such a bottleneck is detected.

A complementary explanation is that for large data set, the PCIe interface cannot sustain both the input and output flows, leading to the FIFO bottleneck previously explained. In that case, the solution could be to add a local memory on the board and to use it both:

- as a cache for storing the most often used data from the host in order to lower down the PCIe traffic;
- as a temporary tank for storing results as soon as they are available from the PSC operator output, and only triggering a PCIe transfer (from this memory) during periods of low PCIe activity.

The last point which needs to be highlighted is the flexibility of the PSC operator. Whatever the number of processors, the global control and the clock frequency remain the same. The software needs only to be parametrized by the number of processor for sending packets of data.

Finally, we also have to stress out that this operator doesn't bring performances for all the genomic sequence comparison process. It only focuses on step 2 of the IRISA-TBLASTN algorithm presented in section 2. Here, measurements report the time for computing step 1 and 2, that is the dynamic indexing of the protein set, together with the computation of the ungapped extensions. The genome indexing is supposed to have been previously done and stored on disk as it is done for the BLAST family programs. It is obvious that now step 3 is the critical section which will greatly determine the execution time of the overall process.

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Annex 1: Ungapped Alignment Procedure Code

Original code

```

int ComputeDistance(ff,rq)
    FILE *ff;
    REQUEST *rq;
{
    int i,i1,i2,j,k,r,score,maxi,idx_nt;
    char s[256];
    REC_OUT DT[SIZE_BUF];

    r=0;
    for (j=0; j<rq->Qry->nb; j++)
    {
        fseeko(ff,rq->offset,SEEK_SET);
        for (i1=0; i1<rq->nbelt; i1=i1+SIZE_BUF)
        {
            i2 = SIZE_BUF;
            if (i1+i2 >= rq->nbelt) i2 = rq->nbelt - i1;
            fread (DT,i2*sizeof(REC_OUT),1,ff);
            for (i=0; i<i2; i++)
            {
                score = 0;
                maxi = 0;
                for (k=0; k<SIZE_BLOCK; k++)
                {
                    score = score + MATRIX[CODE_AA[rq->Qry->seq[j][k]][CODE_AA[DT[i].seq[k]]];
                    if (score<0) score = 0;
                    else if (maxi<score) maxi = score;
                }
                if (maxi > X1)
                {
                    rq->Res[r].idx_nt = DT[i].idx;
                    rq->Res[r].idx_prot = rq->Qry->idx[j];
                    rq->Res[r].num_seq_prot = rq->Qry->num_seq[j];
                    r++;
                }
            }
        }
    }
    rq->nbRes=r;
    return r;
}

```

Modified code for including the PSC operator

```

int ComputeDistanceFPGA(handle, ff, rq)
    fpga_handle handle;
    FILE *ff;
    REQUEST *rq;
{
    int j,i,k,rc,r,rt;
    int resMissed =0;
    REC_OUT DT[SIZE_BUF];
    int i2,i1;
    rt=0;

    /* initializing and copying Query data into Actor 0's memory area */

    memset(addr[0], 0x1b, SIZE_FPGA*NB_QUERY);
    for (j=0; j<rq->Qry->nb; j++){
        for (k=0; k<SIZE_FPGA; k++){
            *((char *)addr[0]+j*SIZE_FPGA+k) = FPGA_AA[rq->Qry->seq[j][k]];
        }
    }

    fseeko(ff,rq->offset,SEEK_SET);

    for(i1=0 ; i1 < rq->nbelt ; i1 += SIZE_BUF) {

        i2 = SIZE_BUF;
        if (i1+i2 >= rq->nbelt){ i2 = rq->nbelt - i1;}

        fread (DT,sizeof(REC_OUT),i2,ff);

        /* Resizing and copying indexed Genome into Actor 1's memory area */

        adjustMemAllocActor(handle,SIZE_FPGA*i2,1);
        memcpy((void *)addr[1],&DT[0],SIZE_FPGA*i2);

        /* Actors/DMA setup */

        alen[0] = SIZE_FPGA*NB_QUERY;
        alen[1] = SIZE_FPGA*i2;
        alen[2] = mlen[2];
        for (i=0 ; i<ACTS ; i++) {
            rc = fpga_act_request(handle, act_id[i], 0, alen[i], 1, 1);
        }

        /* Actors/DMA start */

        for(i=ACTS-1;i>=0;i--){
            rc = fpga_act_start(handle, act_id[i]);
        }
    }
}

```

```
/* Waiting for completion... */

for (;;) {
    int alldone = 1;
    int sct = fpga_wait(handle, 2000);
    if (sct !=0) {
        int status;
        int rc;
        for (i=0 ; i<ACTS ; i++) {
            rc = fpga_act_get_status(handle, act_id[i], &status);
            printf("Actor %d status: %08x\n", act_id[i], status);
            if ((status & (1<<0))) alldone = 0;
        }
        if (alldone) break;
    }
}

/* copying actor 2's memory data into Results structure */

r=0;
int idfpga0=0;
int idfpga1=0;
while(*(int *)addr[2]+2*r)!=0){

    idfpga0 = *((unsigned char *)addr[2]+8*r+1)*256+*((unsigned char *)addr[2]+8*r)-1;
    idfpga1 = *((unsigned char *)addr[2]+8*r+3)*256+*((unsigned char *)addr[2]+8*r+2)-1;

    rq->Res[rt].idx_nt = *((int *)((unsigned char *)addr[1]+(48*(idfpga1))+44));
    rq->Res[rt].idx_prot = rq->Qry->idx[idfpga0];
    rq->Res[rt].num_seq_prot = rq->Qry->num_seq[idfpga0];
    r++;rt++;
}

memset(addr[2], 0x00, 8*r);

rc = fpga_act_hold(handle, 0);
rc = fpga_act_hold(handle, 1);
rc = fpga_act_hold(handle, 2);

}

rq->nbRes=rt;
return r;
}
```



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